

Uses of carotenoid-rich ingredients to design functional foods: a review

Shahida Anusha Siddiqui^{a,b*}, Salome Dini^c, Yasaman Esmaeili^d, Sahar Roshanak^e,
Ali Ali Redha^{f,g} and Sajad Ahmad Wani^{h*}

^aTechnical University of Munich Campus Straubing for Biotechnology and Sustainability, Essigberg 3, 94315 Straubing, Germany

^bGerman Institute of Food Technologies (DIL e.V.), Prof.-von-Klitzing-Straße 7, 49610, D, Quakenbrück, Germany

^cDepartment of Food Science, University of Otago, Dunedin, New Zealand

^dDepartment of Food Science and Technology, Isfahan (Khorasgan) Branch, Islamic Azad University, Isfahan, Iran

^eDepartment of Food Science and Technology, Faculty of Agriculture, Ferdowsi University of Mashhad, Mashhad, Iran

^fThe Department of Public Health and Sport Sciences, University of Exeter Medical School, Faculty of Health and Life Sciences, University of Exeter, Exeter, EX1 2LU, United Kingdom

^gCentre for Nutrition and Food Sciences, Queensland Alliance for Agriculture and Food Innovation (QAAFI), The University of Queensland, Brisbane, QLD 4072, Australia

^hDepartment of Food Technology, Islamic University of Science and Technology, Awantipora, India

***Corresponding author:** Shahida Anusha Siddiqui, Technical University of Munich Campus Straubing for Biotechnology and Sustainability, Essigberg 3, 94315 Straubing, Germany. E-mail: S.Siddiqui@dil-ev.de; Sajad Ahmad Wani, Department of Food Technology, Islamic University of Science and Technology, Awantipora, India. E-mail: sajad04@outlook.com

DOI: 10.31665/JFB.20xx.000xx

Received: November 08, 2022; **Revised received & accepted:** December 26, 2022

Citation: Siddiqui, S.A., Dini, S., Esmaeili, Y., Roshanak, S., Redha, A.A., and Wani, S.A (2022). Uses of carotenoid-rich ingredients to design functional foods: a review. *J. Food Bioact.* 000: 000–000.

Abstract

Carotenoids are isoprenoids that are extensively dispersed in foods that have always been part of the human diet. Certain carotenoids can be transformed into retinoids with vitamin A activity, which is needed for humans. Additionally, they are far more flexible, since they may be found in foods not just as sources of vitamin A, and also as natural colors, antioxidants, and health-promoting substances. Functional foods provide health advantages in addition to basic nourishment. They can be found in a variety of forms, including whole, fortified, enriched, or enhanced meals. A flood of information about the health advantages of functional foods has been supplied by several epidemiological research. This review discusses the factor for healthy and sustainable usage of carotenoid-rich ingredients for the design of functional food products primarily intended for health promotion. Furthermore, data on sources, intakes, and variables influencing bioavailability are summarized.

Keywords: Carotenoids; Food production; Bakery products; Extruded snacks; Gluten-free products; Meat products.

1. Introduction

Carotenoids are natural substances and are biosynthesized by photosynthetic organisms (algae, plants, and cyanobacteria), fungi, and bacteria. Carotenoids are classified as xanthophylls or carotenes depending on their chemical makeup. Carotenes are made up of hydrogen and carbon atoms, whereas xanthophylls also include oxygen. Carotenoids play a part in photosynthetic activities

in plants and provide pigments. Tissues with a yellow-red tint act as an attractant in many fruits and flowers (Kalac, 2012). Carotenoids cannot be biosynthesized by the great majority of animals, but they may be integrated into the diet and structurally changed afterwards. Certain invertebrates, including hemipterans (phylloxerids, adelgids, and aphids) and dipterans (gall midges), have been demonstrated to synthesize carotenoids from scratch (Rodriguez-Concepcion et al., 2018).

