

1 **Title:** Climatic controls of decomposition drive the global biogeography of forest tree symbioses

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428 **Manuscript**

429 **The identity of the dominant microbial symbionts in a forest determines the ability of**
430 **trees to access limiting nutrients from atmospheric or soil pools^{1,2}, sequester carbon^{3,4} and**
431 **withstand the impacts of climate change¹⁻⁶. Characterizing the global distribution of**
432 **symbioses, and identifying the factors that control it, are thus integral to understanding**
433 **present and future forest ecosystem functioning. Here we generate the first spatially explicit**
434 **global map of forest symbiotic status using a database of over 1.1 million forest inventory**
435 **plots with over 28,000 tree species. Our analyses indicate that climatic variables, and in**
436 **particular climatically-controlled variation in decomposition rate, are the primary drivers of**
437 **the global distribution of major symbioses. We estimate that ectomycorrhizal (EM) trees,**
438 **which represent only 2% of all plant species⁷, constitute approximately 60% of tree stems on**
439 **Earth. EM symbiosis dominates forests where seasonally cold and dry climates inhibit**

440 decomposition, and are the predominant symbiosis at high latitudes and elevation. In
441 contrast, arbuscular mycorrhizal (AM) trees dominate aseasonally warm tropical forests
442 and occur with EM trees in temperate biomes where seasonally warm-and-wet climates
443 enhance decomposition. Continental transitions between AM and EM dominated forests
444 occur relatively abruptly along climate driven decomposition gradients, which is likely
445 caused by positive plant-microbe feedbacks. Symbiotic N-fixers, which are insensitive to
446 climatic controls on decomposition compared with mycorrhizal fungi, are most abundant in
447 arid biomes with alkaline soils and high maximum temperatures. The climatically driven
448 global symbiosis gradient we document represents the first spatially-explicit, quantitative
449 understanding of microbial symbioses at the global scale and demonstrates the critical role
450 of microbial mutualisms in shaping the distribution of plant species.

451 Microbial symbionts strongly influence the functioning of forest ecosystems. They exploit
452 inorganic, organic² and/or atmospheric forms of nutrients that enable plant growth¹, determine
453 how trees respond to elevated CO₂⁶, regulate the respiratory activity of soil microbes^{3,8}, and affect
454 plant species diversity by altering the strength of conspecific negative density dependence⁹.
455 Despite growing recognition of the importance of root symbioses for forest functioning^{1,6,10} and
456 the potential to integrate symbiotic status into Earth system models that predict functional changes
457 to the terrestrial biosphere¹⁰, we lack spatially-explicit, quantitative maps of the different root
458 symbioses at the global scale. Generating these quantitative maps of tree symbiotic states would
459 link the biogeography of functional traits of belowground microbial symbionts with their 3.1
460 trillion host trees¹¹, spread across Earth's forests, woodlands, and savannas.

461 The dominant guilds of tree root symbionts, arbuscular mycorrhizal (AM) fungi,
462 ectomycorrhizal (EM) fungi, ericoid mycorrhizal (ErM) fungi, and nitrogen (N)-fixing bacteria

463 (N-fixer) are all based on the exchange of plant photosynthate for limiting macronutrients. The
464 AM symbiosis evolved nearly 500 million years ago, with EM, ErM and N-fixer plant taxa
465 evolving multiple times from an AM basal state. Plants that form the AM symbiosis comprise
466 nearly 80% of all terrestrial plant species, and principally rely on AM fungi for enhancing mineral
467 phosphorus (P) uptake¹². In contrast to AM fungi, EM fungi evolved from multiple lineages of
468 saprotrophic ancestors, and as a result some EM fungi are more capable of mobilizing organic
469 sources of soil nutrients (particularly nitrogen)². Association with EM fungi, but not AM fungi,
470 has been shown to allow trees to accelerate photosynthesis in response to increased atmospheric
471 CO₂ when soil nitrogen (N) is limiting⁶ and to inhibit soil respiration by decomposer microbes^{3,8}.
472 Because increased plant photosynthesis and decreased soil respiration both reduce atmospheric
473 CO₂ concentrations, the EM symbiosis is associated with buffering the Earth's climate against
474 anthropogenic changes.

475 In contrast to mycorrhizal fungi, which extract nutrients from the soil, symbiotic N-fixers
476 (Rhizobia and Actinobacteria) convert atmospheric N₂ to plant-usable forms. Symbiotic N-fixers
477 are responsible for a large fraction of biological soil-N inputs, which can increase N-availability
478 in forests where they are locally abundant¹³. Both N-fixing bacteria and EM fungi often demand
479 more plant photosynthate than does the AM symbiosis^{12,14,15}. Because tree growth and
480 reproduction are limited by access to inorganic, organic and atmospheric sources of N, the
481 distribution of root symbioses is likely to reflect both environmental conditions that maximize the
482 cost-benefit ratio of symbiotic exchange as well as physiological constraints on different
483 symbionts.

484 In one of the earliest efforts to understand the functional biogeography of plant root
485 symbioses, Sir David Read¹⁶ categorically classified biomes by their perceived dominant

486 mycorrhizal type and hypothesized that seasonal climates favor hosts associating with EM fungi
487 due to their ability to compete directly for organic N. In contrast, it has been proposed that
488 sensitivity to low temperatures has prevented N-fixers from dominating outside the tropics, despite
489 the potential for N-fixation to alleviate N-limitation in boreal forests^{15,17}. However, global scale
490 tests of these proposed biogeographic patterns and their climate drivers are lacking. To address
491 this research gap, we compiled the first global ground-sourced survey database to reveal numerical
492 abundances of each symbiosis across the globe, which is essential for identifying the potential
493 mechanisms underlying transitions in forest symbiotic state along climatic gradients^{18,19}.

494 We determined the abundance of tree symbioses using GFBI, an extension from the plot-
495 based Global Forest Biodiversity (GFB) database, which contains over 1.1 million forest inventory
496 plots of individual-based measurement records from which we derive abundance information for
497 entire tree communities (Figure 1). Using published literature on the evolutionary histories of
498 mycorrhizal and N-fixer symbioses, we assigned plant species from the GFBI to one of 5 symbiotic
499 guilds: AM, EM, ErM, N-fixer, and non- or weakly-mycorrhizal (NM). We then used the random
500 forest algorithm with K-fold cross validation to determine the importance and influence of
501 variables related to climate, soil chemistry, vegetation, and topography on the relative abundance
502 of each tree-symbiotic guild (Figure 2). Because decomposition is the dominant process by which
503 soil nutrients become available to plants, we calculated annual and quarterly decomposition
504 coefficients according to the Yasso07 model²⁰, which describes how temperature and precipitation
505 gradients influence mass-loss rates of different chemical pools of leaf litter (with parameters fit
506 using a previous global study of leaf decomposition, Figures 3, S5). Finally, we projected our
507 predictive models across the globe over the extent global biomes that fell within the multivariate
508 distribution of our model training data (Figures 4, S14-15, see Methods for full description).

509 Our analysis shows that the three most numerically abundant tree symbiotic guilds each
510 have reliable environmental signatures, with the four most important predictors accounting for 81,
511 79, and 52% of the total variability in EM, AM, and N-fixer relative basal area, respectively.
512 Models for ErM and NM lack strong predictive power given the relative rarity of these symbiotic
513 states amongst trees, although the raw data do identify some local abundance hotspots for ErM
514 (Figure S1). As a result, we focus the remainder of results and discussion on the three major tree
515 symbiotic states (EM, AM, N-fixer). Despite the fact that data from N. and S. America constitute
516 65% of the training data (at the 1 by 1 degree grid scale), our models accurately predict the
517 proportional abundances of the three major symbioses across all major geographic regions (Figure
518 S10). The high performance of our models, which is robust to both K-fold cross-validation and
519 rarefying samples so that all continents are represented with equal depth (Figures S11-12), suggest
520 that regional variations in climate (including indirect effects on decomposition) and soil pH (for
521 N-fixers) are the primary factors influencing the relative dominance of each guild at the global
522 scale (geographic origin only explained ~2-5% of the variability in residual relative abundance)
523 (Table S8, Figure S10).

