### Influence of iodide ingestion on dietary nitrate metabolism and blood pressure following short-term nitrate supplementation in healthy normotensive adults

## Stephen J. Bailey<sup>a</sup>, Jamie R. Blackwell<sup>a</sup>, Lee J. Wylie<sup>a</sup>, Annabelle Emery<sup>a</sup>, Ellie Taylor<sup>a</sup>, Paul G. Winyard<sup>b</sup> and Andrew M. Jones<sup>a</sup>

<sup>a</sup> Sport and Health Sciences, College of Life and Environmental Sciences and <sup>b</sup> Exeter Medical School, St. Luke's Campus, University of Exeter, Heavitree Road, Exeter, UK.

Address for Correspondence: Stephen J Bailey, Ph.D. E-mail: <u>S.Bailey2@lboro.ac.uk</u> School of Sport, Exercise and Health Sciences Loughborough University Ashby Road Loughborough Leicestershire LE11 3TU Tel: +44 (0) 1509 226433 Fax: +44 (0) 1509 22630

#### ABSTRACT

Uptake of inorganic nitrate  $(NO_3)$  into the salivary circulation is a rate-limiting step for dietary  $NO_3^-$  metabolism in mammals. It has been suggested that salivary  $NO_3^-$  uptake occurs in competition with inorganic iodide ( $\Gamma$ ), but it has yet to be determined whether  $\Gamma$  ingestion compromises NO3<sup>-</sup> metabolism and blood pressure reductions after dietary NO3<sup>-</sup> supplementation. Therefore, this study tested the hypothesis that I<sup>-</sup> ingestion would interfere with NO3<sup>-</sup> metabolism and blunt blood pressure reductions after dietary NO3<sup>-</sup> supplementation. Nine healthy adults (mean  $\pm$  SD, age 20  $\pm$  1 yr, body mass 71  $\pm$  16 kg) reported to the laboratory for initial baseline assessment (CON) and following six day supplementation periods with 140 ml·day<sup>-1</sup> NO<sub>3</sub><sup>-</sup>-rich beetroot juice (8.4 mmol NO<sub>3</sub><sup>-</sup>·day<sup>-1</sup>) and 198 mg potassium gluconate day<sup>-1</sup> (NIT), and 140 ml day<sup>-1</sup> NO<sub>3</sub>-rich beetroot juice and 450  $\mu$ g potassium iodide day<sup>-1</sup> (NIT + I) in a randomized, cross-over experiment. Salivary [I<sup>-</sup> ] was higher in NIT + I compared to CON and NIT (P < 0.05). Salivary and plasma [NO<sub>3</sub><sup>-</sup>] and  $[NO_2^-]$  were higher in NIT and NIT + I compared to CON (P<0.05). Plasma  $[NO_3^-]$  was higher  $(474 \pm 127 \text{ vs. } 438 \pm 117 \text{ } \mu\text{M})$ , salivary  $[NO_3^-] + [NO_2^-]$  was lower  $(8582 \pm 3450 \text{ } \text{ vs.}$  $10487 \pm 2382 \ \mu\text{M}$ ) and the salivary-plasma [NO<sub>3</sub><sup>-</sup>] ratio was lower (14 ± 6 vs. 20 ± 6  $\mu$ M), indicative of a lower salivary  $NO_3^-$  uptake, in NIT + I compared to NIT (P<0.05). Plasma and salivary  $[NO_2]$  were not different between NIT and NIT + I (P>0.05). Systolic blood pressure was lower than CON ( $112 \pm 13 \text{ mmHg}$ ) in NIT ( $106 \pm 13 \text{ mmHg}$ ) and NIT + I ( $106 \pm 13 \text{ mmHg}$ )  $\pm$  11 mmHg; P<0.05), with no differences between NIT and NIT + I (P>0.05). In conclusion, co-ingesting NO<sub>3</sub><sup>-</sup> and I<sup>-</sup> perturbed salivary NO<sub>3</sub><sup>-</sup> uptake, but the increase in salivary and plasma [NO<sub>2</sub><sup>-</sup>] and the lowering of blood pressure were similar, compared to NO<sub>3</sub><sup>-</sup> ingestion alone. Therefore, increased dietary I intake, which is imposed in several countries worldwide as an initiative to offset hypothyroidism, does not appear to compromise the blood pressure reduction afforded by increased dietary NO<sub>3</sub><sup>-</sup> intake.

Key Words: Entero-salivary circulation; nitrite; nitric oxide; vascular health; nutrition

#### HIGHLIGHTS

- Inorganic iodide interfered with salivary nitrate uptake
- The change in plasma [nitrite] was not adversely impacted by iodide supplementation
- Nitrate supplementation lowered blood pressure with or without iodide co-ingestion
- Iodide supplementation did not compromise the hypotensive effects of nitrate supplementation

#### **1. INTRODUCTION**

The gaseous molecule, nitric oxide (NO), regulates an array of physiological processes, but is perhaps best known for its vasodilatory and cardioprotective properties [1,2]. It has been demonstrated that NO can be generated through the  $O_2$ -independent reduction of nitrite (NO<sub>2</sub><sup>-</sup>) to compliment  $O_2$ -dependent NO generation through the NO synthases [3-5]. The circulating plasma [NO<sub>2</sub><sup>-</sup>] can be increased through dietary supplementation with inorganic nitrate (NO<sub>3</sub><sup>-</sup>) in association with a reduction in blood pressure and arterial stiffness [6-8], important predictors of future cardiovascular events [9,10]. In addition, NO<sub>3</sub><sup>-</sup> supplementation can improve vascular function in healthy older adults [11] and some patient populations including peripheral artery disease [12] and heart failure [13] patients. Therefore, increasing dietary NO<sub>3</sub><sup>-</sup> intake appears to confer cardioprotective effects and might hold promise as a nutritional intervention to lower the epidemiological and economic burden of cardiovascular diseases on society [14].

Approximately 25% of  $NO_3^-$  consumed through the diet passes into the entero-salivary circulation where it is delivered to the mouth for second-pass metabolism [15]. Upon arrival at the oral cavity, microflora on the tongue reduce  $NO_3^-$  to nitrite ( $NO_2^-$ ) [15-19]. After ingestion, NO<sub>2</sub><sup>-</sup> is chemically reduced to nitric oxide (NO) and other reactive nitrogen intermediates in the acidic environment of the stomach [20,21], but it is well documented that the circulating plasma  $[NO_2^-]$  is also increased after increased  $NO_3^-$  intake [6-8,18]. This circulating plasma  $NO_2^-$  can then exert a beneficial effect on the vasculature either through direct NO<sub>2</sub><sup>-</sup> action [22,23] or through its subsequent reduction to nitric oxide (NO) via numerous  $NO_2^-$  reductatses [24]. While mammalian tissue is capable of reducing  $NO_3^-$  to  $NO_2^{-1}$  [25], the rate limiting steps for the chemical  $NO_3^{-1}$  reduction in mammals are  $NO_3^{-1}$ transport into the entero-salivary circulation and NO<sub>3</sub><sup>-</sup> reduction to NO<sub>2</sub><sup>-</sup> by the oral microflora [26]. Importantly, the anions perchlorate  $(CIO_4)$ , thiocyanate (SCN), iodide (I)and NO<sub>3</sub><sup>-</sup> share a common transporter for uptake into the salivary glands, with the order of affinity for salivary uptake being  $CIO_4^- > SCN^- > I^- > NO_3^-$  [27]. Although  $CIO_4^-$  has the highest affinity for salivary uptake of the aforementioned anions [27], environmental exposure to  $CIO_4^-$  is limited [28,29]. Consequently, the competition between SCN<sup>-</sup>, I<sup>-</sup> and  $NO_3^-$  is more likely to be pertinent for  $NO_3^-$  transfer into the entero-salivary circulation [28,29] and, subsequently, the stepwise reduction of  $NO_3^-$  to  $NO_2^-$  and then NO.

