

1 **Fluctuating asymmetry, parasitism and reproductive fitness in two species of**
2 **gammarid crustacean**

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10 Running page head: Fluctuating asymmetry and parasitism in *Gammarus* sp.

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13

14 **Abstract**

15 Fluctuating asymmetry (FA), defined as random deviations from perfect bilateral symmetry, is assumed
16 to reflect developmental instability. FA is predicted to increase in response to environmental stress,
17 including parasite infection. In addition, theory predicts higher FA in sexually selected traits, due to their
18 greater sensitivity to stress. Here we investigate the relationships between FA, parasitism and
19 reproductive fitness in two species of gammarid crustacean, incorporating both sexual and non-sexual
20 traits. We found little evidence for a relationship between FA and fecundity in *Gammarus*; FA was not
21 correlated with brood size or size in females, or with sperm number in males. In contrast to prediction,
22 we report lower relative FA in response to sexual traits than non-sexual traits. However, FA in sexual
23 traits was found to be higher in males than females, supporting the theory that sexual selection leads
24 to increased FA. Additionally, we report a negative correlation between FA and both trematode
25 (*Podocotyle atomon*) and PCR positive microsporidian (*Nosema granulosis* and *Dictyocoela*
26 *duebenum*) infection and interpret these results in the context of the parasites' transmission strategies.

27

28 **1. Introduction**

29 Fluctuating asymmetry (FA) are random deviations from perfect bilateral symmetry (Van Valen, 1962)
30 and has long been used as an indicator of developmental stability; as it is thought to reflect the inability

31 of the genome to buffer developmental processes against stresses (Zakharov 1992). Hence, FA is often
32 used as a measure of genetic quality in empirical studies (Clarke 1995, Møller & Swaddle 1997), with
33 symmetry assumed to reflect high quality of individuals. However, the response of FAs to stresses
34 appears to be taxon and trait specific (Leung & Forbes 1996), leading to much debate over the
35 relationship between FA and fitness, with levels of FA decreasing with success in sexual competition
36 and fitness in some organisms and traits, but not others (Lens et al. 2002, Tomkins & Simmons 2003,
37 Dongen 2006). Initial meta-analyses reported an overall weak to moderate negative relationship
38 between FA and reproductive fitness (Møller & Thornhill 1998), but have proved controversial, with
39 inconsistency and publication bias in FA studies cited as a likely cause of discrepancies (Clarke 1998,
40 Palmer 1999, Palmer 2000). This has prompted the emergence of a number of publications that attempt
41 to set out guidelines for the appropriate conduction of studies of FA (Palmer & Strobeck 1986, Palmer
42 1994, Lens et al. 2002).

43 Secondary sexual traits are hypothesised to be particularly sensitive to environmental stress during
44 development and have higher mean FA for their size than morphological traits that are not under sexual
45 selection (Møller & Pomiankowski 1993, Watson & Thornhill 1994). Sexually selected traits are also
46 predicted to show a negative relationship between absolute FA and trait size. Essentially, individuals
47 with larger sexual traits should have lower levels of FA, because they are able to pay the cost of
48 expressing the larger trait, indicating that they are of high genetic quality and perhaps more sexually
49 attractive (Møller & Pomiankowski 1993).

50 Empirical support for increased FA in sexual traits in comparison to non-sexual traits is again mixed,
51 although few studies have directly compared FA in sexual and non-sexual traits in the same individuals
52 (Bjorksten et al. 2000). Literature surrounding the relationship between FA and measures of
53 reproductive investment is also inconsistent, with few studies having been conducted in invertebrates.
54 Woods et al. (2002) report no relationship between FA in non-sexual traits and egg number in
55 *Drosophila*. Similarly, in male yellow dung flies (*Scathophaga stercoraria*), no relationship was found
56 between leg FA and sperm size or testis size (Hosken et al. 2003). A negative association between limb
57 FA and ejaculate size was seen in male decorated field crickets (*Gryllodes sibilates*), but not in the moth
58 *Plodia interpunctella* (Gage 1998). These examples suggest that multiple sexual and non-sexual traits
59 may be developmentally linked in certain species but without correlation in others.

60 Multiple parasite groups, including ectoparasites and endoparasites, exert stress and metabolic costs
61 and cause physical changes to the tissues they infect. Due to the associated costs and potential
62 damage to host tissues, infection is predicted to lead to increased FA at all stages of development. The
63 time of infection in the animal's life stage (juvenile vs adult) is also of interest and may have specific
64 consequences for developmental alterations and specifically FA. This is supported by many empirical
65 studies reporting a positive relationship between prevalence and intensity of infection, and levels of FA
66 (Møller 1996). Alibert et al. (2002) report a positive association between FA in non-sexual traits and
67 infection by two acanthocephalan species in *Gammarus pulex*. There are several ways in which an
68 association between infection and FA can be viewed (Alibert et al. 2002). Firstly, the metabolic costs of
69 parasite infection may directly lead to developmental instability (Polak 1993). Alternatively, lower quality

70 individuals that experience greater developmental instability may simply be more susceptible to
71 infection (Møller 1996). A correlation between infection and developmental instability may occur due to
72 more asymmetrical individuals living in more stressful environments (Møller & Swaddle 1997). Further,
73 the duration of infection and the life stage of the host may also pose significant factors to the likelihood
74 of resultant FA (Møller 1996). Some parasites can initiate early infections in an animal's life cycle
75 through vertical transmission, such as infections caused by *Pleistophora mulleri* (Microsporidia) that
76 can be passed from mother to offspring and emerge at any stage in the development of the young
77 (Bunke et al. 2015).

78 As crustaceans, gammarids moult throughout their lives in order to grow, and it is not clear whether or
79 not levels of FA are conserved through successive moults in amphipods, with contrasting results in
80 other crustacean species. In the brachyuran crab *Hemigrapsus nudus*, sign and magnitude of FA were
81 found to persist throughout three successive moults, indicating that FA levels are determined early in
82 life (Chippindale & Palmer 1993). Conversely, in *Daphnia magna*, FA varies randomly in sign and
83 magnitude between moults, suggesting that FA reflects recent growth history and that developmental
84 instability may increase with age (Stige et al. 2006). If FA levels are not conserved across moults in
85 *Gammarus*, the age at which an individual acquires an infection may be less important than in other
86 systems. Whilst it might be expected that microsporidian parasites would have a greater impact on FA
87 due to their presence from the onset of development, this may be counteracted by the differing virulence
88 transmission trade-offs in horizontally and vertically transmitted parasites. As vertically transmitted
89 parasites rely on host reproduction for transmission, virulence levels are generally expected to be lower
90 (Bandi et al. 2001), which may reduce impact on FA levels.

