

1 **Inter-generational costs of oxidative stress: reduced fitness in daughters of**  
2 **mothers that experienced high levels of oxidative damage during reproduction**

3

4

5 Ana Ángela Romero-Haro<sup>1\*</sup>, Lorenzo Pérez-Rodríguez<sup>2</sup> & Barbara Tschirren<sup>1</sup>

6

7 <sup>1</sup> Centre for Ecology and Conservation, University of Exeter, Penryn TR10 9FE, UK

8 <sup>2</sup> Instituto de Investigación en Recursos Cinegéticos, IREC (CSIC, UCLM, JCCM).

9 Ronda de Toledo 12, 13005 Ciudad Real, Spain

10

11 \*corresponding author: a.romero-haro@exeter.ac.uk

12

13 ORCID:

14 0000-0002-7127-4733 (AAR-H), 0000-0002-5926-1438 (LP-R), 0000-0003-4806-

15 4102 (BT)

16

17 Running title: Parental oxidative damage and reproductive success.

18

19 Keywords: maternal effects, paternal effects, transgenerational effects, oxidative

20 shielding hypothesis, oxidative stress, life history evolution.

21    **Abstract**

22    Parental condition transfer effects occur when the parents' physiological state during  
23    reproduction affects offspring performance. Oxidative damage may mediate such  
24    effects, yet evidence that oxidative damage experienced by parents during  
25    reproduction negatively affects offspring fitness is scarce and limited to early life  
26    stages. We show in Japanese quail (*Coturnix japonica*) that maternal levels of  
27    oxidative damage, measured during reproduction, negatively predict the number of  
28    offspring produced by daughters. This maternal effect on the daughter's reproductive  
29    success was mediated by an effect on hatching success, rather than the number of  
30    eggs laid by daughters. We also observed a negative association between the father's  
31    oxidative damage levels and the number of eggs laid by daughters, but a positive  
32    association between the father's oxidative damage levels and the hatching success of  
33    these eggs. These opposing paternal effects cancelled each other out, resulting in no  
34    overall effect on the number of offspring produced by daughters. No significant  
35    association between a female's own level of oxidative damage during reproduction  
36    and her reproductive success was observed. Our results suggest that oxidative damage  
37    experienced by parents are a better predictor of an individual's reproductive  
38    performance than oxidative damage experienced by the individual itself. Although the  
39    mechanisms underlying these parental condition transfer effects are currently  
40    unknown, changes in egg composition or (epi-)genetic alterations of gametes may  
41    play a role. These findings highlight the importance of an inter-generational  
42    perspective when quantifying costs of physiological stress.

43

44

45

46 **Introduction**

47 The phenotype of the parents can shape the phenotype of the offspring (Mousseau and  
48 Fox 1998; Qvarnström and Price 2001; Crean and Bonduriansky 2014). Such parental  
49 effects can be exerted during the gamete stage or during embryo or juvenile  
50 development, and can often have long-lasting consequences for offspring fitness  
51 (Mousseau and Fox 1998). Parental effects can enhance the offspring's ability to cope  
52 with environmental conditions, and thus be adaptive (Mousseau and Fox 1998;  
53 'predictive adaptive response' Gluckman et al 2005). Alternatively, parental effects  
54 may merely be a (non-adaptive) consequence of the parents' condition during  
55 reproduction, often referred to as parental transmissive effects or parental condition  
56 transfer effects (Qvarnström and Price 2001; Marshall and Uller 2007; Crean and  
57 Bonduriansky 2014). As a consequence, offspring of parents with enhanced  
58 physiological state display a better performance than those of parents with poor  
59 physiological state ('silver spoon effect' Grafen 1988; Monaghan 2008). Oxidative  
60 stress is a key physiological driver of biological processes (Metcalf and Alonso-  
61 Alvarez 2010), and may mediate such parental condition transfer effects.  
62 Oxidative stress is a dynamic process where the rate of reactive oxygen species (ROS)  
63 production exceeds the antioxidant defence capacity of an organism (Halliwell and  
64 Gutteridge 2007; Monaghan et al. 2009). Oxidative damage to proteins, lipids or  
65 DNA is a direct consequence of non-alleviated oxidative stress and can impair  
66 physiological processes and individual performance (Halliwell and Gutteridge 2007;  
67 Monaghan et al. 2009). Oxidative stress may also disrupt the redox signalling  
68 pathways, but with poorly understood consequences (Schieber and Chander 2014;  
69 Ayala et al. 2014). Cell metabolism, and cellular respiration in particular, contribute  
70 substantially to ROS production (Halliwell and Gutteridge 2007). However, also

71 extrinsic factors, such as radiation, changes in temperature, exposure to xenobiotics or  
72 pollution can increase ROS levels in an organism (Araujo et al. 2008; Isaksson 2010)  
73 and indeed this additional burden may often tip the balance and result in oxidative  
74 damage (Isaksson 2010). High levels of oxidative damage are associated with the  
75 occurrence of certain diseases, like cancer (Valko et al. 2007; Ayala et al. 2014) and  
76 they have been related to the ageing process (Martinez de Toda et al 2020), shorter  
77 lifespan (Noguera et al. 2012; Vitikainen et al. 2016), as well as impaired  
78 reproductive success (Stier et al. 2012). However, the causal role of oxidative damage  
79 in mediating these processes is still debated (e.g. Speakman and Selman 2011;  
80 Metcalfe and Alonso-Alvarez 2010; Blount et al. 2015; Alonso-Alvarez et al. 2017).  
81 Importantly, the negative consequences of oxidative damage experienced by an  
82 individual might not be limited to the individual itself but may expand to the next  
83 generation (Blount et al. 2015). A parent experiencing oxidative damage may, for  
84 example, be constrained in its ability to optimally provision or care for the developing  
85 offspring (e.g. because of impaired uterus and placental function or reduced milk  
86 yield; Al-Gubory et al. 2010; Burton and Jauniaux 2011; Napierala et al. 2016, 2019).  
87 Alternatively, oxidized molecules may be directly transferred from the mother to the  
88 offspring (e.g. via the egg; Mohiti-Asli et al. 2008; Surai et al. 2016; placenta;  
89 Rossner et al 2009; or the milk; Napierala et al. 2019) and influence offspring  
90 development. Furthermore, oxidative damage may induce (epi-)genetic changes in the  
91 parents that may be transferred to the next generation (Velando et al. 2008; Aitken et  
92 al. 2016). For example, oxidative damage might shorten telomeres (Kalmbach et al  
93 2013; Heidinger and Young 2020) or affect DNA methylation patterns (Tunc and  
94 Tremellen 2009; Menezo et al. 2016) in gametes. As a result, oxidative damage  
95 experienced by a parent during reproduction may negatively affect offspring

96 development and, ultimately, offspring fitness. Because of such potential parental  
97 condition transfer effects, it has been proposed that parents may actively decrease  
98 oxidative damage levels before reproduction in order to shield the offspring from  
99 harmful effects (Oxidative Shielding Hypothesis; Blount et al. 2015). Yet to date,  
100 evidence for negative fitness consequences of oxidative damage experienced by the  
101 parents for the next generation is scarce and limited to early life stages (i.e. low birth  
102 weight and illness in infants; Al-Gubory et al. 2010; Napierala et al. 2016, 2019,  
103 Viblanc et al. 2018; and reduced early life survival; Bize et al. 2008; Vitikainen et al.  
104 2016; Dupoué et al. 2020) or are indirectly inferred from parental exposure to  
105 oxidative stress-related factors, such as xenobiotics (Hamlin and Guillette 2011).  
106 However, the consequences of oxidative damage experienced by parents for the long-  
107 term performance of the offspring are currently unknown. A long-term perspective is  
108 required, however, to fully understand the fitness consequences of oxidative damage  
109 experienced by parents for the next generation, and to quantify the relative importance  
110 of oxidative damage-mediated parental condition transfer effects vs. effects of  
111 oxidative damage experienced by the adult individual itself. Such an inter-  
112 generational view is essential not only to gain insights into the role of oxidative  
113 damage in life history evolution, but also to assess the consequences of environmental  
114 stressors on the resilience and stability of natural populations.  
115 Here, we quantified if maternal and / or paternal levels of oxidative damage to lipids  
116 (quantified as plasma levels of malondialdehyde, MDA) when the offspring were  
117 conceived predict key fitness components in their daughters, namely reproductive  
118 success and lifespan, using Japanese quail (*Coturnix japonica*) as a study system.  
119 Furthermore, we quantified the relative importance of oxidative damage-mediated  
120 parental condition transfer effects vs. effects of oxidative damage experienced by the

121 focal individual itself on these fitness components. Given the short-term negative  
122 fitness consequences (i.e. reduced early life survival) of parental oxidative damage  
123 observed in previous studies (Bize et al. 2008; Vitikainen et al. 2016; Dupoué et al.  
124 2020), we predict that levels of parental oxidative damage will be negatively  
125 associated with the daughters' fitness. Furthermore, based on previous findings where  
126 high within-individual levels of oxidative damage were related to reduced fitness  
127 (Stier et al. 2012; Costantini et al. 2016; Noguera et al. 2012; Vitikainen et al. 2016),  
128 we predict a negative association between a focal female's oxidative damage levels  
129 and her reproductive success and lifespan.

130

## 131 **Methods**

### 132 *Breeding conditions*

133 The study was conducted in a captive population of Japanese quail (*Coturnix*  
134 *japonica*) maintained in large outdoor aviaries (7m × 5.5 m each) at the University of  
135 Zurich, Switzerland. Females were maintained in a single sex aviary, and males in a  
136 mixed sex aviary together with non-experimental females (see Pick et al. 2016 for a  
137 detailed description of the breeding and husbandry conditions). For this study, adult  
138 males and females (age: 189-295 days old), were randomly selected from the  
139 population and housed during three weeks in pairs (N = 22 randomly assigned  
140 breeding pairs) in breeding cages (122 x 50 x 50 cm) in the animal facility on a 16:8  
141 light:dark cycle at approximately 20 °C. Eggs were collected on the day they were  
142 laid, weighed (to the nearest 0.01g), and artificially incubated (Favorit, HEKA  
143 Brutgeräte). During the first 14 days, eggs were maintained at 37.8 °C and 55%  
144 humidity. They were then transferred to a hatcher (Favorit, HEKA Brutgeräte) and  
145 kept at 37.6 °C and 80% humidity until hatching. Chicks were kept in a heated cage

146 (109 × 57 × 25 cm, Kükenaufzuchtbox Nr 4002/C, HEKA Brutgeräte) for two weeks  
147 after hatching. The first five days the temperature was kept at 35–38 °C, then slowly  
148 lowered to 25 °C over the next 9 days. After two weeks, chicks were transferred to  
149 rearing cages within the breeding facility. At the age of four weeks, the birds were  
150 released into the outdoor aviaries (see Pick et al. 2016 for details). Thus, all offspring  
151 were reared in mixed-family groups under standardised conditions. Body mass was  
152 measured at hatching (to the nearest 0.01g) and at adulthood (i.e. when six months  
153 old; to the nearest 1g). We focused on daughters because males show a low variation  
154 in reproductive performance (Pick et al. 2017).

155 At the age of six months, one randomly chosen virgin daughter per breeding pair  
156 (hereafter referred to as ‘focal females’; N = 22) was brought into the breeding facility  
157 and housed in a breeding cage with a random male from the population during three  
158 weeks to determine their reproductive success during the reproductive event. To avoid  
159 potential effects of incubation conditions, the eggs were removed on the day they  
160 were laid and incubated under standardised conditions as described above to  
161 determine hatching success and the number of offspring produced. After breeding, the  
162 focal females were moved back into the outdoor aviaries where they were kept for  
163 their entire life to record their lifespan. All focal females thus experienced the same  
164 standardised conditions from incubation to death. Death of the focal females occurred  
165 either naturally (N = 17) or they were euthanized because they had reached a pre-  
166 defined humane endpoint (N = 5).