Vegetable consumption provides the largest source of dietary  $NO_3^-$  intake [30]. A diet rich in fruit and vegetables is accompanied by a lower cardiovascular disease morbidity and incidence of future cardiovascular events [31,32]. This evidence has prompted government initiatives in numerous countries to increase fruit and vegetable consumption, including the Dietary Approaches to Stop Hypertension (DASH) diet in the United States of America [33], the 5-A-Day diet in the United Kingdom [34] and variations of this latter diet in countries within the European Union [35]. However, it has been suggested that the cardioprotective effect of  $NO_3^-$ -rich vegetables outweighs that afforded by vegetables low in  $NO_3^-$  [36-38]. Therefore, while adhering to global initiatives to increase fruit and vegetable consumption affords cardioprotection, incorporating a greater proportion of  $NO_3^-$ -rich vegetables into the diet appears to hold greater potential as a dietary approach to increase cardioprotection, and is currently being actively encouraged [36,39,40].

We have recently reported that cigarette smoking [41], which increased salivary and plasma [SCN<sup>-</sup>], perturbed aspects of dietary NO<sub>3</sub><sup>-</sup> metabolism, and thwarted the lowering of blood pressure, after dietary NO<sub>3</sub><sup>-</sup> supplementation. However, while SCN<sup>-</sup> can interfere with dietary NO<sub>3</sub><sup>-</sup> metabolism and its lowering of blood pressure, it has yet to be determined whether this is also the case when  $NO_3^-$  and  $I^-$  are co-ingested. Although  $I^-$  has the potential to interfere with salivary  $NO_3^-$  uptake (27), and therefore its subsequent reduction to  $NO_2^-$  in the oral cavity, this potential perturbation to dietary NO<sub>3</sub><sup>-</sup> metabolism might be offset by a compensatory increase in NO generation in the stomach. Indeed, nitrous acid (HNO<sub>2</sub>), which is formed from the deprotonation of ingested salivary NO2<sup>-</sup> in the stomach [42], can react with I<sup>-</sup> to form NO at an acidic pH [43]. Accordingly, further research is required to assess the extent to which dietary I<sup>-</sup> enrichment impacts dietary NO<sub>3</sub><sup>-</sup> metabolism and associated vascular health benefits. This is important because I<sup>-</sup> is present in numerous food sources, with seafood and dairy products, particularly seaweed, white fish, yogurt and milk, abundant in I<sup>-</sup> [44]. Moreover, in excess of 100 countries fortify their salt with I<sup>-</sup>, or mandate the use of iodised salt for the production of products such as bread, in an effort to alleviate the prevalence of hypothyroidism [45,46]. These government initiatives have been successful at increasing I<sup>-</sup> exposure [46], but it is unclear if this might be to the detriment of dietary NO<sub>3</sub><sup>-</sup> metabolism and its beneficial effect on blood pressure.

The purpose of this study was to examine the effect of co-supplementation with  $NO_3^-$  and I<sup>-</sup>, compared to  $NO_3^-$  supplementation alone, on dietary  $NO_3^-$  metabolism and blood pressure.

We hypothesised that  $NO_3^-$  supplementation would increase salivary and plasma  $[NO_3^-]$  and  $[NO_2^-]$  and lower blood pressure, but that concurrent  $NO_3^-$  and  $I^-$  supplementation would attenuate: 1) salivary  $NO_3^-$  uptake, 2) the increase in circulating plasma  $[NO_2^-]$  and 3) the lowering of blood pressure compared to  $NO_3^-$  supplementation alone.

#### 2. METHODS

#### 2.1 Subject characteristics

We recruited nine healthy non-smoking adults (4 males, mean  $\pm$  SD, age 20  $\pm$  1 yr, body mass 71  $\pm$  16 kg, height 1.72 m) to participate in this study. All procedures employed in this study were approved by the Institutional Research Ethics Committee and subjects gave their written informed consent to participate after the experimental procedures, associated risks, and potential benefits of participation had been explained. Subjects were instructed to arrive at each laboratory testing session in a rested and fully hydrated state, at least 3 h postprandial. Since the reduction of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup> in the oral cavity is abolished by antibacterial mouthwash [47], subjects were required to refrain from mouthwash use for the duration of the study. Each subject was also asked to avoid consumption of NO<sub>3</sub><sup>-</sup>-rich and SCN<sup>-</sup>-rich foods for the duration of the study, and from caffeine and alcohol ingestion 6 and 24 h before each test, respectively. Subjects were instructed to maintain their habitual exercise pattern for the duration of the study. All tests were performed at the same time of day ( $\pm$  2 hours).

#### **2.2 Supplementation Procedures**

All subjects were required to report to the laboratory on three occasions over a 3-4 week period. Subjects did not undergo dietary supplementation prior to their first visit to the laboratory (the control condition; CON). Subjects were asked to record their food and beverage consumption on the day of the CON test and for the 5 days preceding this test and to replicate this prior to the subsequent trials. After completing the CON trial, subjects were randomly assigned to receive six days of supplementation with NO<sub>3</sub><sup>-</sup>-rich beetroot juice and a placebo of potassium gluconate (NIT), or NO<sub>3</sub><sup>-</sup>-rich beetroot juice and potassium iodide (NIT + I) as part of a double-blind, cross-over experimental design. Over the first five days of supplementation, subjects ingested 4.2 mmol NO<sub>3</sub><sup>-</sup> (as 70 ml of concentrated beetroot juice) and a single 99 mg potassium gluconate capsule in the NIT condition, and 4.2 mmol NO<sub>3</sub><sup>-</sup> and a single 225  $\mu$ g potassium iodide capsule in the NIT + I condition, in the morning and

evening. Subjects ingested 140 ml of beetroot juice and two capsules two hours before reporting to the laboratory on day six of NIT and NIT + I supplementation. This was selected to coincide with the peak plasma  $[NO_2^-]$  attained following ingestion of 8.4 mmol  $NO_3^-$  [8]. A 7-10 day washout separated the supplementation periods. Potassium gluconate and potassium iodide capsules were provided by NOW Sports Nutrition (NOW Foods, Bloomingdale, IL, USA) and  $NO_3^-$ -rich beetroot juice was purchased from James White Drinks (Beet It; James White Drinks, Ipswich, UK).

#### **2.3 Measurements**

#### 2.3.1 Blood Pressure

Subjects were required to rest supine for 10 min in an isolated room. Thereafter, blood pressure of the brachial artery was measured whilst the subject was supine using an automated sphygmomanometer (Dinamap Pro, GE Medical Systems, Tampa, USA). Five measurements were taken and the mean of the measurements 2-5 was used for analysis.