91 Gammarid crustaceans are host to many parasite species (Bojko & Ovcharenko 2019) and are an ideal
92 study species in which to conduct a comprehensive investigation into the relationships between FA,
93 parasitism and reproductive fitness. Vertically transmitted parasites are common in the Amphipoda
94 (Bojko & Ovcharenko 2019) and can be present from the onset of host development. In contrast,
95 horizontally transmitted parasites can infect the host at any point throughout its life history, post-hatch.
96 Two such species that exhibit vertically and horizontally transmitted infections include *Gammarus*
97 *duebeni* and its microsporidian parasites, *Nosema granulosis* (vertical transmission) and *Dictyocoela*
98 *duebenum* (vertical transmission), and *Gammarus zaddachi* and its trematode parasite *Podocotyle*
99 *atomon* (horizontally transmitted). In *G. duebeni* microsporidian parasites are vertically transmitted from
100 the mother to offspring via the cytoplasm of her eggs (Terry et al. 1998). Male offspring are an
101 evolutionary dead-end for the parasite as sperm cannot pass on the parasite to the zygote (Dunn et al.
102 2001); hence it is adaptive for the parasites to feminise hosts. It may be possible that higher levels of
103 FA is present in infected females, as these will include feminised males, whose development has been
104 influenced by the parasite. Infection of *G. zaddachi*, an intermediate host for the trophically-transmitted
105 trematode *P. atomon*, has been found to have some impact on host fitness, fecundity and behaviour
106 (Arundell et al. unpublished).

107 In this study, we use two gammarid species, *G. duebeni* and *G. zaddachi* to investigate the relationships
108 between FA, parasitism, and reproductive fitness. To our knowledge, the full extent of sexual selection

109 has not been well studied in amphipods (Nahavandi et al. 2011). However, we incorporate sexual
110 characteristics, as well as more commonly used, morphological traits as a comparison. We test the
111 hypothesis that infection of gammarids by 1) vertically transmitted Microsporidia will result in higher
112 levels of FA than infection by 2) horizontally transmitted trematodes, due to their increased opportunity
113 to influence development. In *G. duebeni*, we investigate the relationship between FA in non-sexual vs.
114 sexual traits, and measures of fecundity, including sperm numbers in males and egg numbers and
115 diameter in females. We also assess the impact of infection by feminising microsporidian parasites on
116 FA. In *G. zaddachi*, we examine the relationship between FA in the same (sexual and non-sexual) traits
117 and infection with *P. atomon*, in both males and females. In contrast to microsporidian infection of *G.*
118 *duebeni*, which is present in the embryo, *P. atomon* prevalence and burden increase with age in *G.*
119 *zaddachi*, providing a system to explore how burden and age of infection may influence FA. Overall,
120 this study provides a comparison of the impacts of vertically transmitted and horizontally transmitted
121 parasites on FA levels in *Gammarus*.

122

123 **2. Materials and Methods**

124 *2.1 Animal collection and husbandry*

125 *Gammarus duebeni* and *G. zaddachi* were collected from Budle Bay, Northumberland, U.K. (55°40'N,
126 1°43'W) during October-November 2010 using a fine mesh net (1mm). Gammarids in both stock tanks
127 and experimental pots were maintained in aerated brackish water (15 ppt) at 14°C, 16h light: 8h dark,
128 with rotted sycamore (*Acer pseudoplatanus*) leaves and algae (*Enteromorpha* spp.) provided for food
129 and shelter. Both the water and food supplies were replaced daily, to ensure the welfare of the animals.
130 Any gammarids remaining at the end of the study were either maintained in stock tanks or returned to
131 the field.

132

133 *2.2 Experimental design*

134 *2.2.1 Fluctuating asymmetry and fecundity in Gammarus duebeni infected with Microsporidia*

135 To investigate the relationship between FA and fecundity in *G. duebeni*, 30 males and 30 gravid females
136 were dissected for measurement. Males were isolated in 200ml plastic pots for seven weeks, to allow
137 maximisation of sperm stores before being anaesthetised in carbonated water and dissected for FA
138 measurement (Fig. 1; Table 1) and sperm counts. The FA measurements were made by the same
139 person using a microscope and graticule throughout the study. To obtain females with eggs in early
140 stages of embryogenesis (stage 1) eggs, 30 pre-copulatory pairs of *G. duebeni* were isolated, inspected
141 twice daily and separated post-copulation. Gravid females were anaesthetised to allow the new eggs
142 to be flushed from the brood pouch, using a fine jet of water from a hypodermic needle and syringe.
143 The eggs were then counted and the diameter of 10 eggs from each female were measured under an
144 Olympus BH-2 compound microscope. All eggs were then stained with DAPI (4', 6-diamidino-2-

145 phenylindole, diluted 1:500 in 0.2M NaH₂PO₄; a fluorescent stain for DNA) using methods from Kelly et
146 al. (2003), and screened for microsporidian infection to explore relationships between infection, FA and
147 fecundity in females (Fig. 2a).

148

149 2.2.2 The relationship between FA and trematode infection in *Gammarus zaddachi*

150 To investigate the relationship between FA and trematode infection in *G. zaddachi*, 60 males and 60
151 females were anaesthetised and dissected for FA measurements to be taken. Infected/uninfected
152 individuals were identified by examination for the presence of parasite cysts below the cuticle, using a
153 dissecting microscope (Leica S6D) (Fig. 2b). Visual identification of infection was deemed acceptable
154 as it had previously been shown that 84.2% of animals with one or more visible cysts were PCR-positive
155 for *P. atomon* infection and only one of 58 individuals with no visible cysts was PCR-positive (1.7%) (K.
156 Arundell et al. unpublished data).

157

158 2.3 Dissection and FA measurements

159 All individuals were anaesthetised, gently blotted dry on tissue paper and weighed. They were placed
160 in a watchglass containing a few drops of Van Harrefeld crustacean saline and observed under a
161 dissecting microscope. Watchmaker's forceps and spring loaded scissors were used to decapitate the
162 gammarid, by making a cut between pereons 1 and 2 and coxal plates 1 and 2 (all nomenclature for
163 *Gammarus* spp. body plan used according to Gledhill et al. (1993). The telson was removed by cutting
164 between pereon 7 and epimera 1 and discarded. The pereon and pleon were transferred to Eppendorf
165 tubes and stored in 70% EtOH at -20°C until required for measurements to be taken.

166 A range of measurements were selected, to include both sexual and non-sexual, as well as metrical
167 and meristic traits (Table 1). All measurements were taken at each side of the individual. Non-sexual
168 traits were selected based upon those in which FA had been found to correlate with acanthocephalan
169 infection in a previous study of FA in *G. pulex* (Alibert et al. 2002). Additional sexual characteristics
170 were also measured to look for a relationship between FA in sexual traits and fecundity. In males,
171 measurements were taken from the genital papillae, which are used to thrust sperm bundles into the
172 brood pouch of the female during mating and the second pereopods. The genital papillae are sexually
173 dimorphic; in males these are enlarged into gnathopods and used to position and hold the female during
174 copulation (Hume et al. 2005). The number of calceoli on antenna 2 were also counted in male *G.*
175 *duebeni* because they are thought to be required for accurate assessment of female quality (Dunn
176 1998). Measurements were taken from the female's oostegites, which are specialised structures used
177 to form the ventral brood pouch.

