

**BIOCHEMICAL AND STRUCTURAL CHARACTERISATION OF
DEHALOGENASES FROM MARINE BACTERIA**

Submitted by

Halina Novak

to the University of Exeter as a thesis for the Doctor of Philosophy in Biological
Sciences
(September 2011)

This thesis is available for Library use from September 2014 on the understanding
that it is copyright material and that no quotation from the thesis may be published
without proper acknowledgement.

I certify that all material in this thesis which is not my own work has been identified
and that no material has previously been submitted and approved for the award of a
degree by this or any other University.

Halina Novak

Abstract

An L-haloacid dehalohexanase from the psychrophilic marine bacteria *Psychromonas ingrahamii* has been cloned, over-expressed in a bacterial expression system and biochemically characterised. The enzyme is stable at temperatures of up to 60°C for 90 min and shows highest activity towards substrates with short carbon chains ($\leq C3$). The enzyme is stable in up to 30% ethanol, methanol and DMSO when incubated for 1 h. The K_m for the enzyme is 1.36 mM.

The genome of AQP5750 from the Aquapharm Biodiscovery Ltd Microbial library was sequenced. An L-haloacid dehalohexanase and a haloalkane dehalogenase gene were identified within the genome.

The AQP5750 L-haloacid dehalohexanase has been cloned, over-expressed in a bacterial expression system, biochemically characterised, crystallized and the native and crystal complex structure with chloropropionic acid (MCP) determined. The L-haloacid dehalohexanase from AQP5750 shows highest activity at 55°C towards brominated substrates with short carbon chains ($\leq C3$). The enzyme shows increased activity of 150% in 40% DMSO and 123% in 30% methanol. The L-haloacid dehalohexanase crystal complex structure with covalently bound MCP confirmed Asp 18 as the main catalytic residue. Residues His 183, Asp 186 and Glu 21 in the active site are proposed to be involved in activation of the catalytic water which attacks the ester intermediate in the second part of the S_N2 dehalogenase mechanism.

The AQP5750 haloalkane dehalogenase has been cloned, over-expressed in a bacterial expression system, crystallized and the native and complex structure with 1-hexanol has been determined. Substrate specificity experiments showed that the haloalkane dehalogenase from AQP5750 does not show high activity towards substrates used by other haloalkane dehalogenases with high amino acid sequence identity. The large active site cavity and the presence of Ser 176 and Arg 136 in the hydrophobic binding pocket may alter the binding of substrates tested which could account for the low activity observed.

Acknowledgements

I would like to thank Richard Tennant for his help with sample collection, Dr Hannah Florance for preparing, running and analysing the protein samples on the LC Q-TOF mass spectrometer and for useful discussions, Alex Moorhouse for the preparation of genomic DNA for genome sequencing, Dr Konrad Paszkiewicz for annotating the AQ5750 genome and for useful discussions, Jana Panning for help with the cloning experiments and Dr Michail Isupov for his help with the X-ray crystallography and for useful discussions.

I would like to say a big thank you to Dr Christopher Sayer for teaching me X-ray crystallography and for his help with biochemical characterisation, for useful discussions and continuous support.

I would also like to thank Dr Carrie Rye, Dr Kirsty Line, Dr Anne Marie Hickey and Dr Rob Gibson for their help throughout the project. I would also like to thank everyone in the Biocatalysis centre.

Thank you to the BBSRC and Aquapharm Biodiscovery Ltd for funding and Professor Jenny Littlechild and Dr Dorothee Gotz for supervision.

Finally, I would like to thank my parents for always supporting me and funding my education and to Nick who has been my biggest support over the last four years.