#### 2.3.2 Blood and saliva collection

Venous blood samples were drawn into 6 mL lithium-heparin tubes (Sarstedt, Leicester, UK). Samples were centrifuged at 4,000 rpm and 4°C for 10 min, within 1-min of collection. Plasma was subsequently extracted and immediately frozen at -80°C for later analysis of  $[NO_3^{-1}]$  and  $[NO_2^{-1}]$ . Unstimulated saliva samples (~ 5 mL) were collected into 30 mL universal containers and 1.5 mL aliquots were frozen at -80°C for later analysis of  $[I^{-1}]$ ,  $[NO_3^{-1}]$  and  $[NO_2^{-1}]$ .

#### 2.4 Data analysis procedures

#### 2.4.1 [*I*<sup>-</sup>] determination

After thawing at room temperature, saliva samples were centrifuged at 1600 g for 10 min, and the supernatant was removed for subsequent analysis. 1 mL of saliva supernatant, 1 mL of deionised water and 40  $\mu$ L of ionic strength adjustor were added to a 30 mL universal container for assessment of salivary [I<sup>-</sup>]. Salivary [I<sup>-</sup>] was determined by plotting the mV signal derived from an iodide-selective electrode (PerfectION<sup>TM</sup>, Mettler-Toledo AG, Switzerland) against a calibration plot of I<sup>-</sup> standards. All measures were completed at 22°C.

#### 2.4.2 $[NO_3^-]$ and $[NO_2^-]$ determination

All glassware, utensils, and surfaces were rinsed with deionized water to remove residual NO intermediates prior to  $[NO_2^-]$  and  $[NO_3^-]$  analysis. Plasma samples were deproteinized using zinc sulfate/sodium hydroxide precipitation prior to determination of [NO<sub>3</sub><sup>-</sup>]. Firstly, 500 μL of 0.18 N NaOH was added to 100 µL of sample followed by 5 min incubation at room temperature. Subsequently, samples were treated with 300 µL aqueous ZnSO4 (5% w/v) and vortexed for 30 seconds before undergoing an additional 10 min incubation period at room temperature. Samples were then centrifuged at 4,000 rpm for 5 min, and the supernatant was removed for subsequent analysis. The [NO<sub>3</sub><sup>-</sup>] of the deproteinized plasma sample was determined by its reduction to NO in the presence of 0.8 % (w/v) VCl<sub>3</sub> in 1M HCl within an air-tight purging vessel. Plasma samples were introduced to the vessel via 50 uL injections into the septum at the top of the vessel. The spectral emission of electronically excited nitrogen dioxide, derived from the reaction of NO with ozone, was detected by a thermoelectrically cooled, red-sensitive photomultiplier tube housed in a Sievers gas-phase chemiluminescence nitric oxide analyzer (Sievers NOA 280i. Analytix Ltd, Durham, UK). The  $[NO_3]$  was determined by plotting signal (mV) area against a calibration plot of sodium nitrate standards. The  $[NO_2]$  of the undiluted (non-deproteinized) plasma was determined by its reduction to NO in the presence of glacial acetic acid and aqueous NaI (4% w/v) from sodium nitrite standards. 100 uL injections were used for plasma [NO<sub>2</sub><sup>-</sup>] determination. After thawing at room temperature, saliva samples were centrifuged for 10 min at 14000 rpm and the supernatant was removed for subsequent analysis. The supernatant was diluted 100 fold with deionized water and  $[NO_3^-]$  and  $[NO_2^-]$  were determined from 50 uL injections using the same reagents describe above for the plasma analyses.

#### 2.4.3 [TSH] and $[T_4]$ determination

Plasma thyroid stimulating hormone ([TSH]) and thyroxine ([T<sub>4</sub>]) concentrations were assessed in duplicate using ELISA kits purchased from DRG Diagnostics (DRG Instruments GmbH, Germany).

#### **2.5 Statistics**

A one-way repeated-measures ANOVA was employed to determine the effects of the different dietary interventions (CON, NIT and NIT + I) on the relevant outcome variables. Where the analysis revealed a significant main effect for supplement, Fishers Least Significant Difference tests were employed to determine the origin of such effects. All data

are presented as mean  $\pm$  SD unless otherwise indicated. Statistical significance was accepted when *P*<0.05.

#### **3. RESULTS**

The  $NO_3^-$  and I<sup>-</sup> supplements administered in this study were well tolerated by all subjects with no negative side effects reported. Subjects consumed all doses of the supplements for each experimental condition and self-reported that their diet was consistent across all the dietary interventions.

#### 3.1 *Salivary* [*I*<sup>-</sup>]

Salivary [I<sup>-</sup>] responses in the CON, NIT and NIT + I conditions are illustrated in figure 1. There was a significant main effect for supplement on salivary [I<sup>-</sup>] (P<0.05), with salivary [I<sup>-</sup>] being lower in the NIT condition (384 ± 245 µg·L<sup>-1</sup>) compared to the CON (487 ± 305 µg·L<sup>-1</sup>) and NIT + I (794 ± 269 µg·L<sup>-1</sup>) conditions (P<0.05), and higher than both the NIT and CON conditions in the NIT + I condition (P<0.05).

#### 3.2 Plasma [TSH] and [T4]

Plasma [TSH] and [T<sub>4</sub>] responses in the CON, NIT and NIT + I conditions are presented in table 1. There were no difference in [TSH] or [T<sub>4</sub>] between the CON, NIT and NIT + I conditions (P>0.05).

#### 3.3 Salivary $[NO_3^-]$ , $[NO_2^-]$ and $[NO_x]$

The changes in salivary  $[NO_3^-]$ ,  $[NO_2^-]$  and [NOx] in the NIT and NIT + I conditions relative to CON are illustrated in figure 2. There were significantly main effects for supplement for salivary  $[NO_3^-]$ ,  $[NO_2^-]$  and [NOx] (P<0.05). Salivary  $[NO_3^-]$ ,  $[NO_2^-]$  and [NOx] were greater in NIT and NIT + I compared to CON (P<0.05). There were no differences between NIT (2168 ± 1302 µM) and NIT + I (1952 ± 1316 µM) for salivary  $[NO_2^-]$  (P>0.05), there was a trend (P=0.07) for a higher salivary  $[NO_3^-]$  in NIT (8318 ± 2399 µM) compared to NIT + I (6630 ± 3516 µM) and salivary [NOx] was higher in NIT (10487 ± 2382 µM) compared to NIT + I (8582 ± 3450 µM; P<0.05).

#### 3.4 Plasma $[NO_3^-]$ and $[NO_2^-]$

The changes in plasma  $[NO_3^-]$  and  $[NO_2^-]$  in the NIT and NIT + I conditions relative to CON are illustrated in figure 3. There were significantly main effects for supplement for plasma  $[NO_3^-]$  and  $[NO_2^-]$  (*P*<0.05). Plasma  $[NO_3^-]$  and  $[NO_2^-]$  were greater in NIT and NIT + I compared to CON (*P*<0.05). Plasma  $[NO_3^-]$  was higher in NIT + I (474 ± 127 µM) compared to NIT (438 ± 117 µM; *P*<0.05), while plasma  $[NO_2^-]$  was not different between NIT (404 ± 142 nM) and NIT + I (407 ± 145 nM; *P*>0.05).