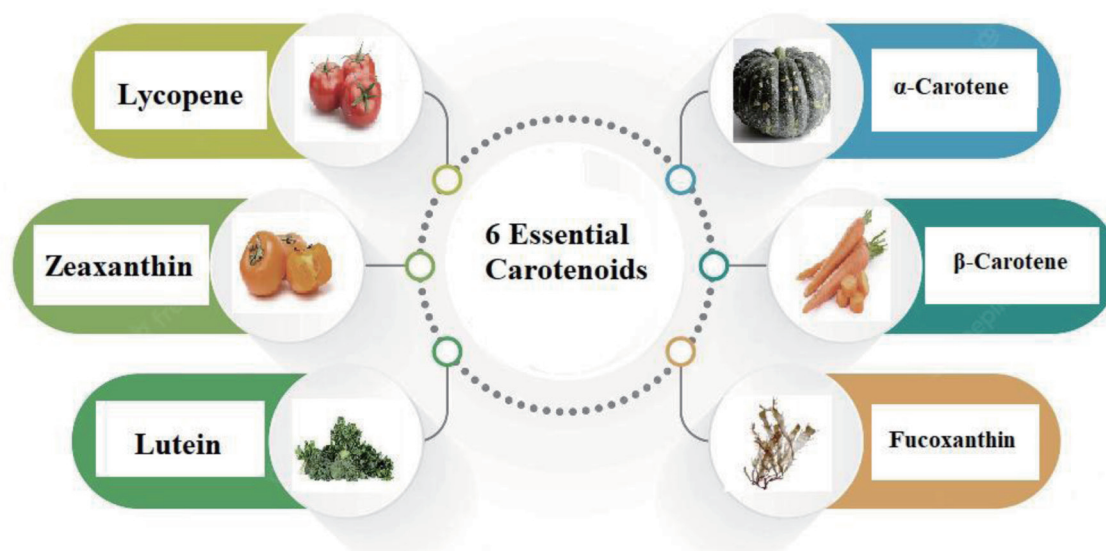


Figure 1. Essential carotenoids and sources.

To execute their biological activities, carotenoids must be released from meals and made accessible for absorption by the human body; yet, when compared to other dietary phytonutrients, they are only 5 to 30% accessible from organic resources (Kopeck and Failla, 2018; Bohn, 2018). As a result, new methods, such as digestion procedures, are guiding the development of innovative delivery systems and functional meals with improved carotenoid availability, as well as the development of technological techniques to increase carotenoid bioavailability from plant-food sources.

Carotenoids have been reported for a wide range of bioactive potentials including anti-cancer, anti-diabetic, anti-obesity, anti-inflammatory, and cardioprotective activities (van Chuyen and Eun, 2017).

Antioxidant and anti-inflammatory effects of carotenoids (Fiedor and Burda, 2014) can improve eye health, cognitive function (Bernstein et al., 2016; Feeney et al., 2013) and provitamin A activity (Conboy Stephenson et al., 2021). Carotenoids having an unsubstituted β -ionone ring, carotenes (such as α -carotene and β -carotene), and the xanthophyll β -cryptoxanthin all exhibit provitamin A activity. Vitamin A is needed for eyesight and is one of the most often deficient vitamins in the world, particularly in developing countries (Gurmu et al., 2014). Fucoxanthin is another carotenoid, a xanthophyll, that has a wide range of bioactivities. It is found mainly in brown seaweed (Meresse et al., 2020). Among its bioactivities, it has shown promising antidiabetic, anti-obesity (Maeda, 2013), antioxidant, anti-inflammatory, and anticancer activity (Meresse et al., 2020). It has been shown to be effective on many of the classical hallmarks of tumor cells. The main carotenoids that have remarkable applications is shown in Figure 1, including their main sources. The structure of key carotenoids is shown in Figure 2 (Fiedor and Burda, 2014).

It is also important to highlight that although higher plants are considered a major source of carotenoids, yet algae also have significant amounts of carotenoids (Shahidi and Brown, 1998). Blue-green algae, red algae, yellow algae, brown algae, and green algae are sources of carotenoid pigments. In addition, some animals such as crustaceans, molluscs, and fish contain a wide range of carotenoids. Salmon fish is abundant in astaxanthin which gives salmon fish the pink-red color (Helgeland et al., 2019). Among the wild salmonids, the highest amount of astaxanthin has been determined

in *Oncorhynchus* species that ranged between 26 to 38 mg/kg flesh (Ambati et al., 2014).