178 All measurements were made at 40X or 100X magnification as appropriate, under an Olympus BH-2
179 compound microscope. Watchmaker's forceps and fine hypodermic needles were used to dissect and
180 manipulate the tissues as required for measurement. To aid visualisation of the genital papillae, the

181 samples were dehydrated in 500µl 100% EtOH for two hours. The EtOH was then removed and
182 replaced with 500µl cedar wood oil to clear the sample. After another two hours, the tissue was removed
183 from the cedar wood oil and transferred to a microscope slide and viewed at 100X magnification for
184 measurements to be taken.

185

186 *2.4 Sperm count data collection*

187 Male *G. duebeni* were dissected for sperm counts as described in Arundell et al. (2014). Briefly, the
188 testes were dissected from the body cavity using spring-loaded scissors, and watchmaker's forceps.
189 The ventral portion of the body was stored with the head in 70% ethanol at -20°C until required for FA
190 measurements. The testes were transferred to 10µl of distilled water on a cavity slide and ruptured
191 using a fine hypodermic needle. The sample was then washed into an Eppendorf tube using a Gilson
192 pipette and diluted to a final volume of 1ml. The tube was gently vortexed in order to evenly distribute
193 the sperm, before three 10 µl aliquots were pipetted onto a microscope slide and allowed to air-dry. The
194 slides were viewed using an Olympus BH-2 microscope at 40X magnification and the sperm were
195 counted and summed together. Counts from the 3 aliquots were averaged and multiplied by the dilution
196 factor (100X) to give an estimate of the total number of sperm for each male.

197 Similar measurements could not be made for *G. zaddachi* because this species did not pair well in
198 laboratory conditions and so reproductive measurements would have been unlikely to be
199 representative.

200

201 *2.5 Microsporidia screening in amphipod hosts*

202 To determine infection status of the mother, eggs were screened for microsporidian parasites (Kelly et
203 al. 2001, Weedall et al. 2006). Briefly, eggs from each female were collected in separate Eppendorf
204 tubes. A Pasteur pipette was used to add 1ml of 5M HCl to each tube, until the membranes had
205 dissolved – evident due to a colour change from dark grey to bright orange. At this point, the HCl was
206 removed and the eggs washed with distilled water, before being stored in 1ml acetone at -20°C until
207 required for DAPI screening (Fig. 2a).

208 When required for visualisation, eggs were transferred to a microscope slide and left to allow excess
209 acetone to evaporate. A cover slip was placed over the eggs and gentle pressure applied, to spread
210 out the cells of the eggs without destroying them. A Gilson pipette was used to deliver 150µl of DAPI
211 quick mounting solution, which fluoresces in the presence of DNA, to the edge of the cover slide, so
212 that it then covered the eggs by capillary action. Any excess DAPI solution was removed with tissue
213 paper and the edges of the coverslip sealed with clear nail varnish. Slides were stored at 4°C in the
214 dark and visualised as soon as possible. The slides were viewed at 200X magnification under a Zeiss
215 Axiovert S100 microscope, using a mercury light source and appropriate filters for visualising the DAPI
216 staining (exciter filter BP 450-490; chromatin beam splitter FT510; barrier filter LP520). Infection by *N.*

217 *granulosis* or *D. duebenum* was determined by the presence of parasite DNA in the cytoplasm around
218 the host nucleus.

219

220 *2.6 Data Analysis and model selection*

221 Unless otherwise stated, all data were analysed using statistical models constructed in R version 3.1.1
222 (R Development Core Team 2014). All models were initially constructed as maximal models, including
223 all relevant terms and interactions. Models were compared using P-values from the “dropterm” function
224 (MASS library; Venables & Ripley 2002) to determine whether terms significantly improved the fit of the
225 model. Those that didn’t were removed in a stepwise fashion until only terms that improved the fit of the
226 model at $P < 0.05$ remained.

227

228 *2.6.1 Testing for directional asymmetry, antisymmetry and measurement error*

229 When testing FA, it is first important to rule out two other forms of symmetry: directional asymmetry
230 (DA) and antisymmetry (AS). These both occur when one side of a bilateral trait is always larger than
231 the other; in DA one specific side (left or right) is consistently the largest, whereas in AS, the side that
232 is larger varies randomly among individuals (Van Valen 1962). Additionally, due to the often small nature
233 of measurements taken, it is important to ensure that measurement error (ME) is not outside acceptable
234 limits, specifically that differences in between-sides measurements are significantly larger than
235 differences in repeated measurements (Palmer & Strobeck 1986). To test for ME, repeated
236 measurements were carried out on all metrical traits for a subset of 60 individuals. Repeated measures
237 were carried out three times, over the course of three days, without reference to the previous
238 measurements, following the method of Palmer (1994). Significant errors in recording meristic data
239 were ruled out by repeat counting of the first sample for each trait. DA and AS were assessed across
240 the entire data set. Whilst expected distributions for meristic data are less clear than for metrical data,
241 because the meristic traits selected generally differed by relatively large amounts, e.g. 4 or 5 (Palmer
242 1994), it was deemed adequate to assume they would approximate metrical traits. All traits were
243 subjected to the same analyses by the same person (KLA) for the rest of preliminary testing.

244 For each trait in each data group, DA was tested for using one-sample t-tests to ensure that the mean
245 of the signed asymmetries did not differ significantly from 0 (Palmer 1994, Alibert et al. 2002). To test
246 for AS, we used IBM SPSS Statistics 20 (IBM, Armonk, NY, USA) to check for any departures from
247 normality, with Shapiro-Wilk and Kolmogorov-Smirnov tests, as well as outputting skewness and
248 kurtosis estimates (Palmer 1994). Sequential Bonferroni correction was used to correct for multiple
249 tests (Rice 1989). Measurement error was evaluated using a linear mixed effects model (Pinheiro et al.
250 2013) for each trait, including side as a fixed factor and individual as a random factor. A significant
251 interaction term between side and individual indicated that measurement error was sufficiently minimal
252 to proceed with FA analysis (Palmer 1994).