#### 3.5 Salivary-plasma $[NO_3^-]$ and $[NO_2^-]$ ratios

The salivary-plasma [NO<sub>3</sub><sup>-</sup>] ratio was lower in the NIT + I ( $14 \pm 6 \mu M$ ) compared to NIT (20  $\pm 6 \mu M$ ; *P*<0.05; Figure 4). The salivary-plasma [NO<sub>2</sub><sup>-</sup>] ratio was not different between the NIT + I (5184  $\pm 3481 \mu M$ ) and NIT conditions (5552  $\pm 3145 \mu M$ ; *P*>0.05; Figure 4).

#### 3.6 Blood pressure responses

SBP, DBP and MAP responses in the CON, NIT and NIT + I conditions are presented in table 2. There were significantly main effects for supplement for SBP, DBP and MAP (P<0.05). SBP and MAP were lower than CON in both NIT and NIT + I (P<0.05), while DBP was only lower than CON in NIT (P<0.05; table 2). There were no differences in any of the blood pressure variables between NIT and NIT + I (P>0.05). The change in plasma [NO<sub>2</sub><sup>-</sup>] tended to be negatively correlated with the change in SBP between the CON and NIT groups (r =-0.65; P=0.06).

#### 4. DISCUSSION

The main original findings from this study were: 1) salivary  $NO_3^-$  uptake was lower after coingesting  $NO_3^-$  and I<sup>-</sup> compared to  $NO_3^-$  ingested alone (as reflected by a higher plasma  $[NO_3^-]$ ], lower salivary  $[NO_3^-] + [NO_2^-]$  and a lower salivary-plasma  $[NO_3^-]$  ratio in the former compared to the latter), 2) circulating plasma  $[NO_2^-]$  was increased and blood pressure was lowered to a similar extent after ingesting  $NO_3^-$  with and without I<sup>-</sup> co-ingestion. Therefore, while increased dietary I<sup>-</sup> has the potential to impede salivary  $NO_3^-$  uptake, this did not blunt the increase in plasma  $[NO_2^-]$  and the lowering of blood pressure after short-term dietary  $NO_3^-$  supplementation. These findings suggest that conforming to global initiatives to increase dietary I<sup>-</sup> intake is unlikely to compromise the lowering of blood pressure after dietary  $NO_3^-$  supplementation, at least at the  $NO_3^-$  dose administered in this study.

Salivary [I<sup>-</sup>] was increased above the un-supplemented control by 63% in the NIT + I condition in this study. Interestingly, ingestion of NO<sub>3</sub><sup>-</sup> alone lowered salivary [I<sup>-</sup>] to 79% of that observed in the un-supplemented control condition. This observation parallels our previous finding of a lower salivary [SCN<sup>-</sup>] after NO<sub>3</sub>-ingestion [41]. Collectively, these findings substantiate notion that SCN<sup>-</sup>, I<sup>-</sup> and NO<sub>3</sub><sup>-</sup> compete for a common salivary transporter [27], with recent evidence pointing to sialin as the key transport protein for salivary NO<sub>3</sub><sup>-</sup> uptake [48]. Importantly, although salivary [I<sup>-</sup>] was lower in NIT compared to the CON trial, TSH and T4 were not different between the CON, NIT and NIT + I trials. These findings corroborate a previous study which reported no differences in thyroid hormones following short-term NO<sub>3</sub><sup>-</sup> supplementation [49]. Therefore, while short-term supplementation with 8 mmol  $NO_3^-$  (a dose that could be achieved through ingestion 200-300 g of NO<sub>3</sub><sup>-</sup>rich vegetables [36-38]) can lower salivary [I<sup>-</sup>], this was not sufficient to perturb thyroid gland function. However, chronic supplementation with  $> 8 \text{ mmol NO}_3^-$  daily is not recommended as blood pressure is not lowered further and exercise capacity is not increased more [8], but there could be a risk of compromising thyroid gland function [50]. In particular, individuals at risk from, or being treated for, hypothyroidism are not recommended to consume excessive  $NO_3^-$ . Further research is required to assess the effects of dietary NO<sub>3</sub><sup>-</sup> supplementation on thyroid function to optimise supplementation guidelines for different populations.

The short-term dietary  $NO_3^{-1}$  supplementation regime employed in this study increased salivary and plasma  $[NO_3^{-1}]$  and  $[NO_2^{-1}]$ , consistent with several previous reports [6,18, 41,47, 51]. However, when the same absolute  $NO_3^{-1}$  dose was co-ingested with  $\Gamma$ , plasma  $[NO_3^{-1}]$  was 8% higher, salivary  $[NO_3^{-1}]$  was 20% lower (*P*=0.07) and the salivary  $[NO_3^{-1}] + [NO_2^{-1}]$  was 18% lower. The increase in plasma  $[NO_3^{-1}]$  and lowering of salivary  $[NO_3^{-1}] + [NO_2^{-1}]$  in the NIT + I condition is indicative of an antagonistic effect of I<sup>-</sup> on salivary  $NO_3^{-}$  uptake, in line with previous observations [27]. Moreover, the salivary-plasma  $[NO_3^{-1}]$  ratio was 30% lower in the NIT + I condition compared to the NIT condition. Therefore, for a given plasma  $[NO_3^{-1}]$ , salivary  $NO_3^{-1}$  uptake was ~ 30% lower when  $NO_3^{-1}$  and I<sup>-</sup> were co-ingested compared to  $NO_3^{-1}$  ingestion alone. Taken together these findings indicate that, compared to  $NO_3^{-1}$  indicate that, compared to  $NO_3^{-1}$  indicate that is shown in the NO\_3^{-1} and I<sup>-</sup> were co-ingested compared to  $NO_3^{-1}$  ingested independently of I<sup>-</sup>, combined dietary supplementation with  $NO_3^{-1}$  and I<sup>-</sup>

compromised salivary NO<sub>3</sub><sup>-</sup> uptake, which is a key rate-limiting step for dietary NO<sub>3</sub><sup>-</sup> supplementation in mammals [26]. However, despite a lower salivary NO<sub>3</sub><sup>-</sup> uptake, salivary [NO<sub>2</sub><sup>-</sup>] was not significantly different between the NIT + I and NIT trials. This observation is similar to our recent study in smokers who exhibited a smaller increase in salivary [NO<sub>3</sub><sup>-</sup>], but a similar increase in salivary [NO<sub>2</sub><sup>-</sup>], compared to non-smokers after ingesting the same NO<sub>3</sub><sup>-</sup> dose (41). In line with the salivary [NO<sub>2</sub><sup>-</sup>] results, and in spite of the potential for increased NO generation in the stomach with NO<sub>3</sub><sup>-</sup> and I<sup>-</sup> co-ingestion [42,43], plasma [NO<sub>2</sub><sup>-</sup>] was not different between the NIT + I and NIT trials. Therefore, while salivary NO<sub>3</sub><sup>-</sup> uptake was compromised in NIT + I compared to NIT, salivary and plasma [NO<sub>2</sub><sup>-</sup>] were similarly increased in NIT and NIT + I. Nevertheless, we cannot exclude the possibility that I<sup>-</sup> ingestion might impact salivary NO<sub>3</sub><sup>-</sup> uptake, and its subsequent metabolism, at lower NO<sub>3</sub><sup>-</sup> doses.