Table of Contents

Abstract.....	2
Acknowledgements	3
List of Figures	10
List of Tables	15
List of Equations	17
Abbreviations	18
Chapter 1- Introduction	20
1.1 Biocatalysts	20
1.2 The marine environment	23
1.2.1 Biopolymers from marine sources	23
1.2.2 Energy from marine sources	23
1.2.3 Pharmaceutically active molecules from the marine environment	24
1.2.4 Enzymes and proteins isolated from the marine environment	25
1.2.5 Extremophiles from the marine environment	26
1.3 Naturally occurring halogenated compounds	28
1.3.1 Using halogens to improve drug activity	29
1.4 Bioremediation	30
1.5 The synthesis of α -halogenated compounds	31
1.5.1 The chemical synthesis of α -halogenated compounds	31
1.6 Enzymes investigated in this project	32
1.6.1 Haloperoxidases (EC 1.11.1.18)	32
1.6.2 Dehalogenases	35
1.7 Marine bacteria containing dehalogenases	50
1.7.1 AQP4626	50
1.7.2 AQP5750	51
1.7.3 <i>Psychromonas ingrahamii</i>	52
1.8 Aims	54
Chapter 2- General Materials and Methods	55
2.1 Microbiology	55
2.1.1 Growth media	55
2.1.2 Growth measurement	56

2.1.3 Marine bacterial growth	56
2.1.4 <i>E. coli</i> growth	56
2.1.5 Bacterial preservation	56
2.1.6 Bacterial harvesting	56
2.2 Molecular biology	56
2.2.1 LB agar/antibiotics plates	56
2.2.2 DNA gel electrophoresis	56
2.2.3 DNA extraction.....	57
2.2.4 PCR reactions	58
2.2.5 DNA purification and concentration	59
2.2.6 Gel extraction	59
2.2.7 Competent <i>E. coli</i> strains	59
2.2.8 Cloning	60
2.3 Biochemistry	64
2.3.1 Induction of protein expression	64
2.3.2 Cell lysis	64
2.3.3 Protein purification	65
2.3.4 Protein parameter calculations	69
2.3.5 Protein concentration determination	69
2.3.6 SDS-PAGE	69
2.3.7 Liquid chromatography mass spectrometry sample (LC-MS) preparation	72
2.3.8 Activity assays	74
2.3.9 Biochemical characterisation.....	75
Chapter 3- Microbial Isolation from the marine environment	77
3.1 Introduction	77
3.2 Materials and Methods	78
3.2.1 Collection of marine microbes	78
3.2.2 Isolation of marine microbial monocultures	78
3.2.3 Marine microbial growth and enzymes assays	78
3.2.4 Identification of the microorganisms in the Exeter microbial library.....	79
3.3 Results and Discussion	79
3.3.1 Identification of microorganism in the Exeter microbial library	79

3.3.2 Activity assays	86
Chapter 4- Purification of L-haloacid dehalohenases from wild type bacteria	88
4.1 Introduction	88
4.1.1 Background	88
4.1.2 Determination of dehalogenase protein sequences	88
4.1.3 Mass spectrometry	89
4.2 Materials and Methods	90
4.2.1 Growth and purification of the L-haloacid dehalohenase from AQP4626 and AQP5750	90
4.2.2 Protein mass spectrometry	91
4.3 Results	91
4.3.1 Purification of the L-haloacid dehalohenase from AQP4626	91
4.3.2 Purification of the L-haloacid dehalohenase from AQP5750	96
4.3.3 Peptide mass spectrometry	102
4.4 Discussion	107
Chapter 5- Determination of L-haloacid dehalohenase gene sequences- the molecular approach	109
5.1 Introduction	109
5.1.1 Determination of L-haloacid dehalohenase gene sequences by degenerate PCR and genome sequencing.	109
5.2 Materials and methods	110
5.2.1 Bioinformatics	110
5.2.2 Degenerate oligonucleotide primer design to amplify the L-haloacid dehalohenase in AQP5750.....	110
5.2.3 Degenerate oligonucleotide primer design to amplify the L-haloacid dehalohenase in AQP4626.....	112
5.2.4 Degenerate PCR compositions and programs	113
5.2.5 Genome sequencing of AQP5750	113
5.3 Results and Discussion	114
5.3.1 Amplification of the L-haloacid dehalohenases from AQP4626 and AQP5750 using degenerate PCR	114
5.3.2 Genome sequencing	114
5.4 Summary	120
Chapter 6- L-haloacid dehalohenase from <i>P. ingrahamii</i>	121