524 Whereas a recent global analysis of root traits concluded that plant evolution has favored
525 reduced dependence on mycorrhizal fungi²¹, we find that trees associating with the relatively more
526 C-demanding and recently-derived EM fungi^{12,14} represent the dominant tree-symbiosis. By taking
527 the average proportion of EM trees, weighted by spatially-explicit global predictions for tree stem
528 density¹¹, we estimate that approximately 60% of trees on earth are EM, despite the fact that only
529 2% of plant species associate with EM fungi (vs. 80% associating with AM fungi)⁷. Outside of the
530 tropics, the estimate for EM relative abundance increases to approximately 80% of trees.

531 Turnover among the major symbiotic guilds results in a tri-modal latitudinal abundance
532 gradient, with the proportion of EM trees increasing (and AM trees decreasing) with distance from
533 the equator, while the upper-quantiles of N-fixing trees reach peak abundance in the arid zone
534 around 30 degrees (Figure 3A, Figure 4). These trends are driven by abrupt transitional regions
535 along continental climatic gradients (Figure 2), which skew the distribution of symbioses among
536 biomes (Figure 3A) and drive strong patterns across geographic and topographic features that
537 influence climate. Moving north or south from the equator, the first transitional zone separates
538 warm (aseasonal), AM-dominated, tropical broadleaf forests (>75% median basal area, vs. 8% for
539 EM trees) from the rest of the EM-dominated world forest system (Figure 2AB; Figure 3A). The
540 transition zone occurs across the globe around 25 degrees N and S latitude, just beyond the dry
541 tropical broadleaf forests (with 25% EM tree basal area; Figure 3A), where average monthly
542 temperature variation reaches 3-5°C (temperature seasonality, Figure 2AB).

543 Moving further N or S, the second transitional climate zone separates regions where
544 decomposition coefficients during the warmest quarter of the year are less than 2 (see Figure 3B
545 for the associated temperature and precipitation ranges). In N. America and China, this transition
546 zone occurs around 50 degrees N, separating the mixed AM / EM temperate forests from their
547 neighboring EM dominated boreal forests (75 vs 100% EM tree basal area, respectively; Figure
548 3A). This transitional decomposition zone bypasses W. Europe, which has temperature seasonality
549 > 5°C, but lacks sufficiently wet summers to accelerate decomposition coefficients beyond values
550 associated with mixed AM/EM forests. The latitudinal transitions in symbiotic state observed
551 among biomes are mirrored by within-biome transitions along elevation gradients. For example,
552 in tropical Mexico, warm and wet quarter decomposition coefficients < 2 occur along the slopes
553 of the Sierra Madre, where mixed AM-exclusive and N-fixer woodlands in arid climates transition

554 to EM dominated tropical coniferous forests (75% basal area, Figure 3A, Figure 4ABC, Figure
555 S16-18). The southern hemisphere, which lacks the landmass to support extensive boreal forests,
556 experiences a similar latitudinal transition in decomposition rates along the ecotone separating its
557 tropical and temperate biomes, around 28 degrees S.

558 The abrupt transitions that we detected between forest symbiotic states along
559 environmental gradients suggest that positive feedbacks may exist between climatic and biological
560 controls of decomposition^{10,20}. In contrast to AM fungi, some EM fungi can use oxidative enzymes
561 to mineralize organic nutrients from leaf litter, converting nutrients to plant-usable forms^{2,5}.
562 Relative to AM trees, the leaf litter of EM trees is also chemically more resistant to decomposition,
563 with higher C:N ratios and higher concentrations of decomposition-inhibiting secondary
564 compounds¹⁰. Thus, EM leaf litter can exacerbate climatic barriers to decomposition, promoting
565 conditions where EM fungi have superior nutrient-acquiring abilities to AM-fungi^{5,10}. A recent
566 game theoretical model has shown that positive plant-soil-nutrient feedbacks can lead to local
567 bistability in mycorrhizal symbiosis²². Such positive-feedbacks are also known to cause abrupt
568 ecosystem transitions along smooth environmental gradients between woodlands and grasses: trees
569 suppress fires, which promotes seedling recruitment, while grass fuels fires, which kill tree
570 seedlings²³. The existence of abrupt transitions also suggests that forests in transitional regions
571 along decomposition gradients should be susceptible to drastic turnover in symbiotic state with
572 future environmental changes²³.

573 To illustrate the sensitivity of global patterns of tree symbiosis to climate change, we use
574 the relationships we developed for current climate to project potential changes in forest symbiotic
575 status in the future. Relative to our global predictions using the most recent climate data, model
576 predictions using the projected climates for 2070 suggest the abundance of EM trees will decline

577 by as much as 10% (using a relative concentration pathway of 8.5 W/m²; Figure S24). Due to their
578 position along decomposition gradients relative to the abrupt shift from EM to AM forests (Figure
579 2AB), our models predict the largest declines in EM abundance will occur along the boreal-
580 temperate ecotone, although this model does not estimate the time lags between climate change
581 and forest community responses. The predicted decline in EM trees corroborates the results of
582 common garden transfer and simulated warming experiments, which demonstrate that some
583 important EM hosts will decline at the boreal-temperate ecotone in altered climates²⁴.

584 The change in dominant nutrient exchange symbioses along climate gradients highlights
585 the interconnection between atmospheric and soil compartments of the biosphere. The transition
586 from AM to EM dominance corresponds with a shift from P to N limitation of plant growth with
587 increasing latitude^{25,26}. Including published global projections of total soil N or P, microbial N, or
588 soil P fractions (labile, occluded, organic, and apatite) did not increase the amount of variation
589 explained by the model or alter the variables identified as most important, and thus were dropped
590 from our analysis. However, our finding that climatic controls of decomposition best predict the
591 dominant mycorrhizal associations mechanistically links symbiont physiology with climatic
592 controls of soil nutrient release from leaf litter. These findings are consistent with Read's
593 hypothesis¹⁶ that slow decomposition at high latitudes favors EM fungi due to their increased
594 capacity to liberate organic nutrients². Thus, while more experiments are necessary to understand
595 the specific mechanism by which nutrient competition favors dominance of AM or EM
596 symbioses¹⁸, we propose that the latitudinal and elevational transitions from AM to EM dominated
597 forests be called Read's Rule.

598 While our analyses focus on prediction at large spatial scales appropriate to the available
599 data, our findings with respect to Read's Rule also provide insight into how soil factors structure

600 the fine-scale distributions of tree symbioses within our grid cells. For example, while at a coarse
601 scale we find that EM trees are relatively rare in many wet tropical forests, individual tropical sites
602 in our raw data span the full range from 0 – 100 % EM basal area. In much of the wet tropics, these
603 EM dominated sites exist as outliers within a matrix of predominantly AM trees. In an apparent
604 exception that proves Read’s Rule, in aseasonal warm neotropical climates, which accelerate leaf-
605 decomposition and promote regional AM dominance (Figure 3), EM dominated tree stands can
606 develop in sites where poor soils and recalcitrant litter slow decomposition and N
607 mineralization^{18,27}. Landscape-scale variation in the relative abundance of symbiotic states also
608 changes along climate gradients, with variability highest in xeric and temperate biomes (Figure
609 S3-4), suggesting that the potential of local nutrient variability to favor particular symbioses is
610 contingent on climate.