Consistent with numerous previous reports [6,8,37,38,41], SBP was lowered by 6 mmHg in the NIT trial in this study compared to CON. Likewise, a 6 mmHg lowering in SBP was observed in the NIT + I group compared to CON. In keeping with previous reports [8,52], the change in SBP following dietary NO<sub>3</sub><sup>-</sup> supplementation tended (P=0.06) to be negatively correlated with the change in plasma [NO<sub>2</sub><sup>-</sup>]. The association between the increase in circulating plasma [NO<sub>2</sub><sup>-</sup>] and lower SBP after NO<sub>3</sub><sup>-</sup> supplementation may be the result of vasodilation evoked from a direct of NO<sub>2</sub><sup>-</sup> on the vasculature [22,23] and/or NO<sub>2</sub><sup>-</sup> reduction to NO [24]. These changes might be mediated by increased plasma [cGMP; 52], or altered renal physiology [53]. Therefore, the similar reduction in SBP in the NIT and NIT + I groups is likely to be a function of a similar increase in plasma [NO<sub>2</sub><sup>-</sup>] in these groups. These findings suggest that increasing dietary NO<sub>3</sub><sup>-</sup> intake concomitant with increased dietary I<sup>-</sup> is unlikely to compromise the increase in circulating plasma [NO<sub>2</sub><sup>-</sup>] and associated lowering of blood pressure.

The impetus for increased dietary I<sup>-</sup> originated from Switzerland and the USA, where salt iodisation was mandated in the 1920s in an attempt to alleviate hypothyroidism [44]. These initiatives have proved effective at increasing I<sup>-</sup> exposure and lowering the incidence of hypothyroidism [46]. Consequently, numerous countries have imposed salt iodisation programmes or the manufacture of certain food products with iodised salt [45,46]. The recommended adequate daily intake for I<sup>-</sup> is 150 µg for adult males and females, and 200 µg during pregnancy and lactation [54]. These doses could readily be achieved through a diet rich in seafood and dairy products [44], but the I<sup>-</sup> dose administered in this study (450  $\mu$ g·day<sup>-1</sup>) exceeded the upper range of average daily I<sup>-</sup> in the US where iodised salt is mandated [55]. However, despite administering a large I<sup>-</sup> dose in this study (i.e., an I<sup>-</sup> dose that is not likely to be achieved by most individuals through the diet), the increase in plasma [NO<sub>2</sub><sup>-</sup>] and the lowering of blood pressure after NO<sub>3</sub><sup>-</sup> supplementation were not different with or without co-ingestion of I<sup>-</sup>. Therefore, our data imply that conforming to global initiatives to increase dietary I<sup>-</sup> intake is unlikely to impede the lowering of blood pressure after supplementation of NO<sub>3</sub><sup>-</sup>-rich vegetables such as spinach, lettuce or rocket [36-38].

#### **5. CONCLUSION**

In conclusion, concurrent  $\Gamma$  and NO<sub>3</sub><sup>-</sup> ingestion lowered salivary NO<sub>3</sub><sup>-</sup> uptake, as indicated by a higher plasma [NO<sub>3</sub><sup>-</sup>], a lower salivary [NO<sub>3</sub><sup>-</sup>] + [NO<sub>2</sub><sup>-</sup>], and a lower salivary-plasma [NO<sub>3</sub><sup>-</sup>] ratio, compared to the same dose of NO<sub>3</sub><sup>-</sup> consumed without I<sup>-</sup> co-ingestion. However, in spite of a lower salivary NO<sub>3</sub><sup>-</sup> uptake with I<sup>-</sup> and NO<sub>3</sub><sup>-</sup> co-ingestion, salivary and plasma [NO<sub>2</sub><sup>-</sup>] were elevated and blood pressure was lowered to a similar extent when NO<sub>3</sub><sup>-</sup> was consumed with or without I<sup>-</sup> co-ingestion. These observations are important because they suggest that increasing dietary I<sup>-</sup>, which is imposed in several countries worldwide through salt iodisation programmes, does not interfere with the lowering of blood pressure after NO<sub>3</sub><sup>-</sup> supplementation. Therefore, our results suggest that conforming to global initiatives to increase dietary I<sup>-</sup> is unlikely to compromise the ability of a NO<sub>3</sub><sup>-</sup> enriched diet to improve blood pressure.

#### Acknowledgment

The authors are grateful to NOW Sports Nutrition for providing the potassium gluconate and potassium iodide capsules that were used in this study. We received no funding from NOW Sports Nutrition for this work.

#### **REFERENCES**

[1] Loscalzo, J.; Welch, G. Nitric oxide and its role in the cardiovascular system. Prog Cardiovasc Dis. 38:87-104; 1995.

[2] Lei, J.; Vodovotz, Y.; Tzeng, E.; Billiar, T. R. Nitric oxide, a protective molecule in the cardiovascular system. Nitric Oxide. 35:175-185; 2013.

[3] Bryan, N. S.; Calvert, J. W.; Gundewar, S.; Lefer, D. J. Dietary nitrite restores NO homeostasis and is cardioprotective in endothelial nitric oxide synthase-deficient mice. Free Radic Biol Med. 45:468-474; 2008.

[4] Carlström, M.; Larsen, F. J.; Nyström, T.; Hezel, M.; Borniquel, S.; Weitzberg, E.; Lundberg, J. O. Dietary inorganic nitrate reverses features of metabolic syndrome in endothelial nitric oxide synthase-deficient mice. Proc Natl Acad Sci U S A. 107:17716-17720; 2010.

[5] Ferguson, S. K.; Glean, A. A.; Holdsworth, C. T.; Wright, J. L.; Fees, A. J.; Colburn, T. D.; Stabler, T.; Allen, J. D.; Jones, A. M.; Musch, T. I.; Poole, D. C. Skeletal Muscle Vascular Control During Exercise: Impact of Nitrite Infusion During Nitric Oxide Synthase Inhibition in Healthy Rats. J Cardiovasc Pharmacol Ther. 21:201-208; 2016.

[6] Bahra, M.; Kapil, V.; Pearl, V.; Ghosh, S.; Ahluwalia, A. Inorganic nitrate ingestion improves vascular compliance but does not alter flow-mediated dilatation in healthy volunteers. Nitric Oxide. 26:197-202; 2012.

[7] Webb, A. J.; Patel, N.; Loukogeorgakis, S.; Okorie, M.; Aboud, Z.; Misra, S.; Rashid, R.; Miall, P.; Deanfield, J.; Benjamin, N.; MacAllister, R.; Hobbs, A. J.; Ahluwalia, A. Acute blood pressure lowering, vasoprotective, and antiplatelet properties of dietary nitrate via bioconversion to nitrite. Hypertension. 51:784-790; 2008.

[8] Wylie, L. J.; Kelly, J.; Bailey, S. J.; Blackwell, J. R.; Skiba, P. F.; Winyard, P. G.; Jeukendrup, A. E.; Vanhatalo, A.; Jones, A. M. Beetroot juice and exercise: pharmacodynamic and dose-response relationships. J Appl Physiol. 115:325-336; 2013.

[9] Sesso, H. D.; Stampfer, M. J.; Rosner, B.; Hennekens, C. H.; Gaziano, J. M.; Manson, J. E.; Glynn, R. J. Systolic and diastolic blood pressure, pulse pressure, and mean arterial pressure as predictors of cardiovascular disease risk in Men. Hypertension. 36:801-807; 2000.