167 To quantify levels of oxidative damage, we took a blood sample from the parents of  
168 the focal females when they were moved into the breeding cages (i.e. when the focal  
169 female was conceived). We also blood-sampled the focal females at the beginning of  
170 their reproductive event. Blood samples (approx. 100µl) were taken from the brachial

171 vein using heparinised capillary tubes. Samples were stored at 4°C until centrifugation  
172 (5 min at 20 °C and 2000 g) within 4 h. Plasma was then separated and frozen at –80  
173 °C until analysis.

174

#### 175 *Quantification of lipid oxidative damage*

176 Although oxidative stress not always results in oxidative damage, oxidative damage is  
177 generated under non-alleviated oxidative stress conditions (Halliwell and Gutteridge  
178 2007; Monaghan et al. 2009). Oxidative damage is thus a direct consequence of  
179 oxidative stress (Monaghan et al. 2009) and lipids are particularly susceptible to such  
180 damage (Del Rio et al 2005; Hulbert et al. 2007). To quantify lipid oxidative damage  
181 in focal females and their parents, we measured plasma levels of malondialdehyde  
182 acid (MDA), one of the end-point molecules in the lipid peroxidation cascade  
183 (Halliwell and Gutteridge 2007; Mateos and Bravo 2007). MDA is a commonly used  
184 marker of oxidative damage (Del Rio et al. 2005; Noguera et al. 2012; Vitikainen et  
185 al. 2016; Vagasi et al. 2019) and plasma levels of MDA have been reported to reflect  
186 those in other tissues, thus being a proxy of whole-body oxidative damage (Argüelles  
187 et al. 2004; Margaritelis et al. 2015). MDA has also been found to be an extremely  
188 toxic and mutagenic molecule with high reactivity, interacting with DNA and proteins  
189 (Del Rio et al. 2005; Nair et al. 2007). High MDA concentrations have been related to  
190 numerous illnesses in humans (reviews in Ayala et al. 2014), as well as impaired  
191 fitness in non-human animals (Noguera et al. 2012; Vitikainen et al. 2016; Vagasi et  
192 al. 2019).

193 MDA quantification was performed using HPLC following the protocol of Agarwal  
194 and Chase (2002) with modifications by Nussey et al. (2009). In short, a standard  
195 curve for calibration was prepared using a 1,1,3,3-tetraethoxypropane stock solution



196 (5  $\mu$ M in 40% ethanol), serially diluted using 40% ethanol. 50  $\mu$ L of a butylated  
197 hydroxytoluene (BHT) solution (0.05% w/v in 95% ethanol), 400  $\mu$ L of a phosphoric  
198 acid solution (0.44 M), and 100  $\mu$ L of a thiobarbituric acid (TBA) solution (42 mM)  
199 were added to 20  $\mu$ L of plasma and 30 $\mu$ L of Milli-Q water or to 50  $\mu$ L of standard,  
200 vortexed and heated at 100°C for 1 h to allow for the formation of MDA-TBA  
201 adducts. The reaction was stopped by placing samples and standards on ice. 250  $\mu$ L of  
202 *n*-butanol was then added to extract the MDA-TBA complex. Tubes were  
203 subsequently vortexed and centrifuged at 18 000 *g* for 3 min at 4°C. 100  $\mu$ L of the  
204 upper (*n*-butanol) phase were then moved to HPLC vials, which were immediately  
205 saturated with N<sub>2</sub> to avoid oxidation (see also Romero-Haro and Alonso-Alvarez  
206 2014). Samples were injected into an Agilent 1100 series HPLC system (Agilent,  
207 Waldbronn, Germany) fitted with a fluorescence detector set and a 5- $\mu$ m ODS-2 C-18  
208 4.0 x 250-mm column maintained at 37°C. The mobile phase was MeOH : KH<sub>2</sub>PO<sub>4</sub>  
209 (50 mM; 40 : 60 v/v), running isocratically for 10 min at a flow rate of 1 mL/min.  
210 Chromatograms were collected at 515 nm (excitation) and 553 nm (emission). Some  
211 samples were measured in duplicate both within and across laboratory sessions to  
212 quantify repeatabilities (intrasession:  $r = 0.95$ ,  $n = 28$ ,  $P < 0.001$ , intersession:  $r =$   
213  $0.76$ ,  $n = 12$ ,  $P = 0.001$ ). Quantification of MDA could not be done in two fathers and  
214 four focal females because samples were lost during handling, resulting in lower  
215 sample sizes for some comparisons. MDA concentrations were log transformed for  
216 the statistical analyses.

217

### 218 *Statistical analyses*

219 First, we ran linear models to test if a focal female's own oxidative damage (i.e.  
220 plasma MDA levels) or the oxidative damage experienced by her parents when she

221 was conceived predicted the number of offspring (i.e. number of hatchlings) produced  
222 by the focal female during the reproductive event. Second, we further explored  
223 variation in focal female reproductive success by running generalised linear models  
224 with a quasibinomial error structure to test if a focal female's own oxidative damage,  
225 or the oxidative damage experienced by her parents when she was conceived,  
226 predicted the number of eggs she laid during the reproductive event or the hatching  
227 success of these eggs. Quasibinomial models were used instead of binomial models  
228 because of overdispersion. Third, we ran linear models to test if a focal female's own  
229 oxidative damage, or the oxidative damage experienced by her parents when she was  
230 conceived, predicted focal female lifespan while accounting for the cause of death  
231 (natural / euthanized). Focal female body mass at adulthood was included as an  
232 additional covariate in the models described above. Fourth, we used linear models to  
233 test if parental oxidative damage levels during reproduction were associated with the  
234 size of the egg the focal female developed in, her hatching mass or her adult body  
235 mass. Finally, we used linear models to test for parent-offspring resemblance in  
236 oxidative damage during the reproductive period. Standardized MDA values were  
237 used for parent-offspring regressions.

238 Absolute MDA levels were used in the models described above. In addition we ran  
239 the same models with MDA levels corrected for circulating triglyceride  
240 concentrations (see Supplementary material S1). Furthermore, analyses of  
241 associations between mother and father oxidative damage levels are presented in  
242 Supplementary material S2.

243 For quasibinomial models, significance of predictors was determined by comparing  
244 two nested models, with and without the factor of interest, using likelihood ratio tests.

245 All statistical analyses were performed in *R* version 3.6.2 (R Development Core Team

246 2014). Terms were removed from the final models if  $P > 0.05$ . Normality of the  
247 residuals of linear models was confirmed by visual inspection and Shapiro-Wilk tests.  
248 Means  $\pm$  SE are presented.

249

## 250 **Results**

251

### 252 *Focal female reproductive success*

253 A focal female's own levels of oxidative damage, measured during the reproductive  
254 event, did not predict the number of offspring she produced ( $t_{1,15} = 0.895$ ,  $P = 0.385$ ,  
255 Fig. 1a). In contrast, levels of oxidative damage in the mother, measured when the focal  
256 female was conceived, were negatively associated with the number of offspring  
257 produced by the focal female ( $\beta \pm$  SE:  $-13.208 \pm 5.816$ ;  $t_{1,20} = -2.271$ ,  $P = 0.034$ , Fig.  
258 1b). No association between the father's oxidative damage, measured when the focal  
259 female was conceived, and the number of offspring produced by the focal female was  
260 found ( $t_{1,12} = -0.107$ ,  $P = 0.916$ , Fig. 1c).

261 In a second step, we further dissected variation in focal female reproductive success  
262 by separately analyzing the number of eggs the focal female laid during the  
263 reproductive event and the hatching success of these eggs. The number of eggs laid by  
264 a focal female was neither predicted by her own oxidative damage levels ( $\chi^2 = 0.440$ ,  
265  $P = 0.715$ ; Fig. 2a) nor by those of her mother, measured when the focal female was  
266 conceived ( $\chi^2 = 3.257$ ,  $P = 0.315$ , Fig. 2b). In contrast, the number of eggs laid by a  
267 focal female was negatively associated with the father's levels of oxidative damage,  
268 measured when the focal female was conceived ( $\beta \pm$  SE =  $-7.724 \pm 2.316$ ;  $\chi^2 = 46.155$ ,  
269  $P < 0.001$ , Fig. 2c). Hatching success was not predicted by the focal female's own  
270 levels of oxidative damage during reproduction ( $\chi^2 = 0.951$ ,  $P = 0.354$ ; Fig. 3a) but

271 the mother's oxidative damage levels, measured when the focal female was  
272 conceived, negatively predicted the hatching success of eggs laid by the focal female  
273 ( $\beta \pm SE = -3.554 \pm 1.715$ ,  $\chi^2 = 4.355$ ,  $P = 0.038$ , Fig. 3b). Furthermore, there was a  
274 positive association between the father's oxidative damage levels, measured when the  
275 focal female was conceived, and hatching success ( $\beta \pm SE = 2.545 \pm 1.267$ ,  $\chi^2 =$   
276  $4.127$ ,  $P = 0.043$ , Fig. 3c).

277 These results did not change when including the focal female's body mass at  
278 adulthood as an additional factor into the models. Focal female body mass was not  
279 significantly associated with a focal female's reproductive success (all  $P > 0.285$ ).

280

#### 281 *Female lifespan*

282 A focal female's lifespan was neither associated with her own levels of oxidative  
283 damage measured during reproduction ( $t_{1, 12} = 0.250$ ,  $P = 0.807$ ) nor by oxidative  
284 damage levels in the mother ( $t_{1, 20} = -1.272$ ,  $P = 0.218$ ) or the father ( $t_{1, 17} = 0.646$ ,  $P$   
285  $= 0.527$ ), measured when the focal female was conceived. Including the cause of  
286 death (natural or euthanized) in the analysis did not change the results, and there was  
287 no association between the parents' or focal female's levels of oxidative damage and  
288 the cause of death (all  $P > 0.176$ ). These results did not change when including the  
289 focal female's body mass at adulthood as an additional factor into the model. A focal  
290 female's body mass was not significantly associated with lifespan ( $t_{1, 16} = 0.493$ ,  $P =$   
291  $0.628$ ).

292

#### 293 *Egg size and female body mass*

294 There was no significant association between the mother's or the father's levels of  
295 oxidative damage when the focal female was conceived and the size of the egg the

296 focal female developed in (mother MDA:  $t_{1,17} = -0.720$ ,  $P = 0.482$ , father MDA:  $t_{1,17}$   
297  $= 0.794$ ,  $P = 0.438$ ), the focal female's body mass at hatching (mother MDA:  $t_{1,17} = -$   
298  $0.501$ ,  $P = 0.623$ , father MDA:  $t_{1,17} = 0.449$ ,  $P = 0.659$ ) or the focal female's body  
299 mass at adulthood (mother MDA:  $t_{1,17} = -0.917$ ,  $P = 0.372$ , father MDA:  $t_{1,17} = 0.979$ ,  
300  $P = 0.341$ ).

301

### 302 *Parent-daughter resemblance in oxidative damage*

303 There was no significant mother-daughter ( $\beta \pm SE = 0.097 \pm 0.230$ ,  $t_{1,16} = 0.422$ ,  $P =$   
304  $0.678$ ) or father-daughter resemblance ( $\beta \pm SE = -0.028 \pm 0.208$ ,  $t_{1,14} = -0.133$ ,  $P =$   
305  $0.896$ ) in the levels of oxidative damage measured during reproduction.