611 Whereas EM trees are associated with ecosystems where plant growth is thought to be
612 primarily N-limited, N-fixer trees are not. Our results highlight the global extent of the “N-cycling
613 paradox,” wherein some metrics suggest that N-limitation is greater in the temperate zone^{25,26}, yet
614 N-fixing trees are relatively more common in the tropics^{15,28} (Figure 3A). We find that N-fixers,
615 which we estimate represent 7% of all trees, dominate forests with annual max temperatures >35°C
616 and alkaline soils (particularly in North America and Africa, Figure 2C). They have the highest
617 relative abundance in xeric shrublands (24%), tropical savannas (21%), and dry broadleaf forest
618 biomes (20%), but are nearly absent from boreal forests (<1%) (Figure 3A, Figure 4). The decline
619 in N-fixer tree abundance we observed with increasing latitude is also associated with a previously
620 documented latitudinal shift in the identity of N-fixing microbes, from facultative N-fixing
621 rhizobial bacteria in tropical forests to obligate N-fixing actinorhizal bacteria in temperate
622 forests²⁸. Our data are not capable of fully disentangling the several hypotheses that have been

623 proposed to reconcile the N-cycling paradox¹⁵. However, our results are consistent with the model
624 prediction¹⁷ and regional empirical evidence^{19,29,30} that N-fixing trees are particularly important in
625 arid biomes. Based primarily on the observed positive, nonlinear association of N-fixer relative
626 abundance with the mean temperature of the hottest month (Figure 2C), our models predict a two-
627 fold increase in N-fixer relative abundance when transitioning from humid to dry tropical forest
628 biomes (Figure 3A).

629 Although soil microbes are a dominant component of forests, both in terms of diversity and
630 ecosystem functioning^{5,6,10}, identifying global-scale microbial biogeographic patterns remains an
631 ongoing research priority. Our analyses confirm that Read's Rule, which is one of the first
632 proposed biogeographic rules specific to microbial symbioses, successfully describes global
633 transitions between mycorrhizal guilds. More generally, climate driven turnover among the major
634 plant-microbe symbioses represents a fundamental biological pattern in the Earth system, as forests
635 transition from low-latitude arbuscular mycorrhizal, to N-fixer, to high-latitude ectomycorrhizal
636 ecosystems. The predictions of our model (which we make available as a global raster layer) can
637 now be used to represent these critical ecosystem variations in global biogeochemical models used
638 to predict climate-biogeochemical feedbacks within and between trees, soils, and the atmosphere.
639 Additionally, the layer containing the proportion of N-fixing trees can be used to map potential
640 symbiotic N-fixation, which links together atmospheric pools of C and N. Future work can extend
641 our findings to incorporate multiple plant growth forms and non-forested biomes, where similar
642 patterns likely exist, to generate a complete global perspective. Our predictive maps leverage the
643 most comprehensive global forest dataset to generate the first quantitative global map of forest tree
644 symbioses, demonstrating how nutritional mutualisms are coupled with the global distribution of
645 plant communities.

646 **References**

- 647 1 Batterman, S. A. *et al.* Key role of symbiotic dinitrogen fixation in tropical forest
 648 secondary succession. *Nature* **502**, 224-227, doi:10.1038/nature12525 (2013).
- 649 2 Shah, F. *et al.* Ectomycorrhizal fungi decompose soil organic matter using oxidative
 650 mechanisms adapted from saprotrophic ancestors. *New Phytol* **209**, 1705-1719,
 651 doi:10.1111/nph.13722 (2016).
- 652 3 Averill, C., Turner, B. L. & Finzi, A. C. Mycorrhiza-mediated competition between
 653 plants and decomposers drives soil carbon storage. *Nature* **505**, 543-+,
 654 doi:10.1038/nature12901 (2014).
- 655 4 Clemmensen, K. E. *et al.* Roots and associated fungi drive long-term carbon
 656 sequestration in boreal forest. *Science* **339**, 1615-1618, doi:10.1126/science.1231923
 657 (2013).
- 658 5 Cheeke, T. E. *et al.* Dominant mycorrhizal association of trees alters carbon and nutrient
 659 cycling by selecting for microbial groups with distinct enzyme function. *New Phytol.*
 660 **214**, 432-442, doi:10.1111/nph.14343 (2017).
- 661 6 Terrer, C., Vicca, S., Hungate, B. A., Phillips, R. P. & Prentice, I. C. Mycorrhizal
 662 association as a primary control of the CO₂ fertilization effect. *Science* **353**, 72-74,
 663 doi:10.1126/science.aaf4610 (2016).
- 664 7 Brundrett, M. C. in *Biogeography of Mycorrhizal Symbiosis* 533-556 (Springer, 2017).
- 665 8 Averill, C. & Hawkes, C. V. Ectomycorrhizal fungi slow soil carbon cycling. *Ecol Lett*
 666 **19**, 937-947, doi:10.1111/ele.12631 (2016).
- 667 9 Bennett, J. A. *et al.* Plant-soil feedbacks and mycorrhizal type influence temperate forest
 668 population dynamics. *Science* **355**, 181-184 (2017).
- 669 10 Phillips, R. P., Brzostek, E. & Midgley, M. G. The mycorrhizal-associated nutrient
 670 economy: a new framework for predicting carbon-nutrient couplings in temperate forests.
 671 *New Phytol.* **199**, 41-51, doi:10.1111/nph.12221 (2013).
- 672 11 Crowther, T. W. *et al.* Mapping tree density at a global scale. *Nature* **525**, 201 (2015).
- 673 12 Heijden, M. G., Martin, F. M., Selosse, M. A. & Sanders, I. R. Mycorrhizal ecology and
 674 evolution: the past, the present, and the future. *New Phytol.* **205**, 1406-1423 (2015).
- 675 13 Binkley, D., Sollins, P., Bell, R., Sachs, D. & Myrold, D. Biogeochemistry of adjacent
 676 conifer and alder-conifer stands. *Ecology* **73**, 2022-2033 (1992).
- 677 14 Leake, J. *et al.* Networks of power and influence: the role of mycorrhizal mycelium in
 678 controlling plant communities and agroecosystem functioning. *Canadian Journal of*
 679 *Botany* **82**, 1016-1045 (2004).
- 680 15 Hedin, L. O., Brookshire, E. N. J., Menge, D. N. L. & Barron, A. R. in *Annual Review of*
 681 *Ecology Evolution and Systematics* Vol. 40 *Annual Review of Ecology Evolution and*
 682 *Systematics* 613-635 (Annual Reviews, 2009).
- 683 16 Read, D. J. Mycorrhizas in Ecosystems. *Experientia* **47**, 376-391, doi:Doi
 684 10.1007/Bf01972080 (1991).
- 685 17 Houlton, B. Z., Wang, Y.-P., Vitousek, P. M. & Field, C. B. A unifying framework for
 686 dinitrogen fixation in the terrestrial biosphere. *Nature* **454**, 327 (2008).
- 687 18 Peay, K. G. The mutualistic niche: mycorrhizal symbiosis and community dynamics.
 688 *Annual Review of Ecology, Evolution, and Systematics* **47**, 143-164 (2016).