253 We checked for any size-dependence of FA using three methods (Alibert et al. 2002, Palmer 1994). 1)
254 To test for within-samples size-dependence, we ran multiple linear models (LMs) of right and left sizes
255 ($|R-L|$) against both weight of the individual and trait size ($(R+L)/2$), using sequential Bonferroni correction
256 when interpreting the outputs. 2) To test for among-samples size-dependence, LMs of $\log(\text{var}(R-L))$ vs.
257 $\text{mean}((R+L)/2)$ across metrical and meristic traits were constructed. 3) As the tests detected evidence
258 of among-samples size dependence, two different indices of FA were calculated for all further analysis
259 (see Section 5.4.1), FA3 which accounts for mean trait size, and FA1 that does not (Palmer 1994):

260 $FA1 = |R-L|$

261 $FA3 = |R-L|/\text{mean}(R+L)$

262 Where R = right measurement and L = left measurement

263

264 2.6.2 *Fluctuating asymmetry and fecundity analysis for Gammarus duebeni and microsporidian* 265 *infection*

266 To investigate any relationship between FA and egg number or egg diameter, general linear models
267 (LMs) were constructed, including the measure of FA, female weight and infection status as predictor
268 variables. For the egg number model, egg diameter was also included as a predictor variable, to look
269 for a potential trade-off between egg number and egg size. To look for an association between FA and
270 microsporidian infection in females, a generalised linear model (GLM), with binomial error structure, of
271 infection status against FA index was used, also including female weight as a potential predictor
272 variable.

273

274 2.6.3 *Fluctuating asymmetry analysis for Gammarus zaddachi and trematode infection*

275 To test for a parasitic association between the trematode *P. atomon* and FA in *G. zaddachi*, GLMs, with
276 binomial error structure, of infection status against FA index were constructed separately for males and
277 females. Both models also included individual weight as a potential explanatory variable.

278

279 2.6.4 *Differences in FA between traits*

280 To test for differences in FA between the various traits measured for this study a GLM of FA3 for all
281 data was used, with quasi-Poisson error structure, as data were non-normal and over-dispersed. FA3
282 was used as this index is corrected for trait size, thus enabling us to look for effects across the entire
283 data set, without bias due to differences in size between *G. dubeni* and *G. zaddachi*, or between males
284 and females. Species, sex, data type (metrical vs. meristic) and trait type (sexual vs. non-sexual) were
285 included as factors.

286

287 3. Results

288 3.1 Exploratory data analyses

289 The mixed model (side x individual) for the replicate measurements showed that the non-directional
290 asymmetry variances (interaction variances from model outputs) were significantly larger than
291 measurement error, for all traits in the subset of 60 individuals tested ($P < 0.001$ in each case). The mean
292 variance due to measurement error ranged from 1.0% to 10.2% of the total between-sides variance and
293 it was therefore deemed acceptable to take just one measurement for the rest of the individuals (Palmer
294 1994). Additionally, it was concluded that these values were sufficiently low to avoid significant bias in
295 subsequent statistical analyses.

296 We found no evidence for directional asymmetry (DA) in any of the specimens analysed with one-
297 sample t-tests found no significant departures from a mean of 0 across all 40 signed asymmetry
298 distributions tested.

299 Kolmogorov-Smirnov tests detected significant departures from normality in 1 of 24 metrical traits and
300 5 of 16 meristic traits, whilst Shapiro-Wilk tests detected significant departures from normality in 3 of 24
301 metrical and 2 of 16 meristic traits. Skewness and kurtosis were significant for 3 of 24 metrical and 4 of
302 16 metrical traits. The same distributions were often significant for more than one of the tests, such that
303 overall, 9 distributions (3 metric and 6 meristic) differed significantly from a normal distribution. However,
304 strong antisymmetry (AS) was ruled out as inspection of histograms for these 9 distributions revealed
305 that none were platykurtic. Due to the low number of significant results and because departures from
306 normality were not concentrated across any particular trait or sample, it was concluded that the samples
307 were suitable for FA analysis.

308 We found little evidence for any within-samples size-dependence of FA, with LMs for both measures of
309 size (individual weight and trait size) across all metrical traits in all samples (48 tests in total) with P -
310 values > 0.05 (following Bonferroni correction). Two of the 32 tests for meristic traits in all samples did
311 indicate significant dependence of FA on trait size - uninfected male *G. zaddachi* count of spines on
312 pereopod 6 PER ($P < 0.05$) and count of flagellum segments on antenna 2 ($P < 0.01$) – yet not on
313 individual weight. As type 1 error is a common hazard in multiple testing, we did not deem this to be
314 significant evidence to justify correcting for within-samples size-dependence. However, we did find
315 strong evidence for among-samples size-dependence across both metrical and meristic traits ($P < 0.001$
316 in both cases). Hence, we accounted for this size-dependence by calculating index FA3, defined as $|R-$
317 $L|/\text{mean}(R+L)$ (Palmer 1994). Palmer and Strobeck (1986) warn that this correction may mask genuine
318 associations with FA, if the factor under investigation is also correlated with size. In this study, it is likely
319 that measures of reproductive potential (egg numbers, and sperm numbers etc.), as well as parasite
320 infection (vertically or horizontally transmitted), may be associated with individual size. Therefore we
321 also used index FA1, defined as $|R-L|$ (Palmer 1994), for comparison in all analyses. Additionally, to
322 look for differences in FA among trait-types, we used subsets of the data to calculate mean FA3 for
323 metrical and meristic, and sexual and non-sexual traits, separately. Number of oostegites in females

324 was found to be exactly equal for the left and right sides in all individuals examined; therefore, this trait
325 was excluded from the analyses.

326

327 3.2 *Fluctuating asymmetry and fecundity in Gammarus duebeni infected with Microsporidia*

328 We found no relationship between FA and brood size in *G. duebeni* females, using either index of FA;
329 e.g. FA1 for all traits ($F_{1,27}=0.027$, $P=0.871$), nor FA3 for all traits ($F_{1,27}=0.075$, $P=0.787$). Egg number
330 was only significantly affected by female weight, with larger females producing larger brood sizes
331 ($F_{1,28}=16.888$, $P<0.001$, Adjusted $R^2=0.35$, Fig. 3a). We found no evidence for a trade-off between egg
332 number and size, with egg diameter having no effect on brood size ($F_{1,27}=0.623$, $P=0.437$). Additionally,
333 we found no effect of microsporidian infection on brood size ($F_{1,27}=0.104$, $P=0.749$).

334 By contrast, we found marginal evidence for a relationship between egg diameter and fluctuating
335 asymmetry, with FA1 for all traits ($F_{1,27}=3.179$, $P=0.086$, Fig. 3b), FA1 for metrical traits ($F_{1,27}=3.742$,
336 $P=0.064$) and FA3 for metrical traits ($F_{1,27}=3.431$, $P=0.075$) all showing non-significant trends for a
337 decrease in egg diameter with increased FA. Additionally, egg diameter was significantly reduced by
338 9.8% in infected individuals (mean \pm S.E. $591.4 \pm 8.8\mu\text{m}$) in comparison to uninfected individuals (mean
339 \pm S.E. $655.2 \pm 11.5\mu\text{m}$) ($F_{1,28}=6.015$, $P=0.021$, Fig. 4a). However, we found no relationship between
340 female weight and egg size ($F_{1,27}=0.001$, $P=0.979$).