306

### 307 **Discussion**

308 Our study provides evidence for an inter-generational link between parental oxidative  
309 damage at offspring conception and key components of offspring reproductive  
310 success. We observed associations between offspring reproductive performance and  
311 the oxidative damage of both parents, but different fitness components were affected  
312 depending on parental sex. The mother's level of oxidative damage at offspring  
313 conception was significantly negatively associated with the number of offspring a  
314 focal female (i.e. her daughter) produced, and this effect was mainly mediated  
315 through an effect on the hatching success of eggs laid by a focal female, rather than  
316 the number of eggs she laid. In contrast, the father's level of oxidative damage was  
317 negatively associated with the number of eggs laid by the focal female, but positively  
318 associated with the hatching success of these eggs. These opposing paternal effects  
319 cancelled each other out, resulting in no significant association between the father's  
320 level of oxidative damage and the number of offspring produced by a focal female.

321 No significant association between a focal female's own oxidative damage during  
322 reproduction and her reproductive success was observed. Furthermore, no direct or  
323 parental effect of oxidative damage on focal female lifespan was observed.  
324 These results show that parental physiological states (i.e. levels of oxidative damage)  
325 can have long-term fitness consequences for the next generation, and suggest that  
326 inter-generational effects may be a stronger predictor of offspring performance than  
327 levels of oxidative damage experienced by the individual itself during reproduction.  
328 Three previous studies have reported a negative association between maternal  
329 oxidative stress-related measurements during reproduction and offspring survival  
330 early in life. In Alpine swifts (*Apus melba*), females with erythrocytes less resistant to  
331 an oxidative attack (KRL bioassay) laid eggs that were less likely to hatch (Bize et al.  
332 2008). In Banded mongoose (*Mungos mungo*), pups of females with high plasma  
333 MDA levels had a lower survival probability until emergence from the den  
334 (Vitikainen et al. 2016). And in common lizards (*Zootoca vivipara*), maternal plasma  
335 levels of oxidative damage (dROMs test, hydroperoxides) were negatively related to  
336 early life offspring survival (Dupoué et al. 2020). We did not find paternal effects on  
337 offspring lifespan. To our knowledge, our study is the first, however, to demonstrate  
338 long-term consequences of parental oxidative damage on key components of offspring  
339 reproductive success.  
340 Currently, we can only speculate about the mechanisms underlying the observed  
341 parental effects on offspring reproductive performance. The environment and  
342 individual encounters during the first stages of life can have long-lasting  
343 consequences (Lindström 1999). The observed effect of oxidative damage  
344 experienced by mothers during reproduction on the reproductive success of their  
345 daughters could thus be mediated by an inability of physiologically stressed females

346 to optimally provision or care for their offspring early in life, with long-term  
347 consequences for their reproductive performance. Given that in our study eggs were  
348 artificially incubated and chicks reared under standardized conditions, such maternal  
349 provisioning effects would have to occur before egg laying, through a change in egg  
350 size or quality. We did not observe a relationship between maternal oxidative damage  
351 and egg size. However, mothers experiencing high levels of oxidative damage could  
352 be constrained in the amount of antioxidants they transfer to the egg because of a  
353 direct trade-off between the use of antioxidants for self-maintenance or reproduction.  
354 Indeed, in great tits (*Parus major*), plasma levels of oxidative damage in females were  
355 negatively associated with the levels of yolk antioxidants in their eggs (Giordano et al.  
356 2015). Similarly, dietary antioxidant supplementation reduces the levels of oxidative  
357 damage and increases the levels of antioxidants in females, and in the eggs and chicks  
358 they produce (Surai et al. 2016). Also, the allocation of different maternal resources to  
359 the eggs is non-independent (Royle et al 2001; Blount et al. 2002; Boulinier and  
360 Staszewski 2008). Thus, maternal oxidative damage may not only affect the allocation  
361 of antioxidants to eggs, but also the allocation of other egg components, such as  
362 hormones (Groothuis and Schwabl 2008) or antibodies (Boulinier and Staszewski  
363 2008), with downstream consequences for offspring development and performance  
364 (Groothuis et al. 2005; Surai et al. 2016), potentially affecting reproductive  
365 performance.

366 Alternatively, oxidized molecules in the mother's circulation may be directly  
367 incorporated into the eggs (Grune et al. 2001; Mohiti-Asli et al. 2008). Both, a  
368 reduced allocation of antioxidants to eggs or the direct transfer of oxidized molecules  
369 to eggs as a consequence of high levels of oxidative damage in the mother, may  
370 increase oxidative stress experienced by focal females during prenatal development.

371 The developing organism is particularly sensitive to oxidative stress (Monaghan et al.  
372 2009; Metcalfe and Alonso-Alvarez 2010), which may negatively affect the  
373 development of the reproductive system and exert negative, long-lasting effects on  
374 reproductive success of the individual. For example, in zebrafish (*Dario rerio*), levels  
375 of oxidative stress experienced during development reduced the odds of breeding and  
376 the rate of fertilized eggs (Newman et al 2015). Furthermore, in laboratory rats, adult  
377 females that developed under gestational hypoxia, a condition commonly used to  
378 generate placental oxidative stress in biomedical studies (Silvestro et al 2020),  
379 showed lower ovarian reserve than females that developed under normoxia (Aiken et  
380 al. 2019a, b). Interestingly, we observed no significant association between maternal  
381 oxidative damage at offspring conception and offspring body mass at hatching or at  
382 adulthood. This makes it unlikely that impaired growth and / or catch-up growth  
383 (Metcalfe and Monaghan 2001) mediated the observed maternal effects on the  
384 daughters' reproductive success.

385 In addition to the significant negative associations between maternal oxidative  
386 damage at offspring conception and a focal female's reproductive success, we also  
387 observed a significant negative association between paternal oxidative damage and  
388 the number of eggs laid by a focal female. Furthermore, and unexpectedly, we found a  
389 significant positive association between paternal oxidative damage and the hatching  
390 success of these eggs. Paternal effects on offspring reproductive performance could  
391 arise if females change the investment in eggs in response to partner quality (Burley  
392 1986; Gowaty 2008). Thus, the observed paternal effects on aspects of offspring  
393 reproductive success may indeed be maternal effects. Females may perceive a male's  
394 oxidative status through condition-dependent signals, such as colour ornaments  
395 (Galván and Alonso-Alvarez 2009), vocalisations (Messina et al. 2017), or



396 behavioural displays (Metcalf and Alonso-Alvarez 2010) and change egg  
397 provisioning accordingly. We did not observe a relationship between the father's  
398 oxidative damage levels and the size of the eggs his partner laid. Yet, changes in egg  
399 composition may occur in response to partner quality. Indeed, females have been  
400 found to differentially deposit hormones and antioxidants in eggs in relation to partner  
401 quality (Gil et al. 1999; Saino et al. 2002; Williamson et al. 2006; Remes 2011),  
402 which may affect offspring reproductive performance later in life (Müller et al. 2009).  
403 Second, the negative link between the father's oxidative status and the number of eggs  
404 laid by daughters could arise because of direct mutagenic effects of oxidative damage  
405 on the male's germline, with negative consequences for the offspring (Aitken and  
406 Krausz 2001; Gavriliouk and Aitken 2015). Spermatozoa are known to be particularly  
407 sensitive to oxidative stress, and indeed, oxidative stress is considered to be one of the  
408 main causes of male infertility (Tremellen 2008) and increased offspring morbidity  
409 (Aitken and Krausz 2001; Aitken et al. 2014). Furthermore, the observed relationships  
410 between paternal oxidative damage and aspects of offspring reproductive performance  
411 could be caused by the transfer of non-genetic information from the father to the  
412 zygote via sperm. Only relatively recently it has been recognized that sperm contains  
413 more than just paternal DNA and that non-genetic sperm components, such as DNA  
414 methylation, chromatin modifications, RNAs or proteins can have long-term  
415 consequences for offspring development and performance (reviewed in Krawetz  
416 2005; Immler 2018). The specific mechanisms underlying the connection between  
417 paternal sperm characteristics and offspring performance are still poorly understood,  
418 although changes in gene expression patterns during the embryonic period have been  
419 pointed out (Chen et al. 2016). Importantly, previous studies suggest that oxidative  
420 damage experienced by a male may affect these non-genetic sperm components, and

421 thus offspring development. For example, in laboratory mice, adult females conceived  
422 by sperm with experimentally increased levels of oxidative damage were smaller and  
423 showed increased adiposity and reduced glucose tolerance compared to control  
424 females (Lane et al. 2014). Furthermore, environmental oxidative stress-related  
425 factors, such as exposure to toxins and pollutants, have also been shown to modify  
426 non-genetic sperm components and affect the phenotype and health of descendants  
427 (reviewed in Jiménez-Chillarón et al. 2015; Xavier et al. 2019), highlighting the role  
428 of oxidative stress and the resulting oxidative damage in mediating environmentally-  
429 induced epigenetic remodeling.

430 Importantly, direct DNA damage in gametes caused by oxidative damage and the  
431 transfer of non-genetic information (e.g. epigenetic states) to the offspring through the  
432 gamete could also contribute to the maternal oxidative damage effect on daughter's  
433 reproductive success.

434 Our study is correlational and experimental manipulations of parental oxidative status  
435 are needed to further understand why and how maternal and paternal physiological  
436 states affect different components of the daughters' reproductive performance. In  
437 addition, analyses of oxidative stress in gametes, eggs and embryos may help to  
438 identify the relative importance of the different, but not mutually exclusive  
439 mechanisms discussed above.

440 In contrast to the associations between parental oxidative damage and a focal female  
441 reproductive success, we found no link between a focal female's own levels of  
442 oxidative damage and her fitness, which is in disagreement with some previous  
443 studies (Bize et al. 2008; Stier et al. 2012; Noguera et al. 2012; Costantini et al. 2016;  
444 Vitikainen et al. 2016, but see Losdat et al. 2012; Fowler et al. 2018; van de  
445 Crommenacker et al. 2017). The potential importance of the mechanisms triggering

446 inter-generational effects discussed above, the enhanced sensitivity to oxidative stress  
447 early in life, and the fact that an individual's levels of oxidative damage during  
448 development and at adulthood appear to be unrelated (Romero-Haro and Alonso-  
449 Alvarez 2014) might explain this unexpected result.

450 In conclusion, our study provides evidence that both the mother's and the father's  
451 oxidative state at offspring conception have long-term consequences for key aspects  
452 of offspring reproductive performance. Such inter-generational oxidative damage  
453 effects may promote the evolution of oxidative shielding mechanisms in parents  
454 during reproduction to protect the descendants and, in turn, increase fitness return via  
455 an enhanced reproductive success of daughters (Blount et al. 2015). Importantly,  
456 parental levels of oxidative damage were stronger predictors of offspring fitness than  
457 levels of oxidative damage experienced by the adult individual itself. This finding  
458 highlights the importance of an inter-and transgenerational perspective in the study of  
459 oxidative stress and life history evolution, and it suggests that natural or human-  
460 induced environmental stressors may have delayed, transgenerational effects on  
461 natural populations, leading to an underestimation of their effect on population health,  
462 resilience and stability.

463

464 Data accessibility

465 Data available from the Dryad Digital Repository:

466 <https://doi.org/10.5061/dryad.1ns1rn8v6>

467

468 Ethics

469 All procedures were conducted under licenses provided by the Veterinary Office of  
470 the Canton of Zurich, Switzerland (permit numbers 195/2010; 14/2014; 156) and the  
471 ethical committee of the University of Exeter (permit eCORN002475).