[10] Maroules, C. D.; Khera, A.; Ayers, C.; Goel, A.; Peshock, R. M.; Abbara, S.; King, K. S. Cardiovascular outcome associations among cardiovascular magnetic resonance measures of arterial stiffness: the Dallas heart study. J Cardiovasc Magn Reson. 16:33; 2014.

[11] Rammos C.; Hendgen-Cotta, U. B.; Sobierajski, J.; Bernard, A.; Kelm, M.; Rassaf, T. Dietary nitrate reverses vascular dysfunction in older adults with moderately increased cardiovascular risk. J Am Coll Cardiol. 63:1584-1585; 2014.

[12] Kenjale, A.A.; Ham, K. L.; Stabler, T.; Robbins, J. L.; Johnson, J. L.; Vanbruggen, M.; Privette, G.; Yim, E.; Kraus, W. E.; Allen, J. D. Dietary nitrate supplementation enhances exercise performance in peripheral arterial disease. J Appl Physiol. 110:1582-1591; 2011.

[13] Zamani, P.; Rawat, D.; Shiva-Kumar, P.; Geraci, S.; Bhuva, R.; Konda, P.; Doulias, P. T.; Ischiropoulos, H.; Townsend, R. R.; Margulies, K. B.; Cappola, T. P.; Poole, D. C.; Chirinos, J. A. Effect of inorganic nitrate on exercise capacity in heart failure with preserved ejection fraction. Circulation. 131:371-80; 2015

[14] Laslett, L. J.; Alagona, P. Jr; Clark, B. A. 3<sup>rd</sup>; Drozda, J. P. Jr; Saldivar, F.; Wilson, S. R.; Poe, C.; Hart, M. The worldwide environment of cardiovascular disease: prevalence, diagnosis, therapy, and policy issues: a report from the American College of Cardiology. J Am Coll Cardiol. 60:S1-S49; 2012.

[15] Spiegelhalder. B.; Eisenbrand, G.; Preussmann, R. Influence of dietary nitrate on nitrite content of human saliva: possible relevance to in vivo formation of N-nitroso compounds. Food Cosmet Toxicol. 14:545-548; 1976.

[16] Tannenbaum, S. R.; Weisman, M.; Fett, D. The effect of nitrate intake on nitrite formation in human saliva. Food Cosmet Toxicol. 14:549-552; 1976.

[17] Duncan, C.; Dougall, H.; Johnston, P.; Green, S.; Brogan, R.; Leifert, C.; Smith, L.; Golden, M.; Benjamin, N. Chemical generation of nitric oxide in the mouth from the enterosalivary circulation of dietary nitrate. Nat Med. 1:546-551; 1995.

[18] Lundberg, J. O.; Govoni, M. Inorganic nitrate is a possible source for systemic generation of nitric oxide. Free Radic Biol Med. 37:395-400; 2004.

[19] Lundberg, J. O.; Weitzberg, E.; Cole, J. A.; Benjamin, N. Nitrate, bacteria and human health. Nat Rev Microbiol. 2:593-602; 2004.

[20] Benjamin, N.; O'Driscoll, F.; Dougall, H.; Duncan, C.; Smith, L.; Golden, M.; McKenzie, H. Stomach NO synthesis. Nature. 368:502; 1994.

[21] Lundberg, J. O.; Weitzberg, E.; Lundberg, J. M.; Alving, K. Intragastric nitric oxide production in humans: measurements in expelled air. Gut. 35:1543-1546; 1994.

[22] Bryan, N. S.; Fernandez, B. O.; Bauer, S. M.; Garcia-Saura, M. F.; Milsom, A. B.; Rassaf, T.; Maloney, R. E.; Bharti, A.; Rodriguez, J.; Feelisch, M. Nitrite is a signaling molecule and regulator of gene expression in mammalian tissues. Nat Chem Biol. 1:290-297; 2005.

[23] Alzawahra, W. F.; Talukder, M. A,;, Liu, X.; Samouilov, A.; Zweier, J. L. Heme proteins mediate the conversion of nitrite to nitric oxide in the vascular wall. Am J Physiol Heart Circ Physiol. 295:499-508; 2008.

[24] van Faassen, E. E.; Bahrami, S.; Feelisch, M.; Hogg, N.; Kelm, M.; Kim-Shapiro, D. B.; Kozlov, A. V.; Li, H.; Lundberg, J. O.; Mason, R.; Nohl, H.; Rassaf, T.; Samouilov, A.; Slama-Schwok, A.; Shiva, S.; Vanin, A. F.; Weitzberg, E.; Zweier, J.; Gladwin, M. T. Nitrite as regulator of hypoxic signaling in mammalian physiology. Med Res Rev. 29:683-741; 2009.

[25] Jansson, E.A.; Huang, L.; Malkey, R.; Govoni, M.; Nihlén, C.; Olsson, A.; Stensdotter, M.; Petersson, J.; Holm, L.; Weitzberg, E.; Lundberg, J. O. A mammalian functional nitrate reductase that regulates nitrite and nitric oxide homeostasis. Nat Chem Biol. 4:411-417; 2008.

[26] Lundberg, J.O.; Weitzberg, E. NO-synthase independent NO generation in mammals. Biochem Biophys Res Commun. 396:39-45; 2010.

[27] Edwards, D. A.; Fletcher, K.; Rowlands, E. N. Antagonism between perchlorate, iodide, thiocyanate, and nitrate for secretion in human saliva; analogy with the iodide trap of the thyroid. Lancet. 266:498-499; 1954.

[28] Knudsen, N.; Brix, T. H. Genetic and non-iodine-related factors in the aetiology of nodular goitre. Best Pract Res Clin Endocrinol Metab. 28:495-506; 2014.

[29] Lewandowski, T. A,; Peterson, M. K.; Charnley, G. Iodine supplementation and drinking-water perchlorate mitigation. Food Chem Toxicol. 80:261-270; 2015.

[30] Ysart, G.; Miller, P.; Barrett, G.; Farrington, D.; Lawrance, P.; Harrison, N. Dietary exposures to nitrate in the UK. Food Addit Contam. 16:521-532; 1999.

[31] Bazzano. L. A.; He, J.; Ogden, L. G.; Loria, C. M.; Vupputuri, S.; Myers, L.; Whelton, P. K. Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first National Health and Nutrition Examination Survey Epidemiologic Follow-up Study. Am J Clin Nutr. 76:93-99; 2002.

[32] Bazzano, L. A.; Serdula, M. K.; Liu, S. Dietary intake of fruits and vegetables and risk of cardiovascular disease. Curr Atheroscler Rep. 5:492-499; 2003.

[33] Appel, L. J.; Moore, T. J.; Obarzanek, E.; Vollmer, W. M.; Svetkey, L.P.; Sacks, F. M.; Bray, G.A.; Vogt, T. M.; Cutler, J. A.; Windhauser, M.M.; Lin, P.H.; Karanja, N. A clinical trial of the effects of dietary patterns on blood pressure. DASH Collaborative Research Group. N Engl J Med. 336:1117-1124, 1997.