341 Of the 30 *G. duebeni* females used in the study, 7 (23% prevalence) were found to be infected with
342 Microsporidia. Due to the low sample size for infected females, individuals infected with *N. granulosis*
343 and *D. duebenum* were combined for statistical analysis purposes. The binomial GLM found a
344 significant relationship between infection status and FA3 for all traits ($\text{LRT}_1=10.082$, $P=0.001$, Fig. 4b),
345 yet not between infection status and FA1 for all traits ($\text{LRT}_1=0.896$, $P=0.343$), indicating that the among-
346 samples size-dependence correction in FA3 is enabling detection of an association that is not apparent
347 from FA1 alone. This highlights the importance of selecting the appropriate index for use in studies of
348 FA. Infection status was not associated with individual weight ($\text{LRT}_1=0.438$, $P=0.508$).

349 In male *G. duebeni*, we found no relationship between sperm number and either index of FA; e.g. FA1
350 for all traits ($F_{1,27}=0.557$, $P=0.470$), and FA3 for all traits ($F_{1,27}=0.135$, $P=0.716$). However, sperm number
351 was significantly associated with male weight, with increased sperm numbers in larger males
352 ($F_{1,28}=87.081$, $P<0.001$; Fig. 5).

353

354 3.3 *The Relationship Between FA and Trematode Infection in Gammarus zaddachi*

355 We found no relationship between female *G. zaddachi* trematode infection and FA1 ($\text{LRT}_1=0.311$,
356 $P=0.577$) or FA3 ($\text{LRT}_1=0.890$, $P=0.346$), for all traits. However, when looking at mean FA for non-
357 sexual traits only, we found a significant association between infection and FA1, with infected individuals
358 exhibiting higher levels of FA in non-sexual traits than uninfected individuals ($\text{LRT}_1=8.752$, $P=0.003$).
359 This relationship was not significant between infection and FA3 for non-sexual traits ($\text{LRT}_1=1.131$,

360 P=0.288). However, infection status in female *G. zaddachi* was strongly correlated with weight
361 (LRT₁=10.398, P=0.001) and hence size of individual. Infected females (mean ± S.E. 16.2 ± 0.9mg)
362 were on average 21.6% heavier than uninfected females (mean ± S.E. 12.7 ± 0.5mg). It is highlighted
363 by Palmer (1994) that if the factor under investigation is also correlated with size, then the size-
364 dependence correction in the FA3 index may mask genuine associations with FA (Palmer & Strobeck
365 1986). The observed association between trematode infection and FA1 in non-sexual traits is likely to
366 be real (Fig. 6a).

367 Similarly, in male *G. zaddachi*, we found a significant relationship between *P. atomon* infection and FA1
368 for all traits (LRT₁=22.756, P<0.001, Fig. 6b), but not between infection and FA3 for all traits
369 (LRT₁=1.668, P=0.197). Again, we found a significant association between infection status and weight
370 (LRT₁=4.065, P=0.044), with infected males (mean ± S.E. 36.1 ± 1.4mg) weighing on average 12.2%
371 more than uninfected males (mean ± S.E. 31.7 ± 2.1mg).

372

373 3.4 Differences Between Traits

374 The GLM for FA3 across the whole data set, found a significant effect of the interaction between trait
375 type and sex on FA3 (F₁=17.466, P<0.001). Non-sexual traits showed higher levels of FA3 than sexual
376 traits in both males and females. FA3 in non-sexual traits did not differ between sexes, whereas FA3 in
377 sexual traits was significantly higher in males than females (Fig. 7). Additionally, we found a significant
378 effect of data type on FA3, with meristic traits (mean ± S.E. 0.0540 ± 0.0024) exhibiting higher levels of
379 FA3 than metrical traits (mean ± S.E. 0.0267 ± 0.0010) (F₁=109.280, P<0.001). Species did not
380 significantly affect levels of FA3; hence *G. duebeni* and *G. zaddachi* did not differ in FA levels (F₁=2.57,
381 P =0.11).

382

383 4. Discussion

384 This study explores the relationship between microsporidian and trematode infections on the body
385 symmetry of amphipod hosts. We discuss these findings in reference to healthy host FA rates, the
386 effects of parasitism, the development and pathology caused by these parasites and conclude on the
387 various areas in which our data could be advanced in other species and with other parasite systems.

388

389 4.1 Host traits and fluctuating asymmetry

390 We found little evidence for any relationship between FA and measures of fecundity in *Gammarus* sp.
391 without disease; with no relationship between any measures of FA and brood size (egg number) in
392 females, or sperm number in males, when not accounting for parasitism. However, we observed a trend
393 for females with higher levels of FA to produce smaller eggs; however, the linking mechanism is
394 unknown. These findings are consistent with the literature, which finds contradictory results regarding

395 the relationship between FA and reproductive fitness, with FA correlated with reproductive fitness and
396 success in some organisms and traits, but not in others (Clarke 1995; Møller & Swaddle 1997; Lens et
397 al. 2002; Dongen 2006). For example, no relationship between leg FA and reproductive fitness or
398 longevity was found in female yellow dung flies (Martin & Hosken 2002). Additionally in male yellow
399 dung flies, no association has been found between leg FA and sperm size or testis size (Hosken et al.,
400 2003). A negative association between limb FA and ejaculate size is seen in male decorated field
401 crickets (*Gryllodes sigillatus*), but not in the moth *Plodia interpunctella* (Gage 1998). Hence relationships
402 between FA and measures of reproductive fitness could be specific to the species examined and include
403 multi-factorial reasons for resulting in FA. In amphipods, only one other study exists to examine FA
404 (Alibert et al. 2002); however, this study provided specific measurements of traits, excluding sperm
405 count, egg size and other measurements of fecundity.

406

407 4.2 Parasitism and fluctuating asymmetry in amphipods

408 To date, the only other study of FA in amphipods investigates the impact of acanthocephalan infection
409 on FA levels in *Gammarus pulex* (Alibert et al. 2002). Levels of FA1 found in the present study were
410 comparable to those observed in *G. pulex* despite differences in the species and range of traits
411 measured. However, their study did not use FA3 and so comparisons between size-corrected measures
412 of FA cannot be drawn. In accord with Alibert et al. (2002), we found compelling evidence for a link
413 between FA and parasitism in gammarids, with increased FA in Microsporidia infected female *G.*
414 *duebeni*, and in both male and female *G. zaddachi* infected by the trematode *P. atomon*. This is
415 consistent with the theory that FA reflects developmental instability. Known microsporidian parasites of
416 *G. duebeni* are vertically transmitted, passing from mother to offspring via the cytoplasm of the egg
417 (Terry et al. 1998). Hence individuals are infected from onset of development, meaning that the
418 increased levels of FA are likely to reflect a reduction in developmental stability caused by infection.
419 This seems particularly plausible as both *N. granulosis* and *D. duebenum* are feminisers (Weedall et al.
420 2006), converting genetic male offspring into functional phenotypic females, in order to avoid the
421 evolutionary dead end of infecting a male. Sperm are comparatively tiny and rarely contribute to the
422 cytoplasm of the zygote, hence they do not transmit cytoplasmic parasites (Dunn & Smith 2001).
423 Therefore, many phenotypic females will have started life as genotypic males and it is not difficult to
424 imagine that these conflicting influences may lead to significant developmental instability (Weedall et
425 al. 2006).