472

473 Authors' contributions

474 AAR-H and BT designed and performed the research, analysed the data and wrote the  
475 manuscript. LP-R enabled the lab analyses. All authors discussed the results and  
476 commented on the manuscript.

477

478 Competing interests

479 The authors declare no competing interests.

480

481 Funding

482 This work was supported by the European Union's Horizon 2020 research and  
483 innovation program under the Marie Skłodowska-Curie grant agreement 842085 (to  
484 AAR-H), the Swiss National Science Foundation (PP00P3\_128386 and  
485 PP00P3\_157455 (to BT)) and the Spanish Ministerio de Ciencia, Innovación y  
486 Universidades (PGC2018-099596-B-I00 (to LP-R), co-financed by  
487 the European Regional Development Fund).

488

489 Acknowledgements

490 We thank the quail husbandry team for help with data collection and Jon Blount and  
491 Magali Meniri for comments on the manuscript.

492

493 **References**

- 494 Agarwal R.J. and S.D. Chase. 2002. Rapid, fluorimetric-liquid chromatographic  
495 determination of malondialdehyde in biological samples. *J Chromatogr B*  
496 775:121–126.
- 497 Aiken C.E., J.L. Tarry-Adkins, A.M. Spiroski, A.M. Nuzzo, T.J. Ashmore, A. Rolfo,  
498 M.J. Sutherland, et al. 2019a. Chronic fetal hypoxia disrupts the peri-conceptual  
499 environment in next-generation adult female rats. *J Physiol-London* 597:2391–  
500 2401.
- 501 Aiken C.E., J.L. Tarry-Adkins, A.M. Spiroski, A.M. Nuzzo, T.J. Ashmore, A. Rolfo,  
502 M.J. Sutherland, et al. 2019b. Chronic gestational hypoxia accelerates ovarian  
503 aging and lowers ovarian reserve in next-generation adult rats. *Faseb J* 33:7758–  
504 7766.
- 505 Aitken R.J. and C. Krausz. 2001. Oxidative stress, DNA damage and the Y  
506 chromosome. *Reproduction* 122:497–506.
- 507 Aitken R.J., T. Smith, M. Jobling, M. Baker, and G. De Iuliis. 2014 Oxidative stress  
508 and male reproductive health. *Asian J Androl* 16:31.
- 509 Aitken R.J., Z. Gibb, M.A. Baker, J. Drevet, and P. Gharagozloo. 2016. Causes and  
510 consequences of oxidative stress in spermatozoa. *Reprod Fertil Dev* 28:1.
- 511 Al-Gubory K.H., P.A. Fowler, and C. Garrel. 2010. The roles of cellular reactive  
512 oxygen species, oxidative stress and antioxidants in pregnancy outcomes. *Int J*  
513 *Biochem Cell Biol* 42:1634–1650.

514 Alonso-Alvarez C., T. Canelo, and A.A. Romero-Haro. 2017. The oxidative cost of  
515 reproduction: theoretical questions and alternative mechanisms. *Bioscience*  
516 67:258–270.

517 Araujo J.A., B. Barajas, M. Kleinman, X. Wang, B.J. Bennett, K.W. Gong, M. Navab,  
518 et al. 2008. Ambient Particulate Pollutants in the Ultrafine Range Promote Early  
519 Atherosclerosis and Systemic Oxidative Stress. *Circ Res* 102:589–596.

520 Argüelles S., S. Garcia, M. Maldonado, A. Machado, and A. Ayala. 2004. Do the  
521 serum oxidative stress biomarkers provide a reasonable index of the general  
522 oxidative stress status? *Biochim Biophys Acta* 1674:251–259.

523 Ayala A., M.F. Munoz, and S. Arguelles. 2014. Lipid peroxidation: production,  
524 metabolism, and signaling mechanisms of Malondialdehyde and 4-Hydroxy-2-  
525 Nonenal. *Oxid Med Cell Longev* 2014:360438.

526 Bize P., G. Devevey, P. Monaghan, B. Doligez, and P. Christe. 2008. Fecundity and  
527 survival in relation to resistance to oxidative stress in a free-living bird. *Ecology*  
528 89:2584–2593.

529 Blount J.D., E.I.K. Vitikainen, I. Stott, and M.A. Cant. 2015. Oxidative shielding and  
530 the cost of reproduction. *Biol Rev* 91, 483–497.

531 Blount J.D., P.F. Surai, R.G. Nager, D.C. Houston, A.P. Moller, M.L. Trewby, and  
532 M.W. Kennedy. 2002. Carotenoids and egg quality in the lesser blackbacked  
533 gull *Larus fuscus*: a supplemental feeding study of maternal effects. *Proc R Soc*  
534 *B-Biol Sci* 269:29–36.

535 Boulinier T. and V. Staszewski. 2008. Maternal transfer of antibodies: raising  
536 immuno-ecology issues. *Trends Ecol Evol* 23:282–288.

537 Burley N. 1986. Sexual Selection for Aesthetic Traits in Species with Biparental Care.  
538 *Am Nat* 127:415–445.

539 Burton G.J. and E. Jauniaux. 2011. Oxidative stress. *Best Pract Res Clin Obstet*  
540 *Gynaecol* 25:287–299.

541 Chen Q., M.H. Yan, Z.H. Cao, X. Li, Y.F. Zhang, J.C. Shi, G.H. Feng, et al. 2016.  
542 Sperm tsRNAs contribute to intergenerational inheritance of an acquired  
543 metabolic disorder. *Science* 351:397–400.

544 Costantini D., G. Casasole, H. AbdElgawad, H. Asard, and M. Eens. 2016.  
545 Experimental evidence that oxidative stress influences reproductive decisions.  
546 *Funct Ecol* 30:1169–1174.

547 Crean A.J. and R. Bonduriansky. 2014. What is a paternal effect? *Trends Ecol Evol*  
548 29:554–559.

549 Del Rio D., A.J. Stewart, and N. Pellegrini. 2005. A review of recent studies on  
550 malondialdehyde as toxic molecule and biological marker of oxidative stress.  
551 *Nutr Metab Cardiovasc Dis* 15:316–328.

552 Dupoué A., P. Blaimont, D. Rozen-Rechels, M. Richard, S. Meylan, J. Clobert, D.B.  
553 Miles, et al. 2020. Water availability and temperature induce changes in  
554 oxidative status during pregnancy in a viviparous lizard. *Funct Ecol* 34:475–  
555 485.

556 Fowler M.A., M. Paquet, V. Legault, A.A. Cohen, and T.D. Williams. 2018.  
557       Physiological predictors of reproductive performance in the European Starling  
558       (*Sturnus vulgaris*). *Front Zool* 15:45.

559 Galván I. and C. Alonso-Alvarez. 2009. The expression of melanin-based plumage is  
560       separately modulated by exogenous oxidative stress and a melanocortin. *Proc R*  
561       *Soc B-Biol Sci* 276:3089–3097.

562 Gavrioliouk D. and R.J. Aitken. 2015. Damage to sperm DNA mediated by reactive  
563       oxygen species: its impact on human reproduction and the health trajectory of  
564       offspring. In *Advances in Experimental Medicine and Biology*, pp. 23–47.  
565       Springer International Publishing.

566 Gil D., J. Graves, N. Hazon, and A. Wells. 1999. Male attractiveness and differential  
567       testosterone investment in zebra finch eggs. *Science* 286:126–128.

568 Giordano M., D. Costantini, J.L. Pick, and B. Tschirren. 2015. Female oxidative  
569       status, egg antioxidant protection and eggshell pigmentation: a supplemental  
570       feeding experiment in great tits. *Behav Ecol Sociobiol* 69:777–785.

571 Gluckman P.D., M.A. Hanson, and H.G. Spencer. 2005. Predictive adaptive responses  
572       and human evolution. *Trends Ecol Evol* 20:527–533.

573 Gowaty P.A. 2008. Reproductive compensation. *J Evol Biol* 21:1189–1200.

574 Grafen, A. 1988 On the uses of data on lifetime reproductive success. In *Reproductive*  
575       *success* (ed. T. Clutton-Brock), pp. 454–471. Chicago, IL: University of  
576       Chicago Press.



577 Groothuis T.G.G. and H. Schwabl. 2008. Hormone-mediated maternal effects in  
578 birds: mechanisms matter but what do we know of them? *Phil Trans R Soc B*  
579 363:1647–1661.

580 Groothuis T.G.G., W. Müller, N. von Engelhardt, C. Carere, and C. Eising. 2005.  
581 Maternal hormones as a tool to adjust offspring phenotype in avian species.  
582 *Neurosci Biobehav Rev* 29:329–352.

583 Grune T., K. Krämer, P.P. Hoppe, and W. Siems. 2001. Enrichment of eggs with n-3  
584 polyunsaturated fatty acids: Effects of vitamin E supplementation. *Lipids*  
585 36:833–838.

586 Halliwell B.H. and J.M.C. Gutteridge. 2007. Free radicals in biology and medicine.  
587 4th ed. Oxford University Press, Oxford.

588 Hamlin H.J. and L.J. Guillette. 2011. Embryos as Targets of Endocrine Disrupting  
589 Contaminants in Wildlife. *Birth Defects Res C* 93:19–33.

590 Heidinger B.J. and R.C. Young. 2020. Cross-Generational Effects of Parental Age on  
591 Offspring Longevity: Are Telomeres an Important Underlying Mechanism?  
592 *Bioessays* 42:1900227.

593 Hulbert A.J., R. Pamplona, R. Buffenstein, and W.A. Buttermer. 2007. Life and  
594 death: metabolic rate, membrane composition, and life span of animals. *Physiol*  
595 *Rev* 87:1175–1213.

596 Immler S. 2018. The sperm factor: paternal impact beyond genes. *Heredity* 121:239–  
597 247.

598 Isaksson C. 2010. Pollution and Its Impact on Wild Animals: A Meta-Analysis on  
599 Oxidative Stress. *EcoHealth* 7:342–350.

600 Jiménez-Chillarón J.C., M.J. Nijland, A.A. Ascensão, V.A. Sardão, J. Magalhães,  
601 M.J. Hitchler, F.E. Domann, and P.J. Oliveira. 2015. Back to the future:  
602 transgenerational transmission of xenobiotic-induced epigenetic remodeling.  
603 *Epigenetics* 10:259–273.

604 Kalmbach K.H., D.M.F. Antunes, R.C. Dracxler, T.W. Knier, M.L. Seth-Smith, F.  
605 Wang, L. Liu, and D.L. Keefe. 2013. Telomeres and human reproduction. *Fertil*  
606 *Steril* 99:23–29.

607 Krawetz S.A. 2005. Paternal contribution: new insights and future challenges. *Nat*  
608 *Rev Genet* 6:633–642.

609 Lane M., N.O. McPherson, T. Fullston, M. Spillane, L. Sanderman, W.X. Kang, and  
610 D.L. Zander-Fox. 2014. Oxidative stress in mouse sperm impairs embryo  
611 development, fetal growth and alters adiposity and glucose regulation in female  
612 offspring. *Plos One* 9:e100832.

613 Lindström J. 1999. Early development and fitness in birds and mammals. *Trends Ecol*  
614 *Evol* 14:343–348.

615 Losdat S., F. Helfenstein, J.D. Blount, V. Marri, L. Maronde, and H. Richner. 2012.  
616 Nestling erythrocyte resistance to oxidative stress predicts fledging success but  
617 not local recruitment in a wild bird. *Biol Lett* 9:20120888.

618 Margaritelis N.V., A.S. Veskokoukis, V. Paschalis, I.S. Vrabas, K. Dipla, A. Zafeiridis,  
619 A. Kyparos, and M.G. Nikolaidis. 2015. Blood reflects tissue oxidative stress: a  
620 systematic review. *Biomarkers* 20:97–108.