- 689 19 Pellegrini, A. F., Staver, A. C., Hedin, L. O., Charles-Dominique, T. & Tourgee, A.
690 Aridity, not fire, favors nitrogen-fixing plants across tropical savanna and forest biomes.
691 *Ecology* **97**, 2177-2183 (2016).
- 692 20 Tuomi, M. *et al.* Leaf litter decomposition—estimates of global variability based on
693 Yasso07 model. *Ecological Modelling* **220**, 3362-3371 (2009).
- 694 21 Ma, Z. *et al.* Evolutionary history resolves global organization of root functional traits.
695 *Nature* (2018).
- 696 22 Lu, M. & Hedin, L. O. Global plant–symbiont organization and emergence of
697 biogeochemical cycles resolved by evolution-based trait modelling. *Nature ecology &*
698 *evolution*, 1 (2019).
- 699 23 Scheffer, M., Carpenter, S., Foley, J. A., Folke, C. & Walker, B. Catastrophic shifts in
700 ecosystems. *Nature* **413**, 591 (2001).
- 701 24 Reich, P. B. *et al.* Geographic range predicts photosynthetic and growth response to
702 warming in co-occurring tree species. *Nature Climate Change* **5**, 148 (2015).
- 703 25 McGroddy, M. E., Daufresne, T. & Hedin, L. O. Scaling of C: N: P stoichiometry in
704 forests worldwide: Implications of terrestrial redfield-type ratios. *Ecology* **85**, 2390-2401
705 (2004).
- 706 26 Reich, P. B. & Oleksyn, J. Global patterns of plant leaf N and P in relation to temperature
707 and latitude. *Proceedings of the National Academy of Sciences of the United States of*
708 *America* **101**, 11001-11006 (2004).
- 709 27 Corrales, A., Mangan, S. A., Turner, B. L. & Dalling, J. W. An ectomycorrhizal nitrogen
710 economy facilitates monodominance in a neotropical forest. *Ecol Lett* **19**, 383-392,
711 doi:10.1111/ele.12570 (2016).
- 712 28 Menge, D. N., Lichstein, J. W. & Ángeles-Pérez, G. Nitrogen fixation strategies can
713 explain the latitudinal shift in nitrogen-fixing tree abundance. *Ecology* **95**, 2236-2245
714 (2014).
- 715 29 Liao, W., Menge, D. N., Lichstein, J. W. & Ángeles-Pérez, G. Global climate change will
716 increase the abundance of symbiotic nitrogen-fixing trees in much of North America.
717 *Global Change Biol* (2017).
- 718 30 Gei, M. *et al.* Legume abundance along successional and rainfall gradients in Neotropical
719 forests. *Nature ecology & evolution*, 1 (2018).
- 720

721 **Figure Legends**

722 **Figure 1. The global distribution of GFBi training data. The global map has n=2,768 grid**
723 **cells at a 1 x 1 degree latitude/longitude resolution. Cells are colored in the red, green and**
724 **blue spectrum according to the % of total tree basal area occupied by N-fixer, AM, and EM**
725 **tree symbiotic guilds, as indicated by the ternary plot. Grey cells show the global distribution**
726 **of forests where we make model projections.**

727

728 **Figure 2. A small number of environmental variables predict the majority of global turnover**
729 **in forest symbiotic status. Panels show the partial feature contributions of different**
730 **environmental variables on forest symbiotic state. Each row plots the shape of the**
731 **contribution of the four most important predictors of the proportion of tree basal area**
732 **belonging to the (a) ectomycorrhizal (EM), (b) arbuscular mycorrhizal (AM), and (c) N-fixer**
733 **symbiotic guilds (n=2,768). Variables are listed in declining importance from left to right, as**
734 **determined by inc node purity, with points colored with a red-green-blue gradient according**
735 **to their position on the x-axis of the most important variable (left-most panels for each guild),**
736 **allowing cross visualization between predictors. Each panel lists two measures of variable**
737 **importance, inc node purity (used for sorting) and %IncMSE (see Supplemental Information**
738 **for description). The abundance of each symbiont type transitions sharply along climatic**
739 **gradients, suggesting that sites near the threshold are particularly vulnerable to switching**
740 **their dominant symbiont guild with climate changes.**

741
742 **Figure 3. The distribution of forest symbiotic status across biomes is related to climatic**
743 **controls over decomposition. (a) Biome level summaries of the median +/- 1 quartile of the**
744 **predicted % tree basal area per biome for ectomycorrhizal (EM), arbuscular mycorrhizal**
745 **(AM), and N-fixer symbiotic guilds (n=100 random samples per biome). (b) The dependency**
746 **of decomposition coefficients (k, solid and dotted lines) on temperature and precipitation**
747 **during the warmest quarter with respect to predicted dominance of mycorrhizal symbiosis.**
748 **The transition from AM forests to EM forests between k=1 and 2 is abrupt, which is**
749 **consistent with positive feedback between climatic and biological controls of decomposition.**
750

751 **Figure 4. Global maps of predicted forest tree symbiotic state. Maps (left) and latitudinal**
752 **gradients (right, with solid line indicating the median and colored ribbon spanning the range**
753 **from the 5% and 95% quantiles) of the % of tree basal area for (a) ectomycorrhizal (EM),**
754 **(b) arbuscular mycorrhizal (AM), and (c) N-fixer symbiotic guilds. All projections are**
755 **displayed a 0.5 by 0.5 degree lat/long scale with n=28,454.**

756 **Acknowledgements**

757 This work was made possible by the Global Forest Biodiversity Database, which represents the
758 work of over 200 independent investigators and their public and private funding agencies (see
759 Supplementary Acknowledgements).

760 **Author Contributions**

761 KGP & TWC conceived the study; TWC, JL, PBR, GN, SdM, MZ, NP, BH, XZ, & CZ
762 conceived and organized the GFBi database; KGP, BSS, GDAW, & MVN compiled the
763 symbiosis database; BSS carried out the primary data analysis; MVN & DR contributed to data
764 compilation and analysis; BSS, TWC, MVN & KGP wrote the initial manuscript; BSS, TWC,
765 JL, MVN, GDAW, PBR, GN, SdM, MZ, NP, BH, XZ, CZ & KGP made substantial revisions to
766 all versions of the manuscript; all other named authors provided forest inventory data and
767 commented on the manuscript.

768 **Data Availability**

769 The GFBi database is available upon written request at <https://www.gfbinitiative.org/datarequest>.
770 Additionally, the symbiotic state assigned to tree species as a supplementary file, as are global
771 rasters of our model projections for EM, AM, and N-fixer proportion of tree basal area.

772 **Conflict of Interest**

773 The authors declare we have no conflict of interests.

774 **Supplementary Information**

775 For more information on symbiotic guild assignments, model selection, and supplementary
776 analyses, refer to SupplementaryInfo_Steidinger_etal2019.pdf. For the full suite of Supplemental
777 Files, including symbiotic guild assignments and rasters of model projections, refer to
778 SupplementalFiles_Steidinger_etal2019.zip

779

780 **Methods**

781 We quantified the relative abundance of tree symbiotic guilds across >1.1 million forest
782 census plots combined in the GFBi database, an extension from the plot-based Global Forest
783 Biodiversity (GFB³¹) database. The GFBi database consists of individual-based data that we
784 compiled from all the regional and national GFBi forest inventory data sets. The standardized
785 GFBi data frame, i.e. tree list, comprises tree ID, a unique number assigned to each individual
786 tree; plot ID, a unique string assigned to each plot; plot coordinates, in decimal degrees of
787 WGS84 datum; tree size, in diameter-at-breast-height; trees-per-hectare expansion factor; year of
788 measurement; data set name, a unique number assigned to each forest inventory data set; and
789 binomial scientific tree species names.

790 We error checked all species names from different forest inventory data sets in three steps.
791 First, we extracted scientific names from original data sets, keeping only the names of genus and
792 species (authority names are removed). Next, we compiled all the species names into five general
793 species lists, one for each continent. Finally, we verified individual species names against 23 online
794 taxonomic databases using the ‘taxize’ package of R programming language³². We assigned each
795 morphospecies a unique name comprising the genus, the string “spp”, followed by the data set
796 name and a unique number for that species. For example, “Picea sppCNI1” and “Picea sppCNI2”

797 represent two different species under the genus “Picea”, observed in the first Chinese data set
798 (CNI).