[34] NHS Choices. Why 5 a Day? 2013. Retrieved 25<sup>th</sup> August 2015 from <u>http://www.nhs.uk/Livewell/5ADAY/Pages/5ADAYhome.aspx</u>

[35] EUFIC. Fruit and vegetable consumption in Europe – do Europeans get enough? 2012 Retrieved 25<sup>th</sup> August 2015 from <u>http://www.eufic.org/article/en/expid/Fruit-vegetable-consumption-Europe/</u>

[36] Hord, N. G.; Tang, Y.; Bryan, N. S. Food sources of nitrates and nitrites: the physiologic context for potential health benefits. Am J Clin Nutr. 90:1-10; 2009.

[37] Ashworth, A.; Mitchell, K.; Blackwell, J. R.; Vanhatalo, A.; Jones, A.M. High-nitrate vegetable diet increases plasma nitrate and nitrite concentrations and reduces blood pressure in healthy women. Public Health Nutr. 16:1-10; 2015.

[38] Jonvik, K. L.; Nyakayiru, J.; Pinckaers, P. J.; Senden, J. M.; van Loon, L. J.; Verdijk, L. B. Nitrate-Rich Vegetables Increase Plasma Nitrate and Nitrite Concentrations and Lower Blood Pressure in Healthy Adults. J Nutr. 146:986-993; 2016.

[39] Bryan, N. S.; Ivy, J. L. Inorganic nitrite and nitrate: evidence to support consideration as dietary nutrients. Nutr Res. 35:643-54; 2015.

[40] Omar, S. A.; Webb, A. J.; Lundberg, J. O.; Weitzberg, E. Therapeutic effects of inorganic nitrate and nitrite in cardiovascular and metabolic diseases. J Intern Med. 279:315-36; 2016.

#### [41] Our smoking study

[42] Lundberg, J. O.; Weitzberg, E.; Gladwin, M. T. The nitrate-nitrite-nitric oxide pathway in physiology and therapeutics. Nat Rev Drug Discov. 7:156-67; 2008.

[43] Abeledo, C. A.; Kolthoff, I. M. The reaction between nitrite and iodide and its application to the iodimetric titration of these anions. J. Am. Chem. Soc. 53:2893–2897; 1931.

[44] National Institute of Health: Office of Dietary Supplements. Iodine. 2011. Retrieved 8<sup>th</sup> August 2016 from <u>https://ods.od.nih.gov/factsheets/Iodine-HealthProfessional/</u>

[45] Leung, A. M.; Braverman, L. E.; Pearce, E. N. History of U.S. iodine fortification and supplementation. Nutrients.;4:1740-1746; 2012.

[46] Fiore, E.; Tonacchera, M.; Vitti, P. Influence of iodization programmes on the epidemiology of nodular goitre. Best Pract Res Clin Endocrinol Metab. 28:577-588; 2014.

[47] Govoni, M.; Jansson, E. A.; Weitzberg, E.; Lundberg, J. O. The increase in plasma nitrite after a dietary nitrate load is markedly attenuated by an antibacterial mouthwash. Nitric Oxide. 19:333-337; 2008.

[48] Qin, L.; Liu, X.; Sun, Q.; Fan, Z.; Xia, D.; Ding, G.; Ong, H. L.; Adams, D.; Gahl, W. A.; Zheng, C.; Qi, S.; Jin, L.; Zhang, C.; Gu, L.; He, J.; Deng, D.; Ambudkar, I. S.; Wang, S. Sialin (SLC17A5) functions as a nitrate transporter in the plasma membrane. Proc Natl Acad Sci U S A. 109:13434-13439; 2012.

[49] Larsen, F. J.; Schiffer, T. A.; Ekblom, B.; Mattsson, M. P.; Checa, A.; Wheelock, C. E.; Nyström, T.; Lundberg, J. O.; Weitzberg, E. Dietary nitrate reduces resting metabolic rate: a randomized, crossover study in humans. Am J Clin Nutr. 99:843-850; 2014.

[50] Bloomfield, R.A.; Welsch, C. W.; Garner, G. B.; Muhrer, M. E. Effect of dietary nitrate on thyroid function. Science. 134:1690; 1961.

[51] Woessner, M.; Smoliga, J. M.; Tarzia, B.; Stabler, T.; Van Bruggen, M.; Allen, J. D. A stepwise reduction in plasma and salivary nitrite with increasing strengths of mouthwash following a dietary nitrate load. Nitric Oxide. 54:1-7; 2016.

[52] Kapil, V.; Milsom, A.B.; Okorie, M.; Maleki-Toyserkani, S.; Akram, F.; Rehman, F.; Arghandawi, S.; Pearl, V.; Benjamin, N.; Loukogeorgakis, S.; Macallister, R.; Hobbs, A. J.; Webb, A. J.; Ahluwalia, A. Inorganic nitrate supplementation lowers blood pressure in humans: role for nitrite-derived NO. Hypertension. 56:274-281; 2010.

[53] Gao, X.; Yang, T.; Liu, M.; Peleli, M.; Zollbrecht, C.; Weitzberg, E.; Lundberg, J. O,; Persson, A. E.; Carlström, M. NADPH oxidase in the renal microvasculature is a primary target for blood pressure-lowering effects by inorganic nitrate and nitrite. Hypertension. 65:161-170; 2015.

[54] EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). Scientific Opinion on Dietary Reference Values for iodine. EFSA Journal. 12:3660; 2014

[55] Murray, C. W.; Egan, S. K.; Kim, H.; Beru, N.; Bolger, P. M. US Food and Drug Administration's Total Diet Study: dietary intake of perchlorate and iodine. J Expo Sci Environ Epidemiol. 18:571-580; 2008.

#### **Figure Legends**

**Figure 1:** Salivary iodide concentration ([I<sup>-</sup>]) following no dietary supplementation (CON), supplementation with nitrate-rich beetroot juice (NIT) and co-supplementation with NIT and potassium iodide (NIT + I). The filled bars represent the group mean  $\pm$  SEM responses in the CON, NIT and NIT + I conditions. The solid grey lines represent the individual responses in the CON, NIT and NIT + I conditions. \* indicates significantly different from CON and NIT + I. # indicates significantly different from CON and NIT.

**Figure 2:** Salivary nitrate concentration ( $[NO_3^-]$ ) (upper panel), nitrite concentration ( $[NO_2^-]$ ) (middle panel) and nitrate + nitrite concentration ([NOx]) (lower panel) following supplementation with nitrate-rich beetroot juice (NIT) and co-supplementation with NIT and potassium iodide (NIT + I). Data are expressed as the change from the control condition without  $NO_3^-$  or I<sup>-</sup> supplementation. The filled bars represent the group mean  $\pm$  SEM responses while the solid grey lines represent the individual responses in the NIT and NIT + I conditions. \* indicates significantly different from NIT.

**Figure 3:** Plasma nitrate concentration ( $[NO_3^-]$ ) (upper panel) and nitrite concentration ( $[NO_2^-]$ ) (lower panel) following supplementation with nitrate-rich beetroot juice (NIT) and co-supplementation with NIT and potassium iodide (NIT + I). Data are expressed as the change from the control condition without  $NO_3^-$  or I<sup>-</sup> supplementation. The filled bars represent the group mean  $\pm$  SEM responses while the solid grey lines represent the individual responses in the NIT and NIT + I conditions. \* indicates significantly different from NIT.