In recent years, the rise of functional foods has prompted significant interest in alternative dietary sources of carotenoids. Although any universally recognized description exists, functional products are entire, enriched, improved, or upgraded foods that provide health benefits. Furthermore, rather than tablets or capsules, functional foods should be ingested as full meals (Conboy Stephenson, Ross, and Stanton 2021).

This review investigates the use of carotenoid-rich components in food product manufacturing, as well as why carotenoid is an important material for the creation of functional products and how it is being employed in modern food technology research.

2. Why choose carotenoids?

Choosing a diet rich in carotenoids has been linked to a decreased risk of developing some types of cancer risk, cardiovascular disease, age-related macular impairment, and eye disease development in epidemiological studies (Coutte et al., 2017; Sharoni et al., 2012; Leong et al., 2018). Clinical indications of the conjunctiva and corneal abnormalities, such as scarring, night blindness, corneal ulceration, keratomalacia, xerophthalmia and permanent blindness, occur when carotenoids are deficient (Sommer, 2008). Aside from the aforementioned, lack of provitamin A carotenoids causes eye impairment in humans as well as higher mortality owing to impaired innate and adaptive immunity (Stephensen, 2013). Carotenoids are interesting candidates for improvement and alteration because of their widespread use in the feed, food, cosmetic, and pharmaceutical sectors. In recent decades, molecular biology and biotechnology improvements have led to a better understanding of carotenoid synthesis in microorganisms and higher plants (Potrykus, 2003).