799 We derived plot-level abundance information in terms of species abundance matrices. Each
800 species abundance matrix consisted of the number of individuals by species (column vectors)
801 within individual sample plots (row vectors). In addition, key plot-level information was also
802 added to the matrices, including plot ID, data set name, plot coordinates, the year of measurement,
803 and basal area, i.e. the total cross-sectional areas (m²) of living trees per one hectare of ground
804 area.

805 Tree genera were assigned to a plant family using a plant taxonomy lookup table generated
806 by Will Cornwell (hosted on Github <https://github.com/traitecoevo/taxonlookup>), which uses the
807 accepted taxonomy from “The Plant List.” The majority (96.5%) of genera from the GFBi species
808 were successfully matched to family; for those that could not be assigned, we manually checked
809 the GFBi genus and species against synonyms from The Plant List. Of the remaining 1,038
810 mismatches, an additional 440 were assigned to family either by updating older genera and species
811 names with their more recent synonyms or else by correcting obvious misspellings. The remaining
812 598 entries that could not be matched to family were excluded from analysis.

813 We used a taxonomically-informed approach to assign symbiotic states to plant species
814 from the GFBi. Plant species were assigned to one of 5 symbiotic guilds – ectomycorrhizal (EM),
815 arbuscular mycorrhizal (AM), ericoid mycorrhizal (ErM), weakly AM or non-mycorrhizal
816 (AMNM), or N-fixer (Table S1). Although we did not model the relative abundance of ErM trees,
817 due to their rarity, we have included a map of their relative abundance from our grid (Figure S1).
818 We also include as a supplementary file the full species list, which includes columns used to assign
819 species to guild. In addition, we include here a list of families and genera assigned to all guilds

820 except AM (Tables S2-5) with notes for cases of species from individual genera that were either
821 assigned to two guilds simultaneously (e.g., *Alnus* is an N-fixer and EM) or where species from
822 individual genera were split between two different guilds (e.g., some *Pisonia* sp. are weakly AM
823 and some are EM). An AM summary table is excluded for length considerations—the same
824 information is available in the Supplementary File “SymbioticGuildAssignment.csv”.

825 The taxonomy of species in our inventory was compared with recently published literature
826 on the evolutionary history of mycorrhizal symbiosis^{7,33,34} and N-fixation³⁵⁻³⁸. Most species
827 symbiotic status could be reliably assigned at the genus (e.g. *Dicymbe*) or family level (e.g.
828 Pinaceae). For the few groups where status was unreliable or variable within a genus (e.g. *Pisonia*)
829 we conducted additional literature searches.

830 We assigned species to the EM category in three stages. First, at the family level (e.g.,
831 Pinaceae); next, as the genus level (e.g., *Dicymbe*); and finally, using literature searches for unclear
832 genera. For example, for the genus *Pisonia*, some species are AM and others are EM. We used a
833 published list from Hayward & Hynson (2014)³⁹ to sort species into the appropriate guild. For the
834 genus *Acacia*, we followed Brundrett (2017)⁷ in assuming that only endemic Australian species
835 associate with EM, while all others are AM (we sorted *Acacia* species according to provenance
836 using <http://worldwidewattle.com/>).

837 The AMNM category lumped together all genera of terrestrial, non-epiphytic plants that
838 either lack arbuscular mycorrhizal fungi (AMF), or have low or inconsistent records of AMF
839 colonization of roots. For example, although there are some published records of AMF
840 colonization in the roots of Proteaceae, these records are inconsistent, and colonization is generally
841 low. Further, as Proteaceae are associated with a non-mycorrhizal root morphology (the “cluster”
842 or “proteoid root system”) that allows them to access otherwise unavailable forms of soil

843 nutrients⁴⁰, we placed the entire family within AMNM. The family Urticaceae, which we also
844 characterized as AMNM, was somewhat problematic – early-successional species from tropical
845 forests, such as those in the genus *Cecropia*, have records of both low and absent AMF
846 colonization⁴¹. Our approach was to use the most broadly inclusive AMNM categorization.

847 N-fixer status was assigned at the genus level, using previously compiled databases of
848 global symbiotic N₂-fixation³⁵⁻³⁸. Given that symbiotic N₂-fixation with rhizobial or *Frankia*
849 bacteria has only evolved in four orders (Rosales, Cucurbitales, Fabales and Fagales)⁴², all species
850 outside of this nitrogen-fixing clade were assigned non-fixing status. Some species could not be
851 assigned a N-fixer status because they were typed to a higher taxonomic level (e.g. family) that is
852 ambiguous from a N-fixer status perspective. We recorded when our assignment of N-fixer status
853 was based on phylogenetic criteria but where symbiotic N-fixation is evolutionarily labile. Since
854 these cases are more likely to be misassigned we excluded them from the N-fixation category. The
855 N-fixer group contains species that are colonized by AMF (e.g., most genera from Leguminosae)
856 and others that are colonized by ectomycorrhizal fungi (e.g., *Alnus* sp.).

857 Most plant species form AM symbiosis, which is the basal symbiotic state to the later
858 derived EM and N-fixing symbioses. Further, many EM and N-fixing plants maintain the ability
859 to form AM symbiosis. Thus, a tree species is most likely AM if it does not form associations with
860 another symbiotic guild (or forgoes root symbiosis entirely), as evidenced by their inclusion in
861 exhaustive databases of plant symbiotic state^{7,33-38,41}. In keeping with other large-scale studies in
862 the field (e.g. ³⁴), we assigned tree species from the GFBi database to an AM-exclusive state if
863 they belonged to taxa that were not matched to EM, ErM, non-or-weakly mycorrhizal or N-fixer
864 symbioses. Thus, the AM and N-fixer groups in our dataset are non-overlapping despite the fact
865 that most N-fixers also associate with AM fungi.

866 The proportions of tree basal area and tree individuals were aggregated to a 1' by 1' degree
867 grid by taking the weighted average of the plot-level proportions (Table S6). This resulted in a
868 total of 2,768 grid cells, each with a score for the proportional abundance of EM, AM, N-fixer,
869 ErM, and AMNM trees. We calculated two measures of relative abundance for each symbiotic
870 guild: proportion of tree stems and proportion of tree basal area. Because the measurements are
871 highly correlated with one another (Figure S2) we chose to model only proportion of total tree
872 basal area, which should scale more approximately to proportion of tree biomass as it accounts for
873 differences in size among individual stems. Additionally, we quantified variability among plots
874 within each grid cell by calculating the weighted standard deviation across the grid (Supplemental
875 Information, Figure S3-4).

876 To identify the key factors structuring symbiotic distributions we assembled 70 global
877 predictor layers: 19 climatic (annual, monthly, and quarterly temperature and precipitation
878 variables), 14 soil chemical (total soil N density, microbial N, C:N ratios and soil P fractions, pH,
879 cation exchange capacity), 5 soil physical (soil texture and bulk density), 26 vegetative indices
880 (leaf area index, total stem density, enhanced vegetation index means and variances), and 5
881 topographic variables (elevation, hillshade) (Table S7). Because decomposition is the dominant
882 process by which soil nutrients become available to plants, we generated 5 additional layers that
883 estimate the climatic control of decomposition. We parameterized decomposition coefficients
884 according to the Yasso07 model^{20,43} using the following equation:

885

$$k = \text{Exp}(0.095T_i - 0.00014 T_i^2) (1 - \text{Exp}[-1.21 P_i]), \quad (1)$$

886 where P_i and T_i are precipitation and mean temperature, either quarterly or annually, and the
887 constants 0.0095 ($=\beta_1$) -0.00014 ($=\beta_2$), and -1.21 ($=\gamma$) are parameters fit using a previous global
888 study of leaf litter mass-loss²⁰. Although local decomposition rates can vary significantly based on

889 litter quality or microbial community composition⁴⁴, climate is the primary control at the global
890 scale²⁰. Decomposition coefficients describe how fast different chemical pools of leaf litter lose
891 mass over time relative to a parameter, α , that accounts for leaf-chemistry. Decomposition
892 coefficients (k) with values of 0.5 and 2 indicate a halving and doubling of decomposition rates
893 relative to α , respectively (Supplemental Information, Figure S5).