**Figure 4:** Salivary-plasma nitrate  $[NO_3^-]$  (upper panel) and nitrite  $[NO_2^-]$  (lower panel) ratios following supplementation with nitrate-rich beetroot juice (NIT) and co-supplementation with NIT and potassium iodide (NIT + I). The filled bars represent the group mean  $\pm$  SEM responses while the solid grey lines represent the individual responses in the NIT and NIT + I conditions. \* indicates significantly different from NIT.

# Table 1. Plasma thyroid stimulating hormone ([TSH]) and thyroxine ([T4]) concentrations following no supplementation (CON), nitrate-rich beetroot juice supplementation (NIT) and co-supplementation with nitrate-rich beetroot juice supplementation and potassium iodide (NIT + I).

	CON	NIT	NIT + I
Plasma [TSH] (mIU/L)	$1.2 \pm 0.6$	$1.1 \pm 0.6$	$1.5 \pm 0.6$
Plasma [T <sub>4</sub> ] (µg/dl)	$4.2\pm0.7$	$3.8 \pm 0.4$	$3.8\pm0.9$

Values are presented as the mean  $\pm$  SD.

Table 2. Resting supine blood pressure measures following no supplementation (CON), nitrate-rich beetroot juice supplementation (NIT) and co-supplementation with nitrate-rich beetroot juice supplementation and potassium iodide (NIT + I).

	CON	NIT	NIT + I
Systolic blood pressure (mmHg)	$112 \pm 13$	106 ± 13*	106 ± 11*
Diastolic blood pressure (mmHg)	$59\pm 6$	$56 \pm 4*$	57 ± 5
Mean arterial pressure (mmHg)	$80\pm8$	75 ± 5*	76 ± 7*

Values are presented as the mean  $\pm$  SD. \* = significantly different from CON (*P*<0.05).

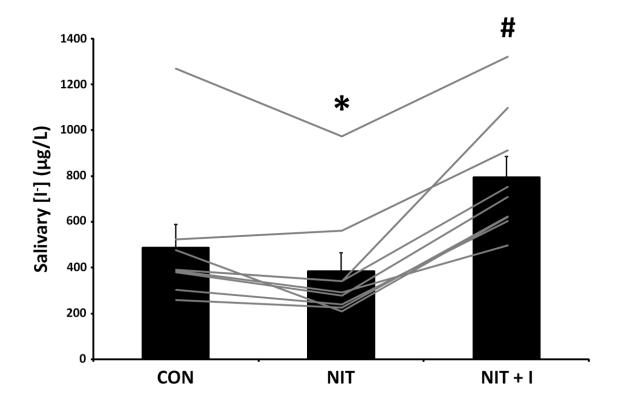
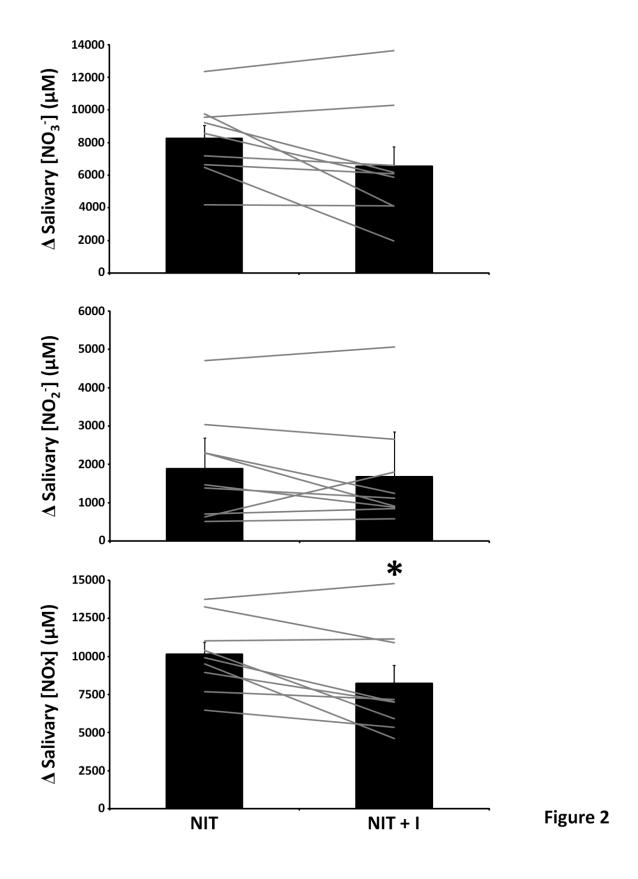


Figure 1



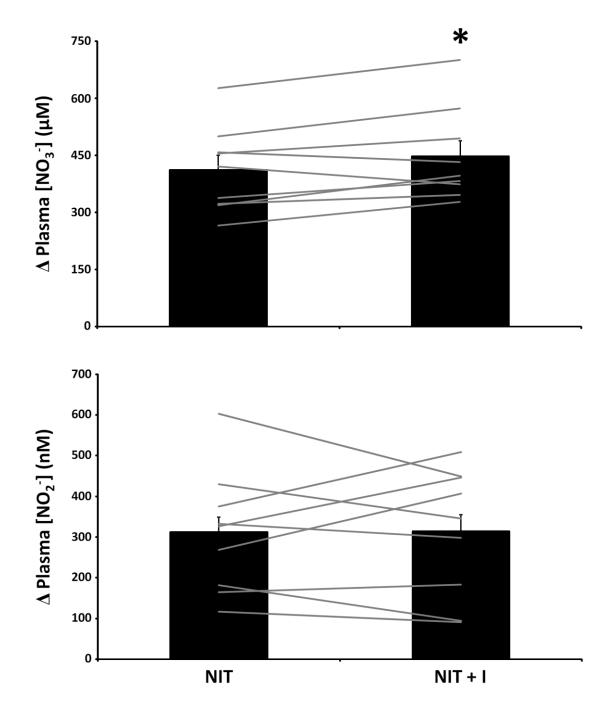


Figure 3

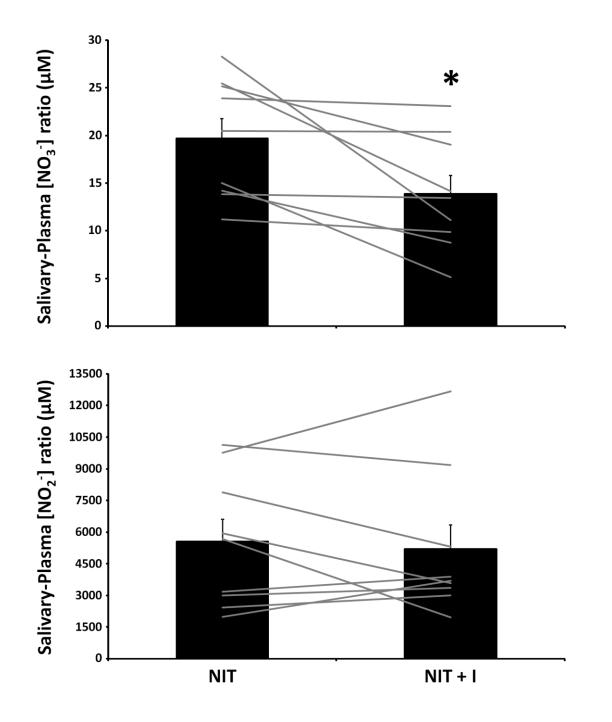


Figure 4