621 Marshall D.J. and T. Uller. 2007. When is a maternal effect adaptive? *Oikos*  
622 116:1957–1963.

623 Martinez de Toda I., C. Vida, A. Garrido, and M. De la Fuente. 2020. Redox  
624 Parameters as Markers of the Rate of Aging and Predictors of Life Span. *J*  
625 *Gerontol A-Biol* 75:613–620.

626 Mateos R. and L. Bravo. 2007. Chromatographic and electrophoretic methods for the  
627 analysis of biomarkers of oxidative damage to macromolecules (DNA, lipids,  
628 and proteins). *J Sep Sci* 30:175–191.

629 Menezo Y.J.R., E. Silvestris, B. Dale, and K. Elder. 2016. Oxidative stress and  
630 alterations in DNA methylation: two sides of the same coin in reproduction.  
631 *33:668–683*.

632 Messina S., M. Eens, G. Casasole, H. AbdElgawad, H. Asard, R. Pinxten, and D.  
633 Costantini. 2017. Experimental inhibition of a key cellular antioxidant affects  
634 vocal communication. *Funct Ecol* 31:1101–1110.

635 Metcalfe N.B. and C. Alonso-Alvarez. 2010. Oxidative stress as a life-history  
636 constraint: the role of reactive oxygen species in shaping phenotypes from  
637 conception to death. *Funct Ecol* 24:984–996.

638 Metcalfe N.B. and P. Monaghan. 2001. Compensation for a bad start: grow now, pay  
639 later? *Trends Ecol Evol* 16:254–260.

- 640 Mohiti-Asli M., F. Shariatmadari, H. Lotfollahian, and M.T. Mazuji. 2008. Effects of  
641 supplementing layer hen diets with selenium and vitamin E on egg quality, lipid  
642 oxidation and fatty acid composition during storage. *Can J Anim Sci* 88:475–  
643 483.
- 644 Monaghan P. 2008. Early growth conditions, phenotypic development and  
645 environmental change. *Phil Trans R Soc B* 363:1635–1645.
- 646 Monaghan P., N.B. Metcalfe, and R. Torres. 2009. Oxidative stress as a mediator of  
647 life history trade-offs: mechanisms, measurements and interpretation. *Ecol Lett*  
648 12:75–92.
- 649 Mousseau T.A. and C.W. Fox. 1998. The adaptive significance of maternal effects.  
650 *Trends Ecol Evol* 13:403–407.
- 651 Müller C., S. Jenni-Eiermann, and L. Jenni. 2009. Effects of a short period of elevated  
652 circulating corticosterone on postnatal growth in free-living Eurasian kestrels  
653 *Falco tinnunculus*. *J Exp Biol* 212:1405–1412.
- 654 Nair U., H. Bartsch, and J. Nair. 2007. Lipid peroxidation-induced DNA damage in  
655 cancer-prone inflammatory diseases: A review of published adduct types and  
656 levels in humans. *Free Rad Biol Med*. 43:1109–1120.
- 657 Napierala M., J. Mazela, T.A. Merritt, and E. Florek. 2016. Tobacco smoking and  
658 breastfeeding: Effect on the lactation process, breast milk composition and  
659 infant development. A critical review. *Environ Res* 151:321–338.

660 Napierala M., T.A. Merritt, I. Miechowicz, K. Mielnik, J. Mazela, and E. Florek.  
661 2019. The effect of maternal tobacco smoking and second-hand tobacco smoke  
662 exposure on human milk oxidant-antioxidant status. *Environ Res* 170:110–121.

663 Newman T.A.C., C.R. Carleton, B. Leeke, M.B. Hampton, and J.A. Horsfield. 2015.  
664 Embryonic oxidative stress results in reproductive impairment for adult  
665 zebrafish. *Redox Biol* 6:648–655.

666 Noguera J.C., S.-Y. Kim, and A. Velando. 2012. Pre-fledgling oxidative damage  
667 predicts recruitment in a long-lived bird. *Biol Lett* 8:61–63.

668 Nussey D.H., J.M. Pemberton, J.G. Pilkington, and J.D. Blount. 2009. Life history  
669 correlates of oxidative damage in a free-living mammal population. *Funct Ecol*  
670 23:809–817.

671 Pick J.L., P. Hutter, and B. Tschirren. 2016. In search of genetic constraints limiting  
672 the evolution of egg size: direct and correlated responses to artificial selection  
673 on a prenatal maternal effector. *Heredity* 116:542–549.

674 Pick J.L., P. Hutter, and B. Tschirren. 2017. Divergent artificial selection for female  
675 reproductive investment has a sexually concordant effect on male reproductive  
676 success. *Evol Lett* 1:222–228.

677 Qvarnstrom A. and T.D. Price. 2001. Maternal effects, paternal effects and sexual  
678 selection. *Trends Ecol Evol* 16:95–100.

679 R Development Core Team. 2014. R: A Language and Environment for Statistical  
680 Computing. R Foundation for Statistical Computing, Vienna, Austria.

681 Remes V. 2011. Yolk androgens in great tit eggs are related to male attractiveness,  
682 breeding density and territory quality. *Behav Ecol Sociobiol* 65:1257–1266.

683 Romero-Haro A.A. and C. Alonso-Alvarez. 2014. Covariation in oxidative stress  
684 markers in the blood of nestling and adult birds. *Physiol Biochem Zool* 87:353–  
685 362.

686 Rossner P., A. Milcova, H. Libalova, Z. Novakova, J. Topinka, I. Balascak, and R.J.  
687 Sram. 2009. Biomarkers of exposure to tobacco smoke and environmental  
688 pollutants in mothers and their transplacental transfer to the foetus. Part II.  
689 Oxidative damage. *Mutat Res-Fund Mol M* 669:20–26.

690 Royle N.J., P.F. Surai, and I.R. Hartley. 2001. Maternally derived androgens and  
691 antioxidants in bird eggs: complementary but opposing effects? *Behav Ecol*  
692 12:381–385.

693 Saino N., V. Bertacche, R.P. Ferrari, R. Martinelli, A.P. Moller, and R. Stradi. 2002.  
694 Carotenoid concentration in barn swallow eggs is influenced by laying order,  
695 maternal infection and paternal ornamentation. *Proc R Soc B-Biol Sci*  
696 269:1729–1733.

697 Schieber M. and N.S. Chandel. 2014. ROS Function in Redox Signaling and  
698 Oxidative Stress. *Current Biol* 24:R453–R462.

699 Silvestro S., V. Calcaterra, G. Pelizzo, P. Bramanti, and E. Mazzon. 2020. Prenatal  
700 Hypoxia and Placental Oxidative Stress: Insights from Animal Models to  
701 Clinical Evidences. *Antioxidants* 9:414.

702 Speakman J.R. and C. Selman. 2011. The free-radical damage theory: Accumulating  
703 evidence against a simple link of oxidative stress to ageing and lifespan.  
704 *Bioessays* 33:255–259.

705 Stier A., S. Reichert, S. Massemin, P. Bize, and F. Criscuolo. 2012. Constraint and  
706 cost of oxidative stress on reproduction: correlative evidence in laboratory mice  
707 and review of the literature. *Front Zool* 9:37.

708 Surai P.F., V.I. Fisinin, and F. Karadas. 2016. Antioxidant systems in chick embryo  
709 development. Part 1. Vitamin E, carotenoids and selenium. *Anim Nutr* 2:1–11.

710 Tremellen K. 2008. Oxidative stress and male infertility—a clinical perspective. *Hum*  
711 *Reprod Update* 14:243–258.

712 Tunc O. and K. Tremellen. 2009. Oxidative DNA damage impairs global sperm DNA  
713 methylation in infertile men. *J Assist Reprod Genet* 26:537–544.

714 Vágási C.I., O. Vincze, L. Pátraş, G. Osváth, J. Péntzes, M.F. Haussmann, Z. Barta,  
715 and P.L. Pap. 2019. Longevity and life history coevolve with oxidative stress in  
716 birds. *Funct Ecol* 33:152–161.

717 Valko M., D. Leibfritz, J Moncol, M.T.D. Cronin, M. Mazur, and J. Telser. 2007.  
718 Free radicals and antioxidants in normal physiological functions and human  
719 disease. *Int J Biochem Cell Biol* 39:44–84.

720 van de Crommenacker J., M. Hammers, J. van der Woude, M. Louter, P. Santema,  
721 D.S. Richardson, and J. Komdeur. 2017. Oxidative status and fitness  
722 components in the Seychelles warbler. *Funct Ecol* 31:1210–1219.

723 Velando A., R. Torres, and C. Alonso-Alvarez. 2008. Avoiding bad genes:  
724 oxidatively damaged DNA in germ line and mate choice. *Bioessays* 30:1212–  
725 1219.

726 Viblanc V.A., Q. Schull, J.D. Roth, J. Rabdeau, C. Saraux, P. Uhlrich, F. Criscuolo,  
727 and F.S. Dobson. 2018. Maternal oxidative stress and reproduction: Testing the  
728 constraint, cost and shielding hypotheses in a wild mammal. *Funct Ecol* 32:722–  
729 735.

730 Vitikainen E.I.K., M.A. Cant, J.L. Sanderson, C. Mitchell, H.J. Nichols, H.H.  
731 Marshall, F.J. Thompson, et al. 2016. Evidence of Oxidative Shielding of  
732 Offspring in a Wild Mammal. *Front Ecol Evol* 4:58.

733 Williamson K.A., P.F. Surai, and J.A. Graves. 2006. Yolk antioxidants and mate  
734 attractiveness in the Zebra Finch. *Funct Ecol* 20:354–359.

735 Xavier M.J., S.D. Roman, R.J. Aitken, and B. Nixon. 2019. Transgenerational  
736 inheritance: how impacts to the epigenetic and genetic information of parents  
737 affect offspring health. *Hum Reprod Update* 25:519–541.

738



739 **Figure legends**

740 Figure 1. Association between the number of offspring produced by the focal female  
741 during a reproductive event and a) her own plasma MDA levels during reproduction,  
742 b) her mother's plasma MDA levels when the focal female was conceived, and c) her  
743 father's plasma MDA levels when the focal female was conceived. A regression line  
744 is presented for significant associations.