894 We implemented the random forest algorithm using the “randomForest” packaged in R.
895 Random forest models average over multiple regression trees, each of which uses a random subset
896 of all the model variables to predict a response. We first determined the influence and relationship
897 of all 75 predictor layers on forest symbiotic state and then optimized our models using a stepwise
898 reduction in variables, from least- to most-important. Variable importance was measured in two
899 ways: Inc Node Purity and %IncMSE (with values reported in each panel of Figure 2). The inc
900 node purity of variable x considers the decrease in the residual sum of squares that results from
901 splitting regression trees using variable x . %IncMSE (mean square error) quantifies the increase
902 in model error as a result of randomly shuffling the order of values in the vector x . We chose to
903 rank variables according to inc node purity because we found that higher inc node purities were
904 associated with larger effect sizes, whereas larger %IncMSE were associated with more linear
905 responses of smaller effect. Whereas our inspection of partial feature contributions is derived from
906 univariate random forest models, we additionally ran multivariate random forests to predict the
907 proportional abundance of EM, AM, and N-fixer trees for each pixel. The multivariate models
908 were run using 50-regression trees each, with the unique set of the best 4 predictor variables for
909 each symbiotic guild in the univariate models (Table S7, Figure 2). Despite strong negative
910 correlations between the proportions of EM and AM basal area (Figure S22), the results from
911 multivariate and univariate random forests are strongly correlated with one another (Figure S23).

912 Using model selection based on eliminating variables with low Inc Node Purity, we
913 removed most soil nutrient, vegetative, and topographic variables from our models (Figure S6-7).
914 Our final models include the remaining 34 predictor layers with climate, decomposition, and
915 certain soil physical and chemical information (Figure S8). To determine the parsimony of our
916 models, we compared the coefficient of determination in models run with a stepwise reduction in
917 the number of variables (starting with those with the lowest Inc Node Purity). Based on
918 performance of the ratio of coefficient of determination in models with 4 vs 34 variables, we
919 determined that the 4 most important variables accounted for >85% of the explained variability
920 (Figure S9). We also compared model performance visually with plots of actual vs predicted
921 proportions of each tree symbiotic guild among continents and geographic subregions (Figure
922 S10). We used the “forestFloor” packaged in R to plot the partial variable response of tree
923 symbiotic guilds to each predictor variable (Figure 2ABC, see Figure S19-21 for partial plots of
924 the partial feature contributions of all 34 variables).

925 In order to test the sensitivity of model performance and predictions, we performed cross
926 validation in R using the “rfUtilities” package²⁴. K-fold cross validation tests the sensitivity of
927 model predictions to losing random subsets from the training data. For EM, AM, and N-fixer
928 models we ran 99 iterations that withheld 10% of the model training data. We assessed the drop in
929 model performance in the 99 iterations by manually calculating the coefficient of determination,
930 which uses the following formula: $1 - \frac{\sum (\text{actual \% basal area} - \text{predicted \% basal area})^2}{\sum (\text{actual \% basal area} - \text{mean actual \% basal area})^2}$. For all symbiotic guilds, withholding 10% of
931 the training data resulted in a mean loss in variance explained of less than 1% (Figure S11). This
932 shows that our training data has sufficient redundancy to ensure that our model conclusions are
933 robust. Similarly, to determine whether our random forest models would make similar predictions
934

935 if data were equally distributed among continents, we rarefied our aggregated grid of symbiotic
936 states and predictor layers to an even depth. Specifically, we sub-sampled all continents – N.
937 America (including Central America and the Caribbean), S. America, Europe, Asia, and Oceania
938 – to match the number of grid-pixels from Africa (n=50). This is a much more aggressive reduction
939 of training data than is typically used in K-fold cross validations, as it involves dropping ~90% of
940 training data rather than retaining the same amount. We performed 99 iterations of rarefaction each
941 for the three symbiotic guilds. On average, models run with the rarefied data explained about 10%
942 less variance over the full training data (the entire predictor / response grid) than did models run
943 with all of the training data (Figure S12-13).

944 To avoid projecting our random forest models outside the ranges of their training data (e.g.,
945 grid cells with higher mean annual temperatures than the maximum used to fit the models), we
946 subset a global grid of predictor layers depending on whether (1) the grid cell fell within the top
947 60% of land surface with respect to tree stem density¹¹ and either (2) fell within the univariate
948 distribution of all the predictor layers from our training data and/or (3) fell within an 8-dimensional
949 hypervolume defined by the unique set of the 4-best predictors of the relative abundance of each
950 guild (Figure S14). We then projected our models across only those grid cells that met these
951 criteria, which constitutes 46% of the global land surface and 88% of global tree stems (Figure 1;
952 Figure S15). Model projections were made at two resolutions: both 1 by 1 degree and 0.5 by 0.5
953 degree resolution (Figure 4). While model validation indicates that our projections are robust,
954 additional ground truthing of predictions to identify any discrepancies would be incredibly
955 valuable. If such discrepancies exist they can help fine tune climate-symbiosis models, or identify
956 areas where climate might favour invasion by symbioses that have not yet evolved or dispersed to
957 a particular biogeographic region.

958 We used the following equation to estimate the % of global tree stems that belong to each
959 tree symbiotic guild: \sum_i (predicted proportion of trees of guild g in pixel i) x (total number of tree
960 stems in pixel i) / \sum_i (total number of tree stems in pixel i). The proportion of tree stems and the
961 proportion of tree basal area in each guild are highly correlated throughout the training data (Figure
962 S4). The figures cited in the main text for each guild were calculated using model projections
963 across all pixels, even those that did not meet the criteria for model projection because they fell
964 outside the multivariate distribution of the predictor layers or had insufficient stem density.
965 However, our estimates for the global % of trees occupied by each tree symbiotic guild change by
966 <1% when using only those pixels that met our criteria for model projection.

967 In the main manuscript we state that sharp transitions between dominant symbiotic states
968 with climate variables could lead to declines in EM trees, particularly in southern boreal forests.
969 To determine this, we projected our random forest models for each symbiotic guild using climate
970 change projections over our 19 bioclimatic variables (Table S7), including the decomposition
971 coefficients that use temperature and precipitation values. Specifically, we considered the 2070
972 scenario with a relative concentration pathway (RCP) of 8.5 (W/m²), which predicts an increase
973 of greenhouse gas emissions throughout the 21st century⁴⁵. We plot difference in the proportion of
974 forest basal area between the projections for 2070 and those using current climate data (Table S7,
975 Figure S24). We qualify this prediction with the note that vegetative changes to forests are
976 constrained by rates of mortality, recruitment, and growth.

977 After training and cross-validating our models with GFBi data exclusively, we additionally
978 tested whether our models accurately predicted the symbiotic state of Eurasian forests previously
979 published by Schepaschenko et al. (2017)⁴⁶. We assigned symbiotic status to all trees in
980 Schepaschenko et al. (2017) and aggregated plot level data to a 1 by 1 degree grid using the same

981 methods as with the GFBi dataset (Figure S25). We found that, on average, our models predicted
982 the symbiotic state in the regional dataset within 13.6% of the value of this other dataset (Figure
983 S26). For projected maps in Figure 4abc, we included the Schepaschenko et al. (2017) data with
984 the GFBi training data to increase geographic coverage throughout Eurasia.