426 The metabolic costs of microsporidian infection may also impact on female reproductive success by
427 reducing investment in reproduction. We found a non-significant trend for a reduction in egg diameter
428 in microsporidia infected female *G. duebeni*. However, the sample size for infected females was
429 relatively small (7 of 30 females). Hence this area of research may warrant further study. It is likely that
430 larger eggs are advantageous to reproductive success as they enable provision of more resources for
431 the developing eggs. For example, female *Gammarus* produce fewer, larger eggs in winter, when
432 conditions are less suitable for reproduction (Dunn & McCabe 1995; Sheader 1996). Additionally, trade-

433 offs between egg size and number are stronger in smaller *Gammarus minus* females. Hence larger
434 females, for which space constraints are less of an issue, maintain egg size with an increase in number,
435 again indicating that larger egg size is preferable (Glazier 2000). In line with previous studies (Terry et
436 al. 1998; Kelly et al. 2003). We found no evidence for an effect of microsporidian infection on egg
437 number or size; however, this could also be due to the limited sample size. The same association was
438 made by Bojko et al. (2018) in *Dikerogammarus haemobaphes* infected with *Cucumispora ornata*,
439 where the sample size was in the hundreds of individuals examined.

440 In contrast to microsporidian infection, *P. atomon* infection increases in prevalence and intensity in
441 larger and hence older gammarids in wild populations; most likely due to increased exposure over time.
442 Therefore, it might be expected that the increased FA in parasitised individuals reflects an increased
443 susceptibility to infection in individuals that experienced higher levels of developmental stability.
444 Crustaceans are particularly interesting for studies of asymmetry because they moult throughout life in
445 order to grow larger. The degree to which FA is conserved throughout successive moults is variable
446 between crustacean species. For example, Chippindale and Palmer (1993) report a persistence in both
447 sign and magnitude of FA throughout three successive moults in the brachyuran crab *Hemigrapsus*
448 *nudus*, indicating that FA levels are determined early in life. Conversely, in *Daphnia magna*, FA was
449 found to vary randomly in sign and magnitude between moults, suggesting that FA reflects recent
450 growth history and that developmental instability may increase with age (Stige et al. 2006). It is not
451 known whether symmetry is conserved across moults for gammarid species and we highlight this as an
452 interesting area for future research. If levels of FA fluctuate between moults, then trematode infection
453 has the potential to increase FA over an individual's life span.

454

455 4.3 Concluding remarks and future considerations

456 In this study, parasitism has been shown to have specific negative impacts on naturally symmetrical
457 traits in amphipods, for both vertically transmitted Microsporidia and horizontally transmitted trematode
458 parasites acquired later in development. Our data suggest that FA can be influenced by parasitism;
459 however, fecundity does not seem to be impacted significantly. This may reflect the parasites need to
460 transmit and utilise the host as well as maintain the host population density and size for continued
461 transmission. For vertically transmitted Microsporidia, their primary method involves a breeding host,
462 therefore the parasite may be evolutionary predisposed to avoid impairing the breeding capability of the
463 host to benefit its own transmission.

464 Our data reveal that parasitism impacts traits linked with asymmetry, including those traits directly
465 involved in reproduction as well as additional morphological traits. Novel associations with disease in
466 amphipods, including viral, bacterial and other infections (Bojko & Ovcharenko 2019), have paved the
467 way to conduct further assessment of asymmetry linked with parasitism and the additional discovery of
468 diseases in amphipods and their study as model systems to explore FA could provide intriguing

469 relationships between specific parasite taxa and the likelihood for parasite-associated host
470 developmental alterations.

471

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476

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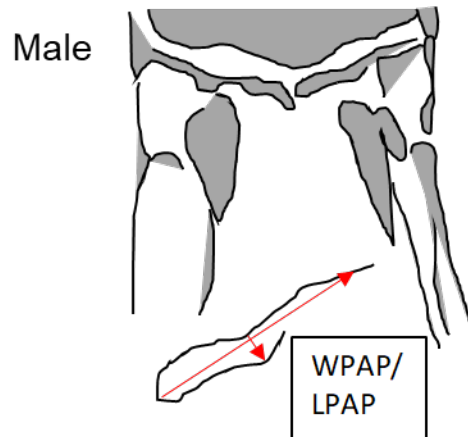
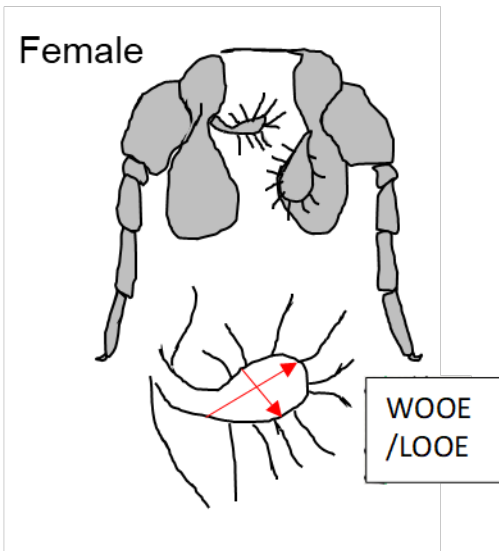
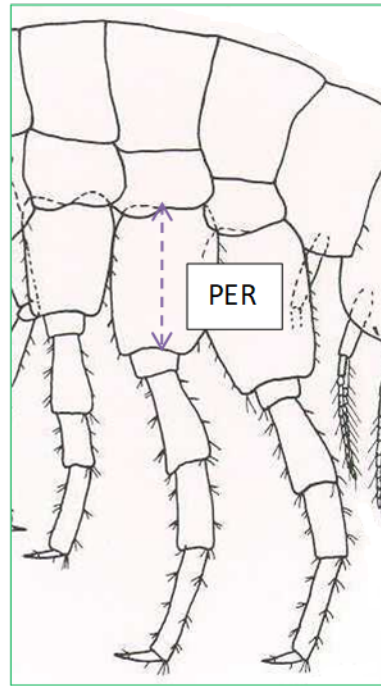
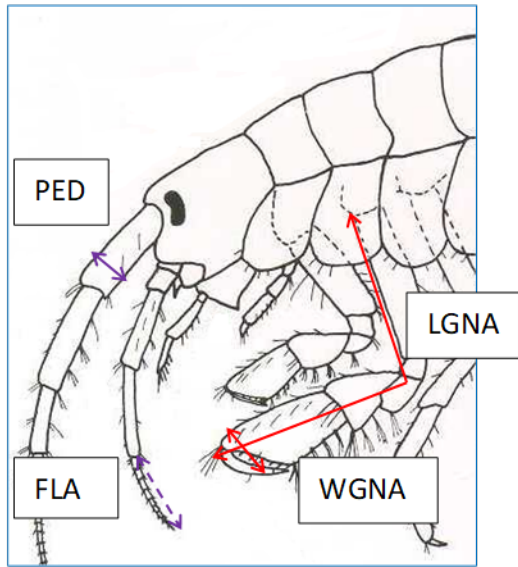
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578 **Figures and Tables**