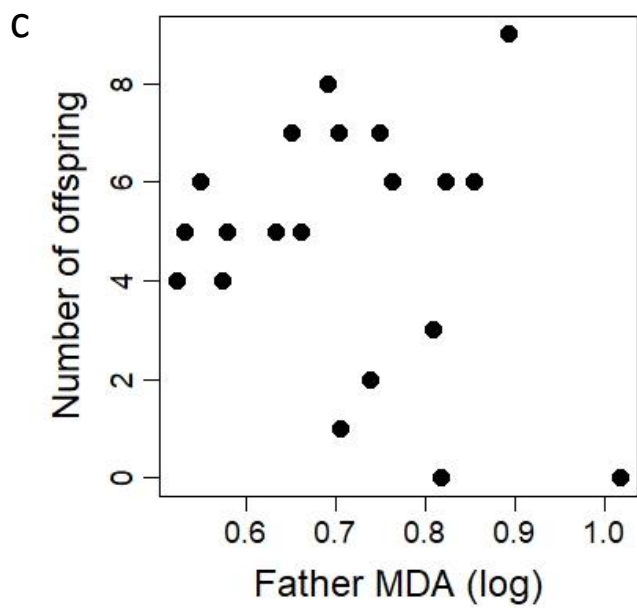
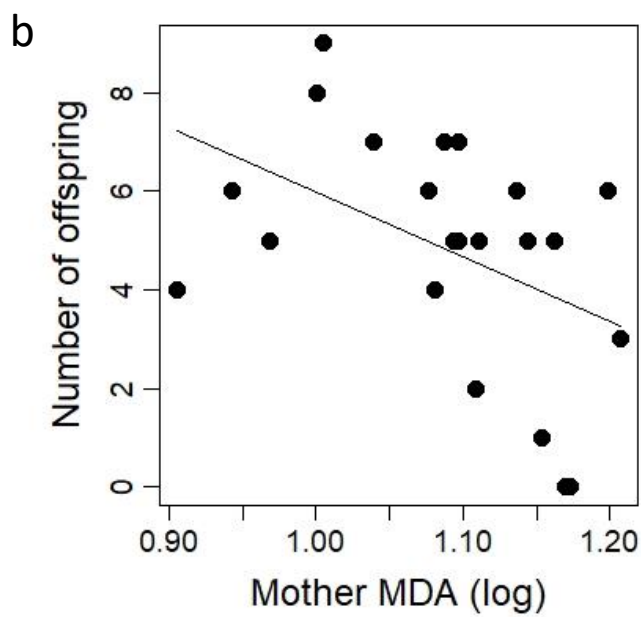
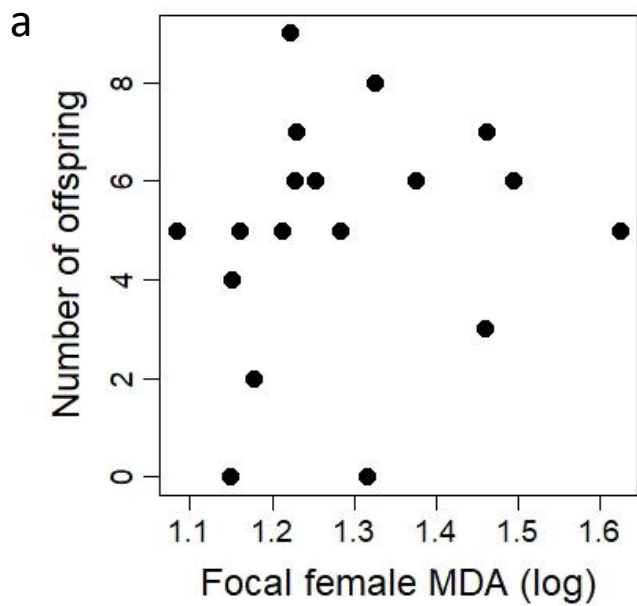
745

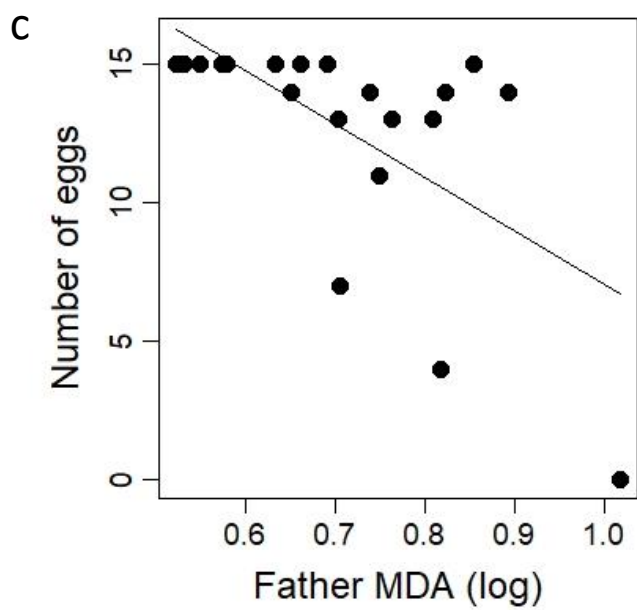
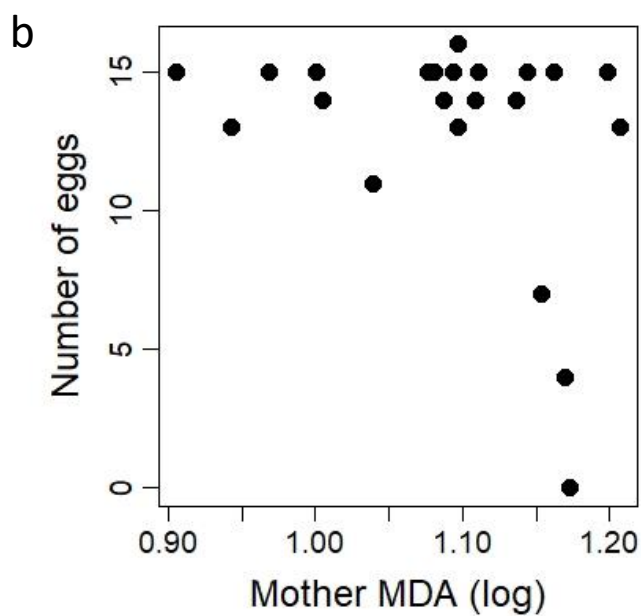
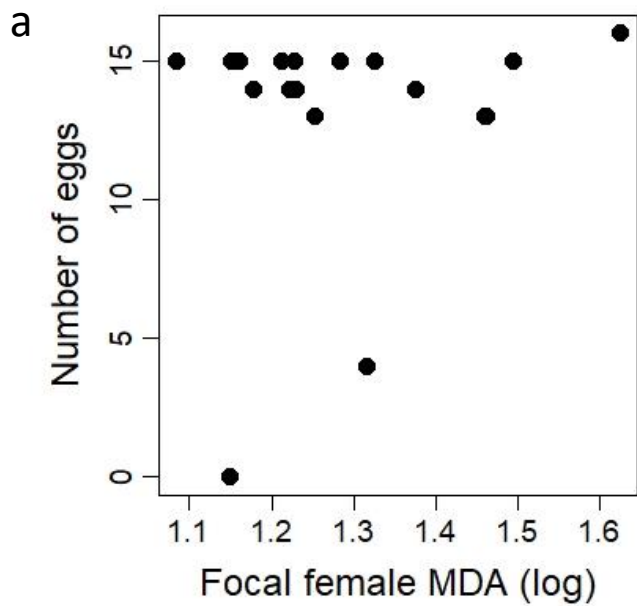
746 Figure 2. Association between the number of eggs laid by the focal female during a  
747 reproductive event and a) her own plasma MDA levels during reproduction, b) her  
748 mother's plasma MDA levels when the focal female was conceived, and c) her  
749 father's plasma MDA levels when the focal female was conceived. A regression line  
750 is presented for significant associations.

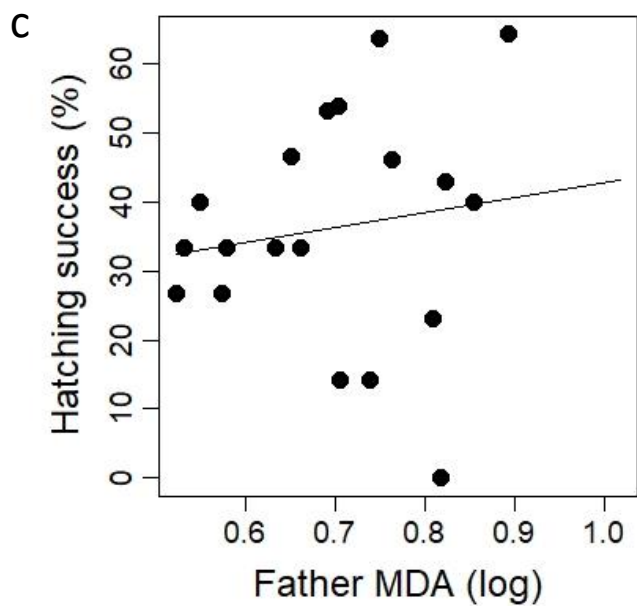
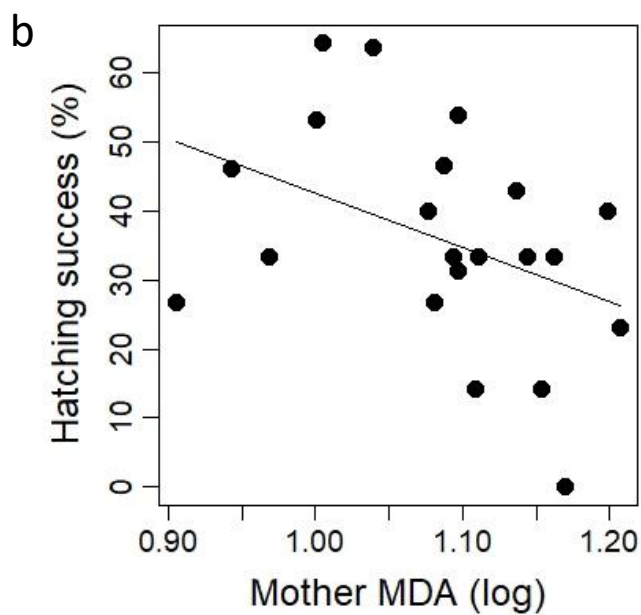
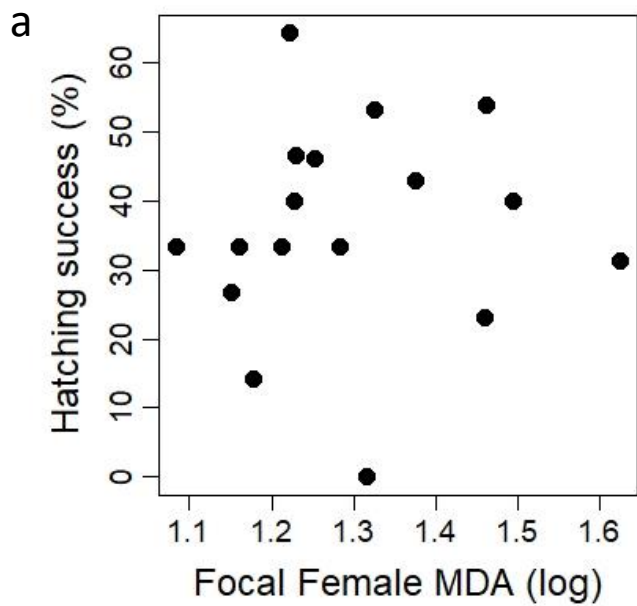
751

752 Figure 3. Association between the hatching success of the eggs laid by the focal  
753 female and a) her plasma MDA levels during reproduction, b) her mother's plasma  
754 MDA levels when the focal female was conceived, and c) her father's plasma MDA  
755 levels when the focal female was conceived. A regression line is presented for  
756 significant associations.

757







**Supplementary material for:**

**Inter-generational costs of oxidative stress: reduced fitness in daughters of mothers that experienced high levels of oxidative damage during reproduction**

Ana Ángela Romero-Haro<sup>1\*</sup>, Lorenzo Pérez-Rodríguez<sup>2</sup> & Barbara Tschirren<sup>1</sup>

1 *Supplementary analysis S1. Analyses of focal female fitness using MDA controlled for*  
2 *circulating levels of triglycerides as explanatory variables*

3 Plasma MDA levels have been shown to positively correlate with plasma triglyceride levels  
4 across bird species (Pérez-Rodríguez et al. 2015). This is expected since triglycerides are the  
5 main form of storage and transport of polyunsaturated fatty acids, which are the main target  
6 of lipid peroxidation (Mateos and Bravo 2007). So, it has been recommended to report both  
7 absolute and relative (controlled for circulating triglyceride levels) MDA levels (Romero-  
8 Haro and Alonso-Alvarez 2014; Pérez-Rodríguez et al. 2015).