3. Processing of carotenoid-rich ingredients

When exposed to various circumstances (e.g., light, heat, oxygen),

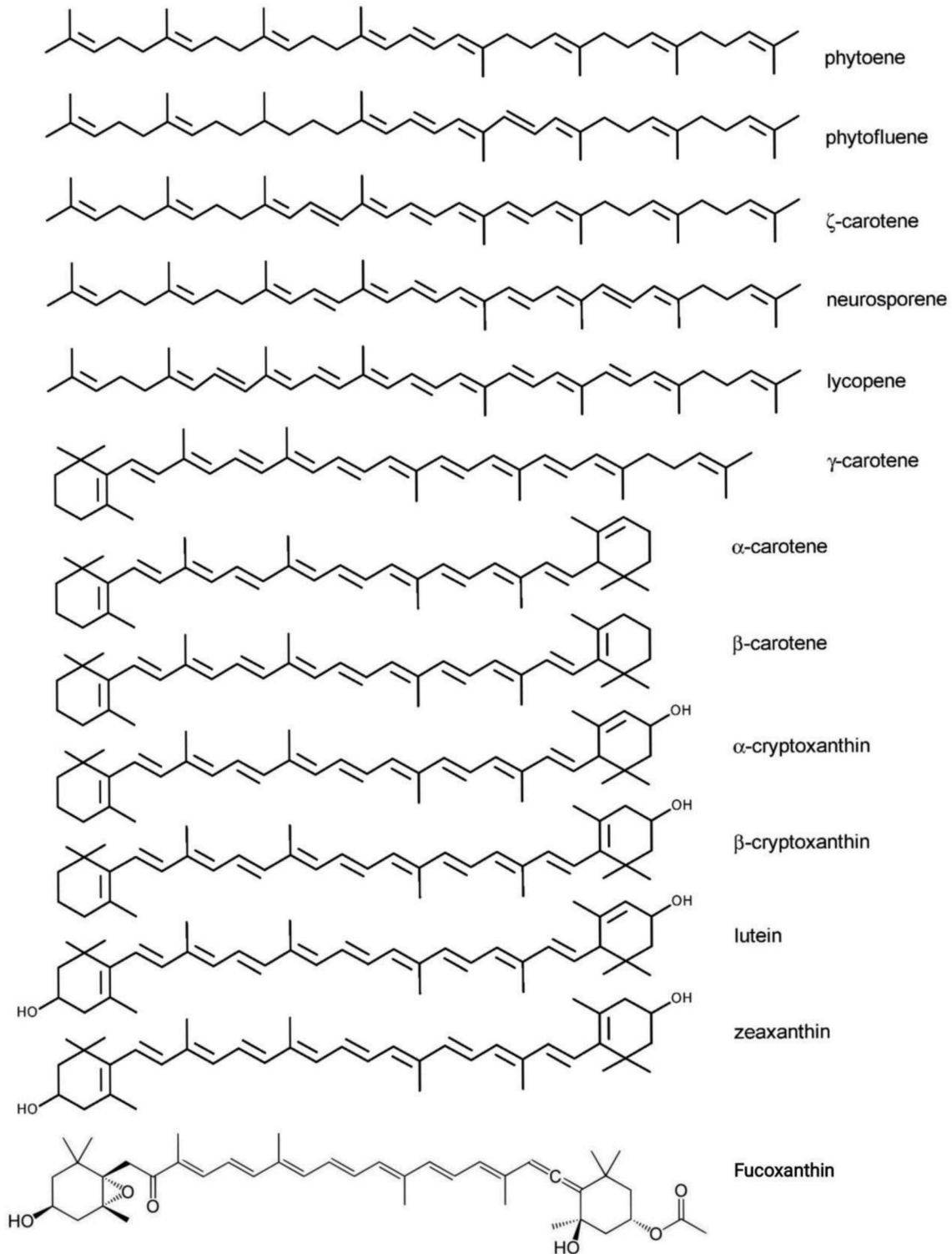


Figure 2. Chemical structure of key carotenoids.

carotenoids degrade, resulting in a loss of benefits, change in physical properties (Figure 3) and prohibition of usage in food products (Mendes-Silva et al., 2020). Their hydrophobicity makes it harder to incorporate them into water-based diets. Microencapsu-

lation methods have been used for decades to keep carotenoid-rich extracts stable throughout ordinary food processing and storage conditions. Bioactive substances are typically enclosed to protect them from degrading elements such as enzymes, heat, oxygen, and