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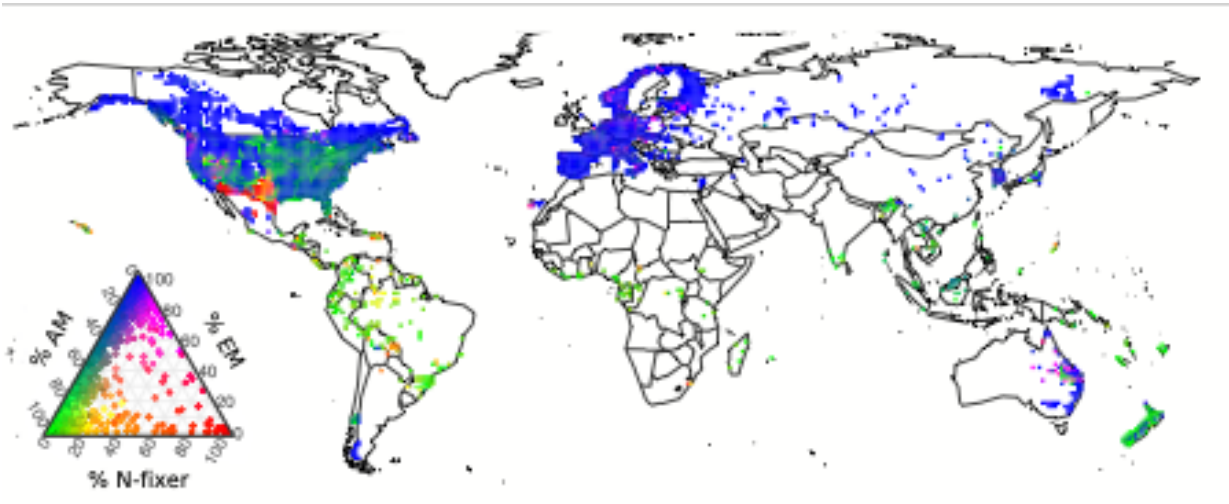
986 **Methods References**

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- 988 31 Liang, J. *et al.* Positive biodiversity-productivity relationship predominant in global
989 forests. *Science* **354**, aaf8957 (2016).
- 990 32 Chamberlain, S. A. & Szöcs, E. taxize: taxonomic search and retrieval in R.
991 *F1000Research* **2** (2013).
- 992 33 Brundrett, M. C. & Tedersoo, L. Evolutionary history of mycorrhizal symbioses and
993 global host plant diversity. *New Phytol.* (2018).
- 994 34 Brundrett, M. C. Mycorrhizal associations and other means of nutrition of vascular
995 plants: understanding the global diversity of host plants by resolving conflicting
996 information and developing reliable means of diagnosis. *Plant and Soil* **320**, 37-77,
997 doi:10.1007/s11104-008-9877-9 (2009).
- 998 35 Werner, G. D., Cornwell, W. K., Sprent, J. I., Kattge, J. & Kiers, E. T. A single
999 evolutionary innovation drives the deep evolution of symbiotic N₂-fixation in
1000 angiosperms. *Nature Communications* **5**, 4087 (2014).
- 1001 36 Werner, G. D., Cornwell, W. K., Cornelissen, J. H. & Kiers, E. T. Evolutionary signals of
1002 symbiotic persistence in the legume–rhizobia mutualism. *Proceedings of the National*
1003 *Academy of Sciences* **112**, 10262-10269 (2015).
- 1004 37 Afkhami, M. E. *et al.* Symbioses with nitrogen-fixing bacteria: nodulation and
1005 phylogenetic data across legume genera. *Ecology* **99**, 502-502 (2018).
- 1006 38 Tedersoo, L. *et al.* Global database of plants with root-symbiotic nitrogen fixation: Nod
1007 DB. *Journal of Vegetation Science* (2018).
- 1008 39 Hayward, J. & Hynson, N. A. New evidence of ectomycorrhizal fungi in the Hawaiian
1009 Islands associated with the endemic host *Pisonia sandwicensis* (Nyctaginaceae). *Fungal*
1010 *Ecol* **12**, 62-69 (2014).
- 1011 40 Lambers, H., Martinoia, E. & Renton, M. Plant adaptations to severely phosphorus-
1012 impoverished soils. *Current opinion in plant biology* **25**, 23-31 (2015).
- 1013 41 Wang, B. & Qiu, Y.-L. Phylogenetic distribution and evolution of mycorrhizas in land
1014 plants. *Mycorrhiza* **16**, 299-363 (2006).
- 1015 42 Soltis, D. E. *et al.* Chloroplast gene sequence data suggest a single origin of the
1016 predisposition for symbiotic nitrogen fixation in angiosperms. *Proceedings of the*
1017 *National Academy of Sciences* **92**, 2647-2651 (1995).
- 1018 43 Palosuo, T., Liski, J., Trofymow, J. & Titus, B. Litter decomposition affected by climate
1019 and litter quality—testing the Yasso model with litterbag data from the Canadian intersite
1020 decomposition experiment. *Ecological Modelling* **189**, 183-198 (2005).

1021 44 Bradford, M. A. *et al.* Climate fails to predict wood decomposition at regional scales.
1022 *Nature Climate Change* **4**, 625 (2014).
1023 45 Allen, M. R. *et al.* IPCC fifth assessment synthesis report-climate change 2014 synthesis
1024 report. (2014).
1025 46 Schepaschenko, D. *et al.* A dataset of forest biomass structure for Eurasia. *Scientific data*
1026 **4**, 170070 (2017).
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1031 Fig. 1. The global map has $n = 2,768$ grid cells at a resolution of $1^\circ \times 1^\circ$ latitude and longitude.

1032 Cells are coloured in the red, green and blue spectrum according to the percentage of total tree

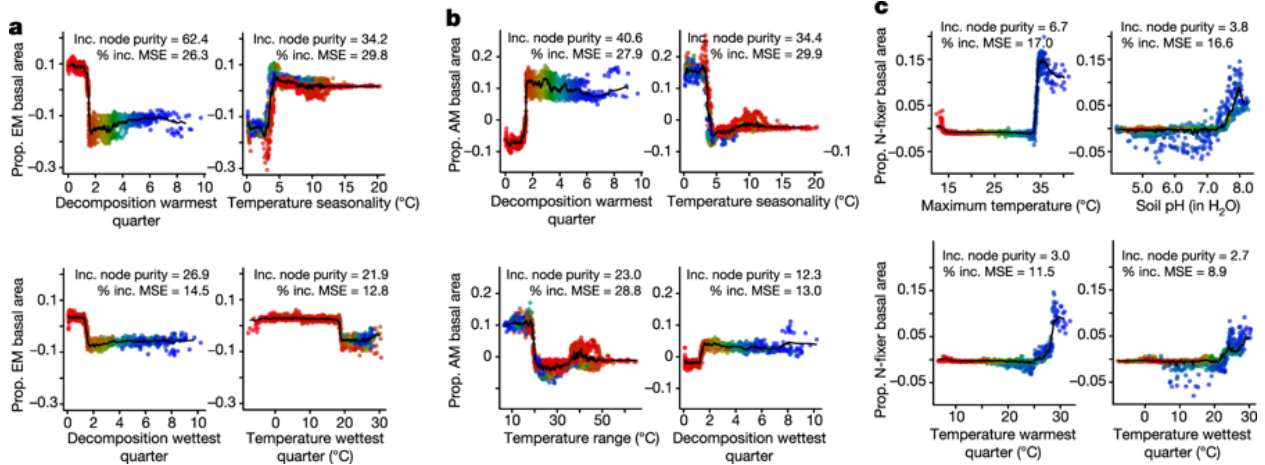
1033 basal area occupied by N-fixer, arbuscular mycorrhizal (AM) and ectomycorrhizal (EM) tree

1034 symbiotic guilds, as indicated by the ternary plot.