9 In addition, differences in absolute MDA levels between sexes and generations (see x-axis of  
10 the main text figures 1-3) were found. Since all individuals in this study, males and females,  
11 as well as parents and daughters, were reared under the same standardized conditions, such  
12 differences could potentially be explained by variation in circulating plasma triglycerides  
13 levels. To confirm the patterns presented in the main text and test if differences in triglyceride  
14 levels explain the sex and generational differences in MDA we repeated the analyses  
15 presented in the main text using triglyceride-controlled MDA levels as explanatory variables  
16 instead of absolute MDA levels.

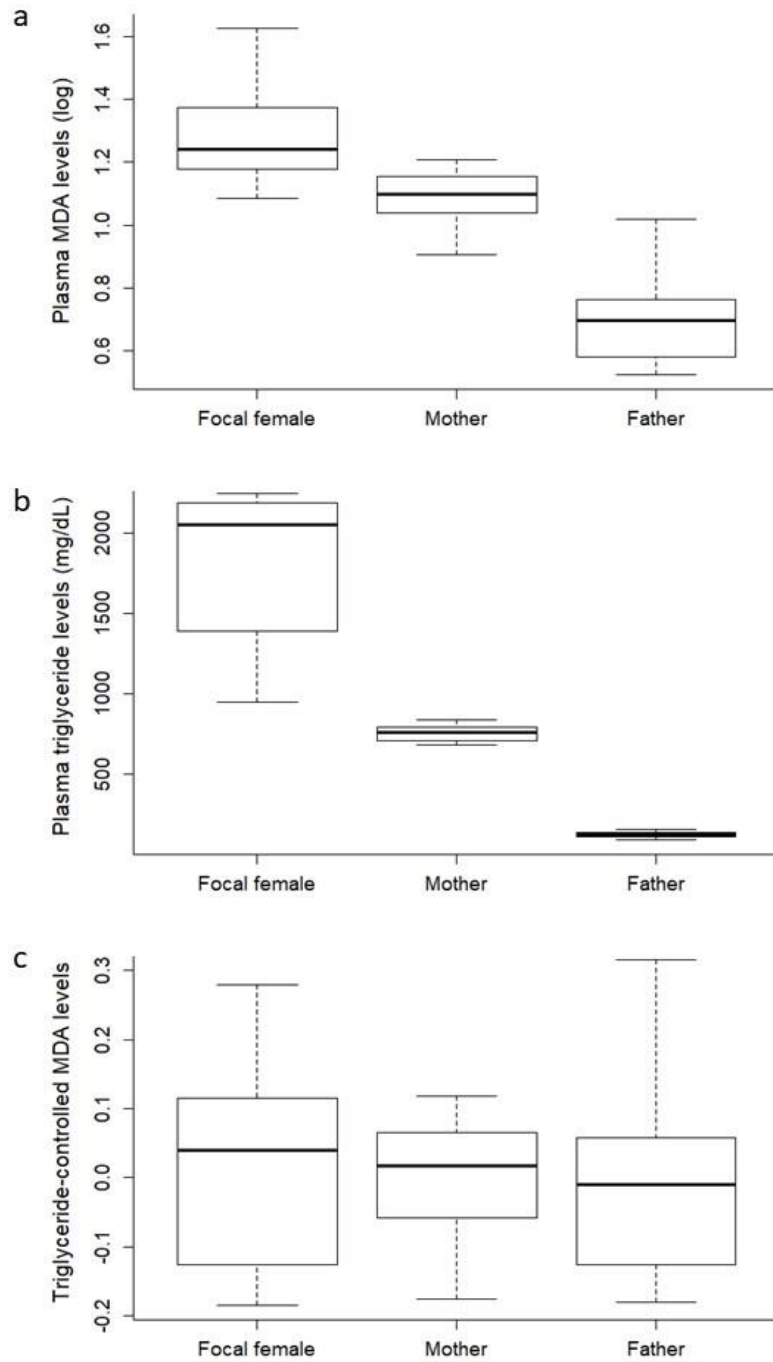
17 To this end, we quantified triglyceride concentrations in all plasma samples using the  
18 glycerol phosphate oxidase/oxidase method. We used a commercial kit (Biosystems,  
19 Barcelona) and followed the manufacturer's instructions. Repeatability of triglyceride  
20 measures was high ( $r = 0.98$ ,  $n = 20$ ,  $P < 0.001$ ). Due to limited plasma samples, triglyceride  
21 levels were not measured in two samples of fathers, resulting in lower sample sizes for some  
22 comparisons.

23 In order to check if triglyceride levels predict MDA levels across all the samples, we ran a  
24 linear model including triglyceride levels as a fixed factor. We found that plasma triglyceride

25 levels were positively related to plasma MDA levels ( $\beta \pm \text{SE}$ :  $0.0003 \pm 0.00002$ ;  $t_{1,56} =$   
26  $11.18$ ,  $P < 0.0001$ ). We then obtained triglyceride-controlled MDA levels separately for each  
27 group (i.e. focal females, mothers and fathers). Triglyceride-controlled MDA was defined as  
28 the residuals of a linear model with MDA levels as response variable and triglyceride level as  
29 predictor.

30 To explore if the differences in MDA levels between focal females, mothers and fathers  
31 observed in the figures of the main text were caused by differences in triglyceride levels, we  
32 ran three linear models including MDA, triglycerides or triglyceride-controlled MDA levels  
33 as response variables and the group of analysis (a three-level factor representing focal  
34 females, mothers or fathers) as a predictor. The post-hoc comparisons were performed by  
35 Tukey tests. We found that MDA and triglyceride levels differed between groups (MDA:  $F_{2,55} =$   
36  $113.29$ ,  $P < 0.001$ , fig. S1a; triglycerides:  $F_{2,55} = 248.92$ ,  $P < 0.001$ , fig. S1b). MDA and  
37 triglyceride levels in focal females were higher than those in mothers and fathers, and levels  
38 in mothers were higher than in fathers (all P-values  $< 0.001$ ). In contrast, triglyceride-  
39 controlled MDA levels did not differ among groups ( $F_{2,55} = 0.021$ ,  $P = 0.979$ , fig. S1c, all P  
40 values of post-hoc comparisons  $> 0.853$ ). This suggests that differences in MDA levels  
41 observed across groups are caused by differences in triglyceride levels (fig. S1). We show the  
42 mean, SE and range of the variables in the table S1.

43



44

45 Figure S1: Focal female, mother and father plasma levels of: a) MDA (i.e. oxidative damage in  
 46 lipids), b) triglycerides and c) triglyceride-controlled MDA. The midline of the boxes represents  
 47 median values, and the lower and upper edges of the boxes represent the first and third quartiles,  
 48 respectively. Whiskers represent greatest and lowest values.

49

50

51

52



53 Table S1: Triglyceride and MDA plasma levels of focal females, mothers and fathers included in this  
54 study.

55

	<b>Mean</b>	<b>SE</b>	<b>Range</b>
<b>Focal females</b>			
Triglycerides (mg/dL)	1873	100.2	947.4 – 2246.7
MDA ( $\mu$ M)	20.6	1.82	12.1 – 42.1
Log-transformed MDA	1.29	0.03	1.08 – 1.62
Triglyceride-controlled MDA	0	0.03	-0.017 – 0.029
<b>Mothers</b>			
Triglycerides (mg/dL)	758	10.2	680.9 – 839.5
MDA ( $\mu$ M)	12.5	0.48	8.05 – 16.1
Log-transformed MDA	1.09	0.02	0.91 – 1.21
Triglyceride-controlled MDA	0	0.02	-0.16 – 0.11
<b>Fathers</b>			
Triglycerides (mg/dL)	123.6	4.83	90.8 – 160.7
MDA ( $\mu$ M)	5.41	0.39	3.34 – 10.4
Log-transformed MDA	0.71	0.03	0.52 – 1.02
Triglyceride-controlled MDA	0	0.03	-0.18 – 0.031

56

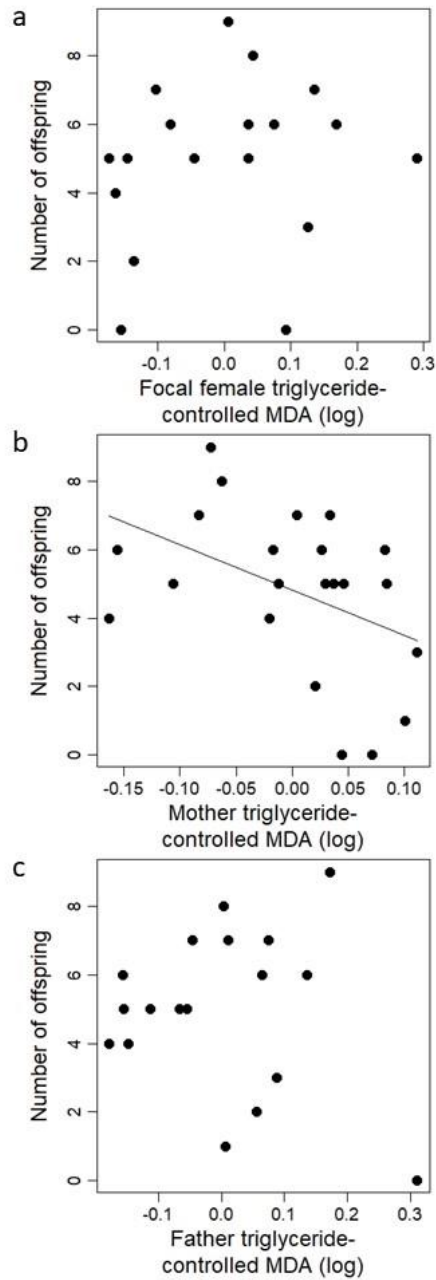
57

58

59 What could cause differences in triglyceride levels across groups? Differences between sexes  
60 could have been originated because of sex differences in the metabolism, use or storage of  
61 lipids for reproduction (Lawrence and Riddle 1916; Riddle and Burns 1927). In other words,  
62 females may need higher circulating levels of triglycerides to produce eggs, which means  
63 higher levels of easily oxidizable substrate (Romero-Haro and Alonso-Alvarez 2014; Perez-  
64 Rodriguez et al. 2015; Alonso-Alvarez et al. 2017). Indeed, males showing lower levels of  
65 circulating lipids than females has been reported previously in birds (Riddle and Burns 1927),  
66 and we have found the same result in another population of quail during reproduction  
67 (unpublished data). Regarding the differences between generations, focal females showing  
68 higher triglyceride levels than mothers, the difference may be caused, for example, by slight  
69 differences between food batches generated by the manufacturer (note that mother and  
70 daughter samples were collected at different time points).

71 Considering the results above, to confirm the patterns presented in the main text (where  
72 absolute MDA levels were used in the analyses) we repeated the same models reported in the  
73 main text but using triglyceride-controlled MDA levels instead of absolute MDA levels as  
74 explanatory variables.

75 In agreement with the results presented in the main text, the number of offspring produced by  
76 a focal female were neither explained by her own triglyceride-controlled MDA levels ( $t_{1,15} =$   
77  $1.182$ ,  $P = 0.256$ , Fig. S2a) nor the father's triglyceride-controlled MDA levels ( $t_{1,10} = -$   
78  $0.370$ ,  $P = 0.719$ , Fig. S2c). However, we again found a negative association with  
79 triglyceride-controlled MDA of the mother during reproduction ( $\beta \pm 1SE: -13.307 \pm 6.188$ ;  $t$   
80  $_{1,20} = -2.150$ ,  $P = 0.044$ , Fig. S2b).