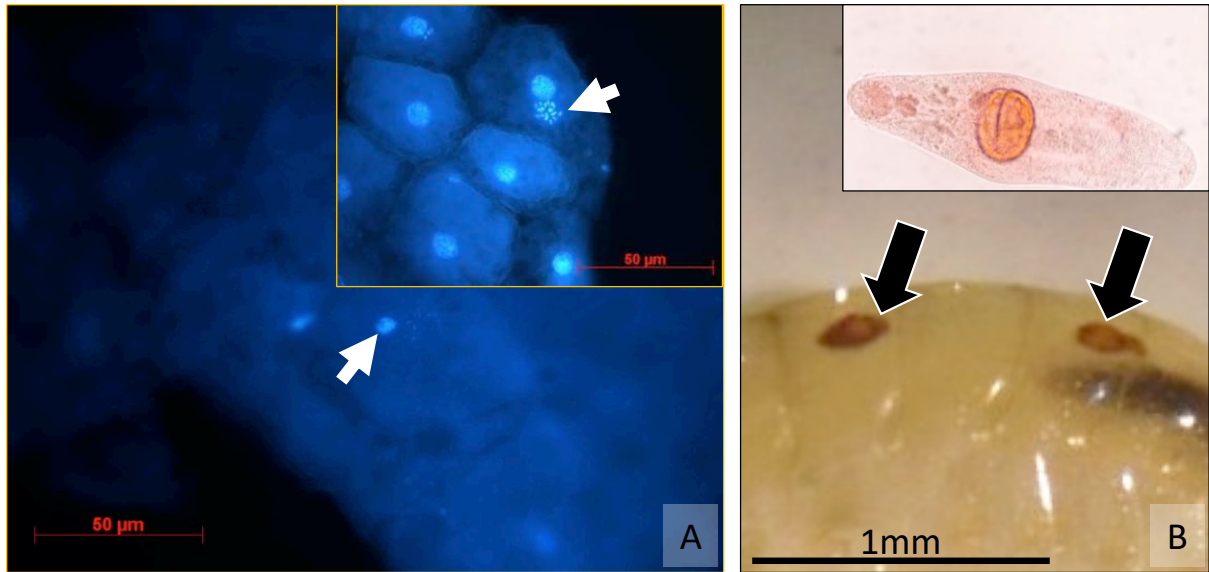
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580

581 Figure 1. Measurements for FA Calculations; Solid arrows indicate metrical measurements and dashed
 582 arrows indicate regions from which meristic counts were taken; Purple arrows indicate non-sexual and
 583 red arrows sexual traits. Diagrams of male and female sexual appendages are reproduced from Gledhill
 584 et al. (1993).

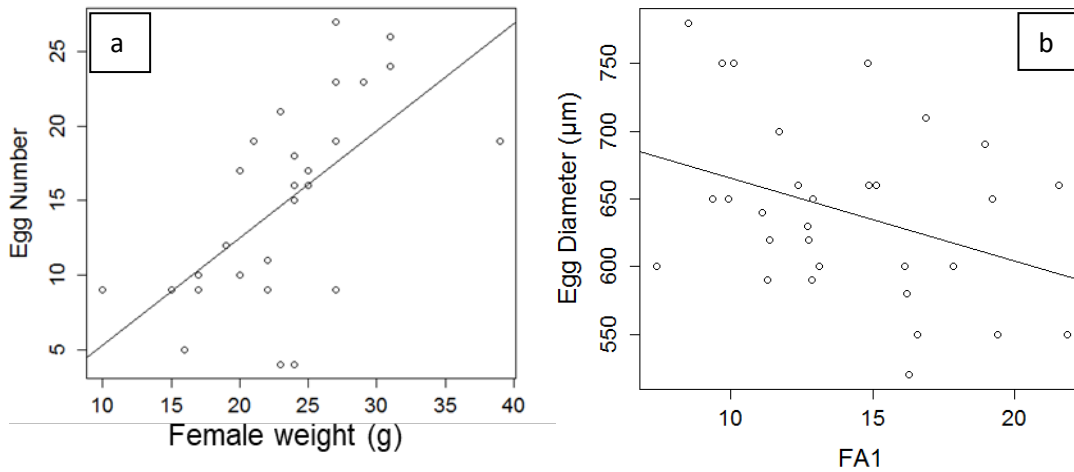
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587 Figure 2: DAPI stained oocytes with and without Microsporidia present in *Gammarus duebeni* and
 588 *Podocotyle atomon* present in the body cavity of *Gammarus zaddachi*. A) Tissue with light
 589 microsporidian infection. The white arrow indicates a cell nucleus. In the inset image the white arrow
 590 identifies Microsporidia present in the cytoplasm of an oocyte. B) Two trematodes encysted in the body
 591 cavity of its host. A biopsied specimen is shown in the inset.

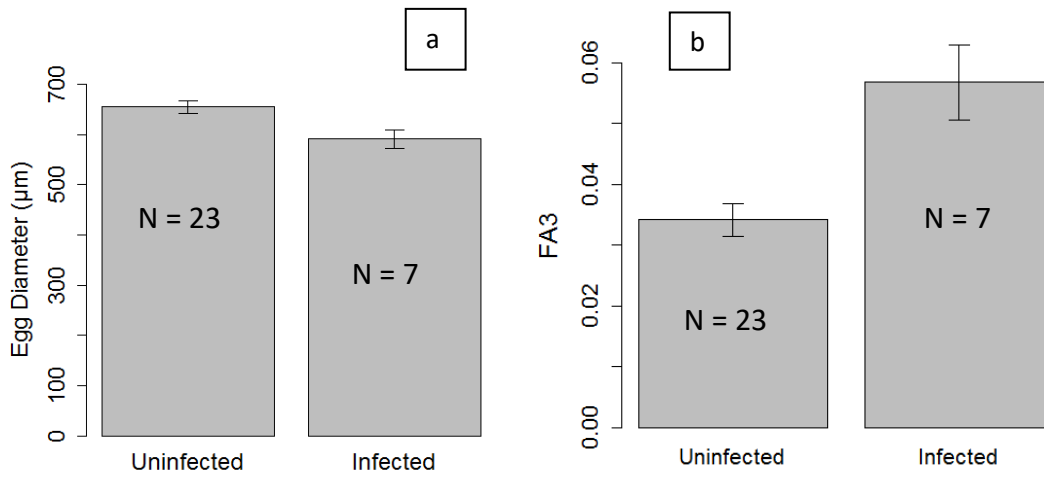
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594 Figure 3. The relationships between a) egg number and female weight (mg); and b) egg diameter (μm)
 595 and FA1 (for all traits) in *Gammarus duebeni*.