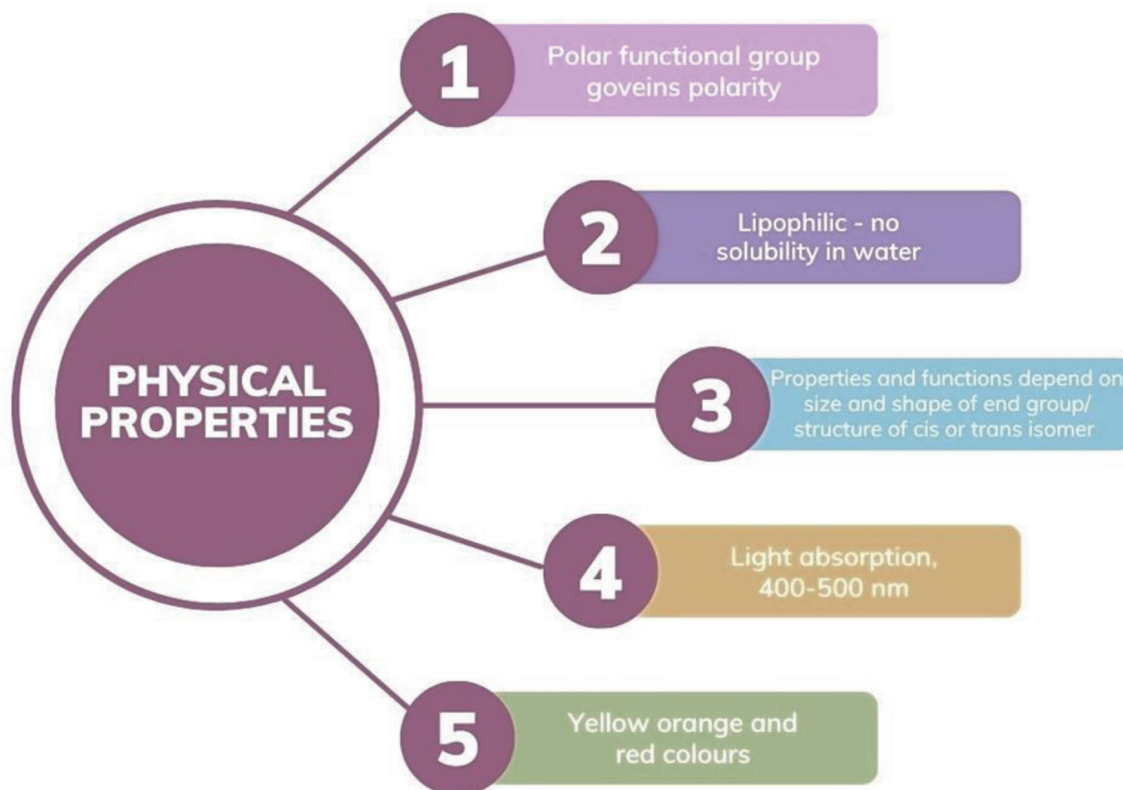


Figure 3. Physical properties of carotenoids.

light. A study by Gomez-Estaca et al. (2016) presented a one-of-a-kind study on the encapsulation of carotenoid-rich animal-derived materials. Regarding microbiological sources, *Haematococcus pluvialis* cells have been employed in several research studies to obtain astaxanthin-rich extracts, a xanthophyll carotenoid with high antioxidant activity (de Freitas Santos et al., 2021a). Because of its ease of use and scale-up, as well as its simple, rapid, and low-cost operation, spray drying is one of the earliest and most extensively used processes for dehydrating or microencapsulating food and bioactive compounds. It also has the ability to create powders from fluids, convert hydrophobic compounds into water-dispersible powders, and increase the durability of critical bioactive compounds (Bakry et al., 2016).

It is important to underline that isomerization can also influence the bioavailability and functionality of carotenoids. For instance, *E*-isomers of β -carotene have a higher bioavailability than the all-*Z*-isomer, yet the opposite is true for lycopene (Honda et al., 2018). Furthermore, different isomers of the same compound can show different bioactive potentials. Many of the *Z*-isomers of carotenoids have shown greater antioxidant activity than the all-*E*-isomers (Honda, 2021). In fact, they also have greater solubility than the all-*E* isomers in organic media and oils. Overall, it has been reported that the *Z*-isomerization of carotenoids can cause a significant change in the physicochemical properties (including solubility and crystallinity) of carotenoids, resulting in enhanced processing efficacy, bioavailability, and bioactivity (Honda, 2022).

Another research employed spray drying to reduce the hydrophobicity and susceptibility of tucum oil particles to deterioration. The study experimented different oil concentrations (100, 200, and 300 g/kg) and temperatures for drying (120 and 180 °C). Be-

cause the carotenoid content of granules dehydrated at 120 °C was greater (52.9–162.1 mg/kg) than that of granules dried at 180 °C (43.7–130.6 mg/kg), they were chosen for further study (de Freitas Santos et al., 2021b).

Red capsicum, an outstanding supply of carotenoids, has been investigated as a new source for the enzymatic liquefaction (EL) of aquatic carotenoid-rich extract (ACE). Utilizing different carbohydrates digestive enzymes: pectinase, cellulase and viscozyme L the effects of liquefaction and the ability to recover greater carotenoids in the aqueous extract were investigated. When viscozyme and pectinase were used instead of cellulase, the restoration of carotenoids and other bioactive components was significantly higher. Viscozyme at a dosage of 0.3 per cent at 60 degrees Celsius produced the greatest results. Valorization of capsicum concentrates via EL might be a promising technique for recovering important food components while decreasing processing waste and maintaining environmental sustainability through green processing (Nath et al., 2016).

4. Advancement in the carotenoid analysis

Colorimetric, thin-layer chromatography, capillary electrophoresis, paper, open-column, and fluorometric, high-performance liquid chromatography (HPLC), and spectrophotometric techniques were used to identify and quantify carotenoids in the food matrix. For characterization and recognition study, HPLC is commonly used in combination with nuclear magnetic resonance spectroscopy (NMR) and mass spectrometry (positive mode atmospheric pressure chemical ionization; APCI+ mode). HPLC is used for dif-

ferentiating *cis* isomers from the *trans* isomers throughout the processing procedure for characterization and analysis of carotenoids in biological and nutritional samples (Gupta et al., 2015; Zhao et al., 2022).