6.1 Introduction	121
6.1.1 Background	121
6.2 Materials and Methods	122
6.2.1 Bioinformatics	122
6.2.2 <i>P. ingrahamii</i> growth	122
6.2.3 Cloning of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	122
6.2.4 Over-expression of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	123
6.2.5 Purification of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	123
6.2.6 Biochemical characterisation of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	124
6.3 Results and Discussion	124
6.3.1 Bioinformatics	124
6.3.2 Cloning and over-expression of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	125
6.3.3 Purification of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	127
6.3.4 Biochemical Characterisation of the L-haloacid dehalohenase from <i>P. ingrahamii</i>	131
6.4 Summary	140
Chapter 7- L-haloacid dehalohenase from AQP5750	141
7.1 Introduction	141
7.1.1 Introduction to crystallography	141
7.2 Materials and Methods	148
7.2.1 Bioinformatics	148
7.2.2 Culturing of AQP5750	148
7.2.3 Cloning of the L-haloacid dehalohenase from AQP5750	149
7.2.4 Over-expression of the L-haloacid dehalohenase from AQP5750	149
7.2.5 Purification of the L-haloacid dehalohenase from AQP5750	150
7.2.6 Biochemical characterisation of the L-haloacid dehalohenase from AQP5750 ..	150
7.2.7 Crystallization	151
7.2.8 X-ray data collection	151
7.2.9 Structure determination	152
7.3 Results	153

7.3.1 Bioinformatics	153
7.3.2 Cloning and over-expression of the L-haloacid dehalohenase from AQP5750 ...	155
7.3.3 Purification of the L-haloacid dehalohenase from AQP5750	157
7.3.4 Biochemical characterisation of the L-haloacid dehalohenase from AQP5750 ..	160
7.3.5 Crystallization	168
7.3.6 Structure determination of the L-haloacid dehalohenase from AQP5750	169
7.3.7 Model building and validation	170
7.3.8 Structure of the L-haloacid dehalohenase from AQP5750	173
7.4 Summary	188
Chapter 8- Haloalkane dehalogenase from AQP5750	189
8.1 Introduction	189
8.2 Materials and Methods	189
8.2.1 Bioinformatics	189
8.2.2 AQP5750 growth.....	189
8.2.3 Cloning of the haloalkane dehalogenase from AQP5750	189
8.2.4 Over-expression of the haloalkane dehalogenase from AQP5750	190
8.2.5 Purification of the haloalkane dehalogenase from AQP5750	191
8.2.6 Substrate specificity of the haloalkane dehalogenase from AQP5750	191
8.2.7 Crystallization	192
8.2.8 X-ray Data Collection	192
8.2.9 Structure determination	193
8.3 Results	194
8.3.1 Bioinformatics	194
8.3.2 Cloning and over-expression of the haloalkane dehalogenase from AQP5750 ..	195
8.3.3 Purification of the haloalkane dehalogenase from AQP5750	197
8.3.4 Biochemical characterisation of the haloalkane dehalogenase from AQP5750 .	200
8.3.5 Substrate specificity of the haloalkane dehalogenase from AQP5750	200
8.3.6 Crystallization	202
8.3.7 Structure determination of the haloalkane dehalogenase from AQP5750	204
8.3.8 Structure solution	205
8.3.9 Model building and validation	205
8.3.10 Structure of the haloalkane dehalogenase from AQP5750	207

8.3.11 Active site	212
8.3.12 Substrate binding	215
8.3.13 Structural comparisons	217
8.3.14 Halide stabilisation	220
8.4 Summary	220
Summary and future work	221
9.1 Summary	221
9.2 Future work	225
10 References	227
11 Appendix	249