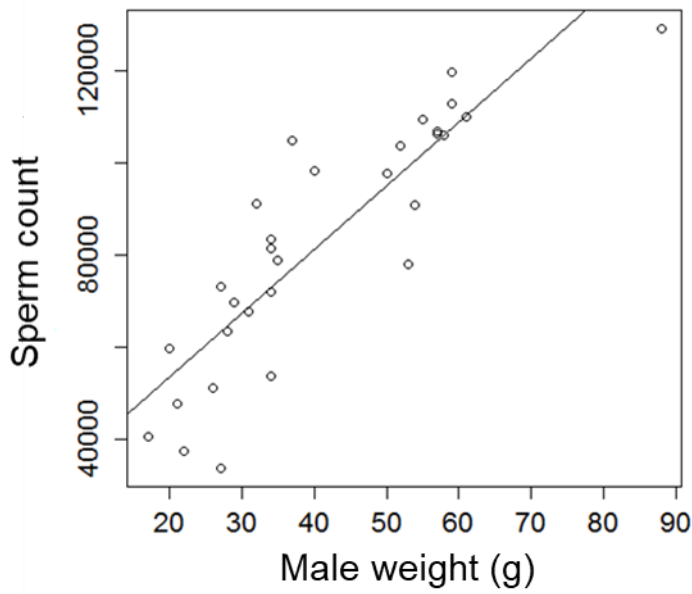
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598 Figure 4. The impact of microsporidian infection expressed on mean a) egg diameter; and b) FA3
 599 (across all traits) in female *Gammarus duebeni*, with ± 1 S.E. bars.

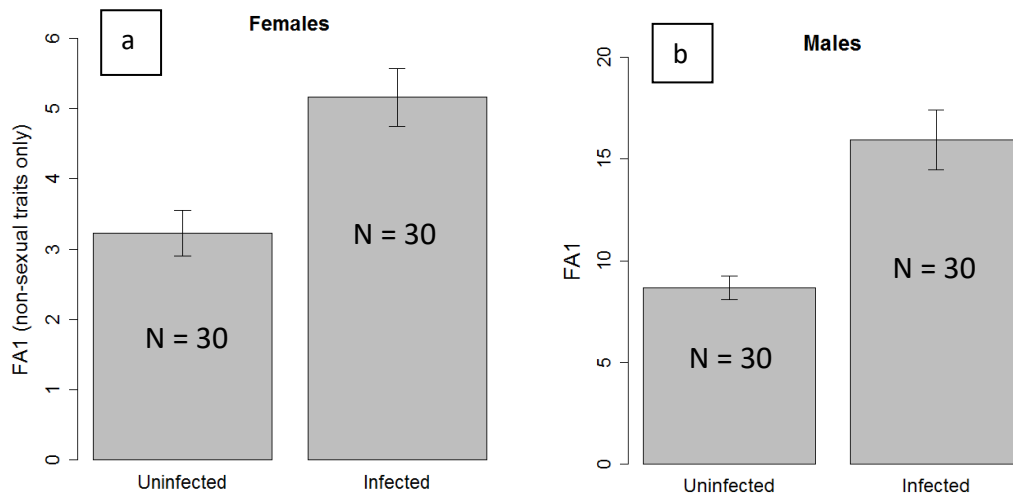
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602 Figure 5. The relationship between weight and sperm number in male *G. duebeni*.

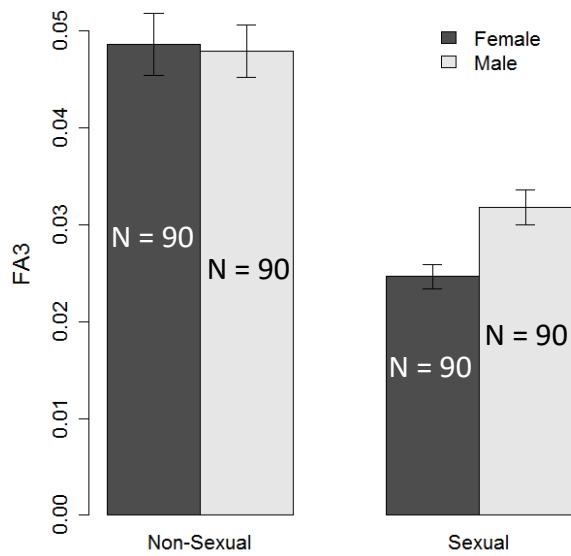
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604

605 Figure 6. The relationship between *P. atomon* infection status and: a) FA1 for non-sexual traits in female
 606 *G. zaddachi*; b) FA1 for all traits in male *G. zaddachi*, with ± 1 S.E. bars.

607



608

609 Figure 7. Mean FA3 in non-sexual and sexual traits, for males and females of both *G. duebeni* and *G.*
 610 *zaddachi*, with ± 1 S.E. bars.

611

612

Trait	Measurement	Abbreviation
Non-sexual: male and females		
<i>Metrical</i>	Width of antenna 1 at the widest part of peduncle article 1	PED

<i>Meristic</i>	Count of flagellum segments on antenna 2	FLA
	Count of spines on the posterior basis of pereopod 6 (walking leg 4)	PER
Sexual: males		
<i>Metrical</i>	Width of pereopod 2 (gnathopod 2) at its widest point	WGNA
	Length of pereopod 2 (gnathopod 2)	LGNA
	Width of tip of genital papillae at widest point	WPAP
	Length of genital papillae	LPAP
<i>Meristic</i>	Count of calceoli on antenna 2*	CAL
Sexual: females		
<i>Metrical</i>	Width at widest point of primary oostegite (found on coxal plate 2) at widest point	WOOE
	Length of bulbous section of primary oostegite	LOOE
<i>Meristic</i>	Count of hairs on primary oostegite	HOOE
	Count of oostegites	NOOE

613 Table 1. Fluctuating Asymmetry measurements and abbreviations used for gammarid
614 amphipods. All measurements taken in both *G. duebeni* and *G. zaddachi*, except for the
615 number of calceoli, which were counted in *G. duebeni* males only (*). Abbreviations refer to
616 the anatomical diagrams in Figure 1 that were measured for both species throughout
617 the study.
618