Because there are over 700 different forms of carotenoids in animals, microbes, fungi, and plants, developing and preserving pure carotenoid levels is a big difficulty in measuring. Aside from that, the complexity of dietary components, along with carotenoids' vulnerability to *cis* and *trans* isomerization and oxidation during analysis and storage, are major impediments to carotenoid assessment. Due to a shortage of inexpensive standards, many laboratories continue to struggle with carotenoid identification and quantification. As a result, improved carotenoid purification processes must be developed. Refined carotenoids are frequently used in the cosmetics sector, in addition to the food industry (Anunciato and da Rocha Filho, 2012).

A single-step application of an organic solvent to extract carotenoids from the matrix is the most common approach for collecting them for further study. Even though diol and silica capsules have only shown good retention for lutein, a far more polar carotenoid, solid-phase extraction (SPE) is rarely acknowledged in carotenoid academic research, the most popular compounds are C30 and C18 (Ferreiro-Vera et al., 2011). SPE may also be used to improve the recovery efficiency of carotenoids extracted using organic solvents. The most widely used approach for detecting and characterizing diverse oxygen-functionalized carotenoids and carotenes, including ketone hydroxyl and epoxy groups, is air pressure chemical ionization (APCI)-tandem mass spectrometry (MS/MS) in positive ion mode (Rivera and Canela-Garayoa, 2012).

Mass spectrometry is a helpful method for detecting new chemicals; fragment signals may even separate pigments with similar structures. In structural elucidation, mass spectrometry (MS) is commonly combined with proton nuclear magnetic resonance (¹H-NMR) data to produce absolute confirmatory results on highly similar isomers (Qiu et al., 2014). Rivera et al. (2014) compiled positive MS fragment information of specific carotenoids acquired using various ionization methods and composites in a recent study. This study's fragmentation pattern and intensity fragment ratios will be very valuable in evaluating MS data for carotenoid identification. Qiu et al. (2014) recently used C30-HPLC-APCI-MS in conjunction with UV/Vis spectroscopy, gel permeation chromatography (GPC), and NMR spectrometry to identify nine different isomers of canthaxanthin. Recently, accelerated solvent extraction (ASE) was utilized to extract carotenoids from orange carrots, and it was shown that the most crucial elements impacting ASE extraction are solvent properties and temperature. Cardenas-Toro et al. (2015) tried to compare the output, carotenoid characteristics, and financial sustainability of pressurized liquid extraction (PLE), percolation (LPSE-PE), and Soxhlet extraction (LPSE-SOX) of crushed palm fiber for the full recovery of carotenoid-rich concentrations to assess the techniques and industrial applicability (PPF). Compared to LPSE processes, the PLE approach is more efficient to extract carotenoids. When compared to PE, however, LPSE has the lowest production cost. A carotenoid separation process for lutein, α -carotene, and β -carotene from a fresh carrot extract was devised using an Englert et al. technique, and double co-current chromatography (CCC). Using dual CCC, 51 mg of β -carotene, 32 mg of α -carotene, and 4 mg of lutein were separated from 100.2 mg of pure carrot concentrate in a short amount of time and with high purity of 95–99 per cent (Englert et al., 2015).

Saponification may result in the degradation or structural modification due to the origin of the carotenoid and the kind of food (Divya et al., 2012). Rasmussen et al. investigated if saponification of standard lutein may produce meso-zeaxanthin, a xantho-

phyll present in the macula, in addition to zeaxanthin. Peaks with zeaxanthin-like spectra, chiral normal-phase and meso-zeaxanthin retention lengths were obtained from pure lutein saponification (Rasmussen et al., 2012). Irakli et al. discovered significant β -carotene and lutein recoveries following grain saponification, but lesser recoveries (ranging from 46.7 per cent for β -carotene to 74.5 per cent for lutein) after extraction without saponification. In recent research on table olives, Amorim-Carrilho et al. (2014) reported limited β -carotene recovery after saponification. A study investigated the concentration of β -carotene in coriander before and after saponification and discovered that saponification destroyed 20–30% of β -carotene and 50% of another carotenoid (Divya et al., 2012). The number of studies that focused on analytical methods for detecting carotenoid compounds was summarized in Table 1.