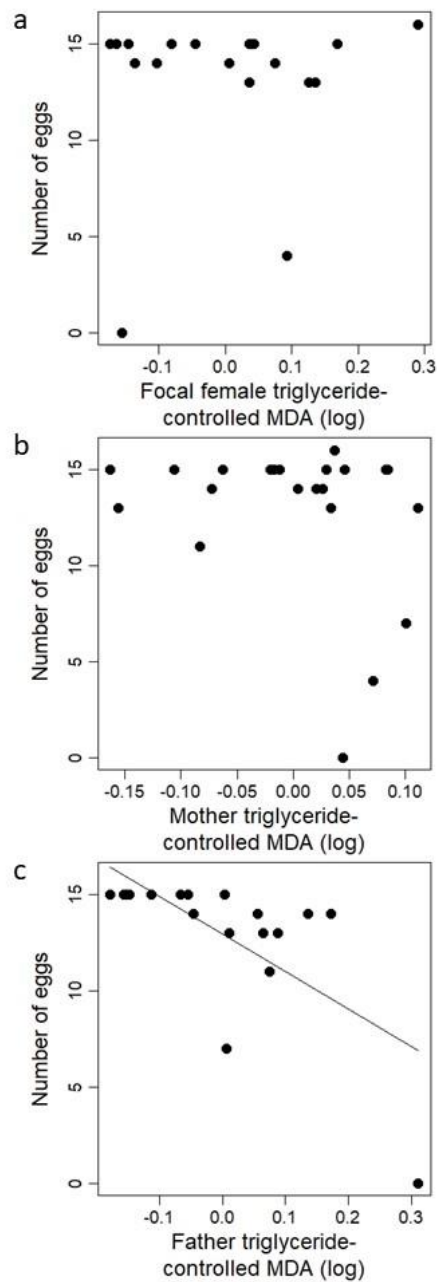


81

82 Figure S2. Association between the number of offspring produced during a reproductive event by the  
 83 focal female and a) her plasma triglyceride-controlled MDA levels during reproduction, b) her  
 84 mother's plasma triglyceride-controlled MDA levels when the focal female was conceived, and c) her  
 85 father's plasma triglyceride-controlled MDA levels when the focal female was conceived. A  
 86 regression line is presented for significant associations.

87

88 Regarding the number of eggs laid by a focal female, the results are again the same as those  
89 presented in the main text: the number of eggs was neither predicted by the focal female's nor  
90 by her mother's triglyceride-controlled MDA levels ( $\chi^2 = 3.657$ ,  $P = 0.144$  and  $\chi^2 = 3.743$ ,  $P$   
91  $= 0.201$ , respectively; Figs. S3a, b). However, we again found a negative association with the  
92 triglyceride-controlled MDA levels of the father ( $\beta \pm 1SE = -8.372 \pm 2.242$ ;  $\chi^2 = 45.072$ ,  $P <$   
93  $0.001$ , Fig. S3c).

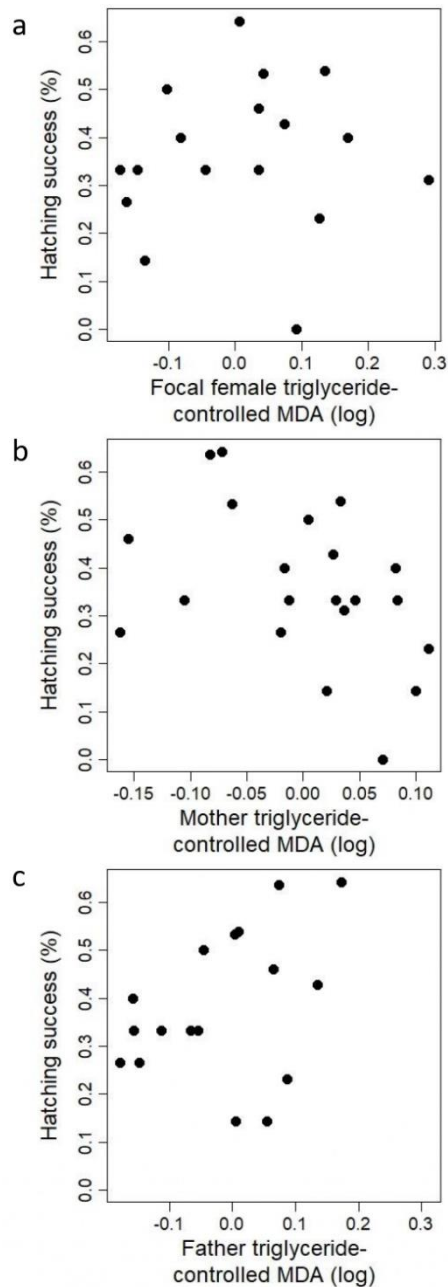


94

95 Figure S3. Association between the number of eggs laid during a reproductive event by the focal  
96 female and a) her plasma triglyceride-controlled MDA levels during reproduction, b) her mother's

97 plasma triglyceride-controlled MDA levels when the focal female was conceived, and c) her father's  
98 plasma triglyceride-controlled MDA levels when the focal female was conceived. A regression line is  
99 presented for significant associations.  
100

101 Slight differences were observed when analyzing hatching success of eggs laid by a focal  
102 female during the reproductive event. As in the main text, the hatching success was not  
103 predicted by the focal female's levels of triglyceride-controlled MDA ( $\chi^2 = 0.855$ ,  $P = 0.376$ ;  
104 Fig. S4a). Unlike in the analyses presented in the main text, also the mother's and father's  
105 levels of triglyceride-controlled MDA did not significantly predict hatching success (mother:  
106  $\chi^2 = -2.698$ ,  $P = 0.112$ ; father:  $\chi^2 = 1.369$ ,  $P = 0.246$ , respectively; Figs. S4b, c). This absence  
107 of association between father triglyceride-controlled MDA levels and the hatching success of  
108 the daughter's eggs could be explained by the lower statistical power of this analysis, as we  
109 were not able to obtain triglyceride levels (and, consequently, triglyceride-controlled MDA  
110 levels) from two males. However, this cannot explain the absence of a negative association  
111 between mother triglyceride-controlled MDA levels and hatching success of the daughter's  
112 eggs, since the sample size is the same for absolute and triglyceride-controlled MDA. This  
113 result suggests that the association found in the main text could be led by maternal  
114 triglyceride levels instead of MDA levels. Anyway, this highlights the modulating role of  
115 circulating triglyceride levels for the interpretation of lipid oxidative damage results  
116 (Romero-Haro and Alonso-Alvarez 2014; Perez-Rodriguez et al. 2015; Alonso-Alvarez et al.  
117 2017).



118

119 Figure S4. Association between the hatching success of the eggs laid by the focal female and a) her  
 120 plasma triglyceride-controlled MDA levels during reproduction, b) her mother's plasma triglyceride-  
 121 controlled MDA levels when the focal female was conceived, and c) her father's plasma triglyceride-  
 122 controlled MDA levels when the focal female was conceived.

123

124 In agreement with the results presented in the main text, lifespan of focal females was neither  
 125 explained by their own triglyceride-controlled MDA levels during reproduction ( $t_{1,10} =$   
 126  $0.075$ ,  $P = 0.942$ ) nor by those of their mother or father when they were conceived, ( $t_{1,20} = -$   
 127  $0.921$ ,  $P = 0.368$  and  $t_{1,15} = 0.646$ ,  $P = 0.733$ , respectively). There was no significant

128 association between the mother's or the father's levels of triglyceride-controlled MDA when  
129 the focal female was conceived and the size of the egg the focal female developed in (mother  
130 MDA:  $t_{1,15} = -0.186$ ,  $P = 0.855$ , father MDA:  $t_{1,16} = 1.467$ ,  $P = 0.162$ ), the focal female's  
131 body mass at hatching (mother MDA:  $t_{1,15} = -0.016$ ,  $P = 0.987$ , father MDA:  $t_{1,16} = 0.644$ ,  $P$   
132  $= 0.528$ ) or the focal female's body mass at adulthood (mother MDA:  $t_{1,17} = -0.402$ ,  $P =$   
133  $0.693$ , father MDA:  $t_{1,16} = 0.644$ ,  $P = 0.528$ ). There was neither a mother-daughter nor father-  
134 daughter resemblance in the levels of triglyceride-controlled MDA measured during  
135 reproduction ( $t_{1,16} = 0.909$ ,  $P = 0.377$  and  $t_{1,12} = 0.002$ ,  $P = 0.999$ , respectively).

136 Although the associations between hatching success and parental levels of triglyceride-  
137 controlled MDA became non-significant, overall, the results obtained from either absolute or  
138 triglyceride-controlled MDA levels are very similar. Thus, the proposed underlying  
139 mechanisms and, more importantly, the conclusions of the study do not change.

140

#### 141 ***Supplementary analyses S2. Correlation between mother and father plasma MDA levels***

142 Although mothers and fathers were randomly selected and blood sampled just before being  
143 placed in cages for breeding (i.e. before meeting each other), their levels could – by chance –  
144 be correlated. We tested for an association between maternal and paternal absolute MDA,  
145 triglyceride and triglyceride-controlled MDA levels using Pearson's correlation coefficient  
146 and standardized values. Unexpectedly, maternal and paternal levels of MDA were positively  
147 correlated ( $r = 0.454$ ,  $P = 0.044$ ,  $n = 20$ ). However, neither triglyceride levels ( $r = 0.153$ ,  $P =$   
148  $0.544$ ,  $n = 18$ ) nor triglyceride-controlled MDA levels ( $r = 0.247$ ,  $P = 0.323$ ,  $n = 18$ ) of both  
149 pair members were correlated. Since they were sampled before they were placed together in  
150 cages (i.e. before they had met), a direct influence on each other can be excluded.

151 Given the opposite effects of maternal and paternal MDA levels on daughter reproductive  
152 success, this correlation does not bias the results or conclusions presented in the main text.  
153 Specifically, the results presented in Supplementary material S1 show that the results are  
154 robust when using triglyceride-controlled MDA levels, for which no correlation between  
155 mothers and fathers is observed.

156

## 157 **References**

158 Alonso-Alvarez C., T. Canelo, and A.A. Romero-Haro. 2017. The oxidative cost of  
159 reproduction: theoretical questions and alternative mechanisms. *Bioscience* 67:258–  
160 270.

161 Lawrence J.V. and O. Riddle. 1916. Studies on the physiology of reproduction in birds VI.  
162 Sexual differences in the fat and phosphorus content of the blood of fowls. *Am J*  
163 *Physiol* 41:430–437.

164 Mateos R. and L. Bravo. 2007. Chromatographic and electrophoretic methods for the analysis  
165 of biomarkers of oxidative damage to macromolecules (DNA, lipids, and proteins). *J*  
166 *Sep Sci* 30:175–191.

167 Pérez-Rodríguez L., A.A. Romero-Haro, A. Sternalski, J. Muriel, F. Mougeot, D. Gil, and C.  
168 Alonso-Alvarez. 2015. Measuring oxidative stress: the confounding effect of lipid  
169 concentration in measures of lipid peroxidation. *Physiol Biochem Zool* 88:345–351.

170 R Development Core Team. 2014. *R: A Language and Environment for Statistical*  
171 *Computing*. R Foundation for Statistical Computing, Vienna, Austria.



- 172 Riddle O. and F.H. Burns. 1927. Studies on the physiology of reproduction in birds XXII.  
173 Blood fat and phosphorus in the sexes and their variations in the reproductive cycle.  
174 Am J Physiol 81:711–724.
- 175 Romero-Haro A.A. and C. Alonso-Alvarez. 2014. Covariation in oxidative stress markers in  
176 the blood of nestling and adult birds. Physiol Biochem Zool 87:353–362.