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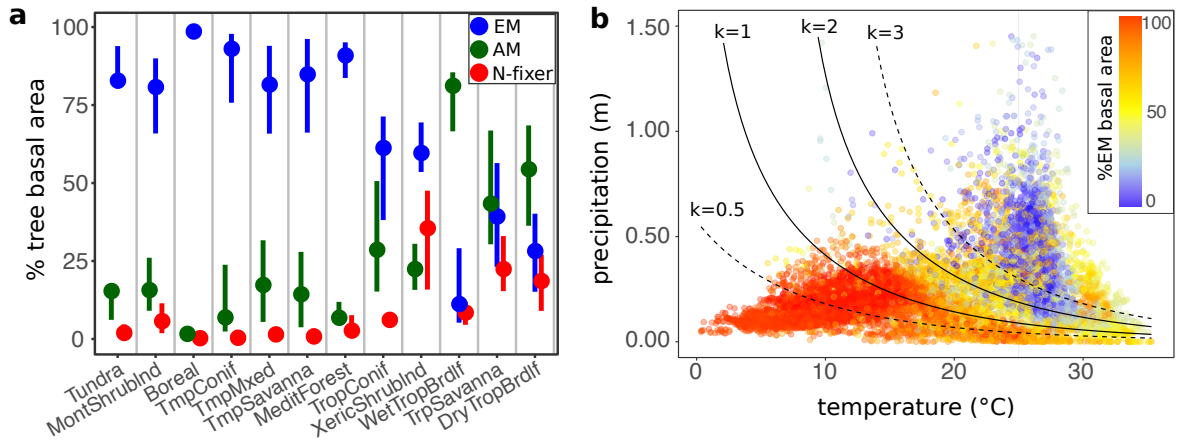
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Fig. 2. a–c, Partial feature contributions of different environmental variables to forest symbiotic state. Each row plots the shape of the contribution of the four most-important predictors of the proportion of tree basal area that belongs to the ectomycorrhizal (a), arbuscular mycorrhizal (b) and N-fixer (c) symbiotic guilds (n = 2,768). Variables are listed in declining importance from left to right, as determined by the increase in node purity (inc. node purity), and with points coloured with a red to green to blue gradient according to their position on the x axis of the most-important variable (left-most panels for each guild), allowing cross-visualization between predictors. Each panel lists two measures of variable importance; inc. node purity (used for sorting) and percentage increase in mean square error (% inc. MSE) (see Supplementary Information). The abundance of each type of symbiont transitions sharply along climatic gradients, which suggests that sites near the threshold are particularly vulnerable to switching their dominant symbiont guild as climate changes. Warmest and wettest quarter, the warmest and wettest quarters of the year, respectively.

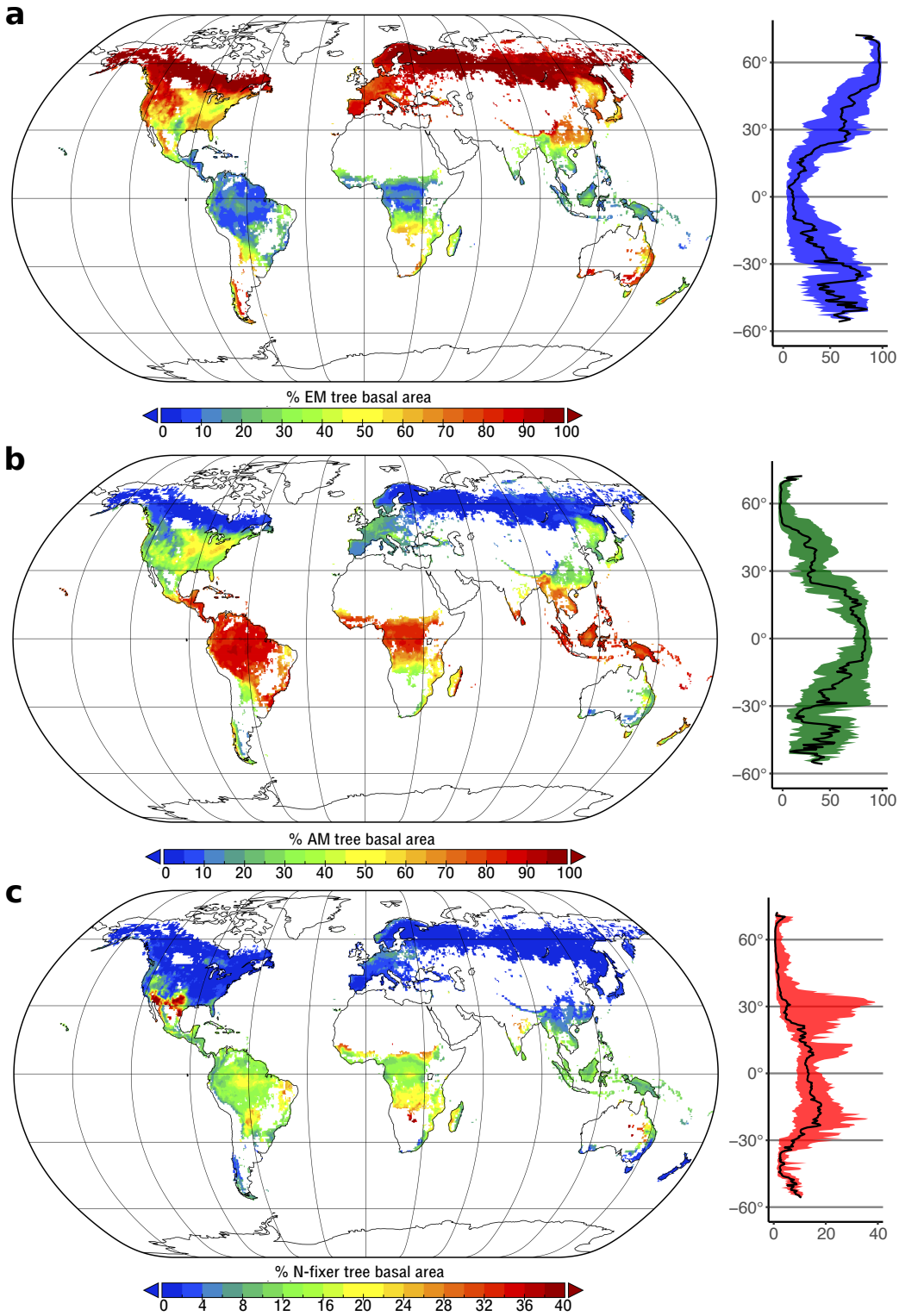
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1057 Fig. 3. a, Biome level summaries of the median \pm 1 quartile of the predicted percentage of tree
1058 basal area per biome for ectomycorrhizal, arbuscular mycorrhizal and N-fixer symbiotic guilds (n
1059 = 100 random samples per biome). b, The dependency of decomposition coefficients (k , solid and
1060 dotted lines; in the region between the solid lines, the model transitions abruptly between dominant
1061 symbiotic status) on temperature and precipitation during the warmest quarter with respect to
1062 predicted dominance of mycorrhizal symbiosis. The transition from arbuscular mycorrhizal forests
1063 to ectomycorrhizal forests between $k = 1$ and $k = 2$ is abrupt, which is consistent with positive
1064 feedback between climatic and biological controls of decomposition.

1065



1066

1067 Fig. 4. a–c, Maps (left) and latitudinal gradients (right; solid line indicates median; coloured ribbon

1068 spans the range between the 5% and 95% quantiles) of the percentage of tree basal area for

1069 ectomycorrhizal (a), arbuscular mycorrhizal (b) and N-fixer (c) symbiotic guilds. All projections
1070 are displayed on a 0.5°-by-0.5° latitude and longitude scale. n = 28,454 grid cells.