More recently, non-invasive analytical techniques started to be applied in determination of carotenoids. These techniques are commonly based on infrared spectroscopy that measures the vibrational spectrum of the sample of interest, without the need of applying any extraction methods. The infrared radiation passes through the samples and records the wavelengths that have been absorbed and the degree of absorption. A study by (Toledo-Martín et al., 2018) applied near infrared spectroscopy to determine the carotenoid content in blackberries. This method was suitable for screening purposes, but not quantification. On the other hand, (Ikeogu et al., 2017), was successful in developing a quantification model using a portable visible and near infrared spectrometer to quantify the carotenoid content of fresh cassava roots. This green technique has also shown promising prediction of provitamin A carotenoids content in maize (Rosales et al., 2022) as well as carotenoids in tomato products (Saad et al., 2021). Infrared spectroscopic methods could be further developed to support the food industry in determining the carotenoid content of the produced products. These methods do not require sample preparation, extraction processes, and use of solvents. In fact, they are relatively quick and cheap in comparison to chromatographic techniques that makes them good candidates as analytical techniques in the industry (Zare et al., 2019).

5. Summary of the role of carotenoids in human health

Carotenoids are intriguing natural substances that can be transformed into a variety of other molecules. Retinoids with vitamin A activity stand out among them, as they are critical for human health and development. Due to the fact carotenoids are colored, one of the most effective aesthetic attributes considered by people to assess food attractiveness. This advantage can be used to develop functional foods contributing to the human health. It is important to highlight that the presence of conjugated double bonds in the structure of carotenoids is responsible for the chromophore of carotenoids (Khoo et al., 2011). These bonds absorb light, and as the number of these double bonds increases, the absorbance of the red color wavelength increases too.

Carotenoids have a lot of evidence that they can assist enhance health and lower the risk of many illnesses. Because carotenoids include the provitamin A carotenoids (PAC) and serve as Vitamin A precursors, they stand out among the wide array of bioactive phytochemicals. Their function as pigments is also important for food marketability, aesthetics, and customer preferences. Carotenoids (and/or their derivatives) are good for one's health, according to an increasing body of data. Carotenoids, too, have been demonstrated in tests to offer cosmetic benefits in addition to their ability to protect the skin (Meléndez-Martínez et al., 2019).

Vitamin A is recognized as the vitamin having the most diverse

Table 1. Analytical techniques applied for the determination of carotenoids

| Sample | Analyte | Analytical technique | References |
|--|--|---|------------------------|
| Tamarillo (<i>Solanum betaceum Cav.</i>) | Zeaxanthin; Lutein; Apocarotenal; Physalins; β -Carotene; β -Cryptoxanthin | LE and LC-PDA-MS analysis, C30 column (250 \times 4.6 mm, 5 μ L), mobile phase: (A) water/20 mm ammonium acetate, (B) MeOH/20 mm ammonium acetate and (C) MTBE gradient elution | Giuffrida et al., 2018 |
| Tomatoes | Lycopene; Lutein; β -Carotene; α -Tocopherol | LC-UV/VIS: RP C18 column (125 \times 4.6 mm, 5 μ m), mobile phase (MeOH:ACN:DCM, 50:48:2 v/v/v) | Zanfini et al., 2010 |
| Carrot | β -Carotene; Carotenes; Xanthophylls | LC-PDA: RP C18 column (250 mm), mobile phase (A): MeOH and (C) 10% n-hexane with ACN, (B) 1% water in MeOH | Agócs et al., 2007 |
| Different citrus species (lime, mandarin, orange, clementine, grapefruit, lemon and kumquat) | β -Carotene; α -Carotene; β -Citraurin; Lutein Violaxanthin; (9z)-violaxanthin; Mutatoxanthin; β -cryptoxanthin; zeaxanthin | LC-PDA: C18 column (250 \times 4.6 mm, 5 μ m), mobile phase (A): 30 percent (v/v) DCM in MeOH, (B): MeOH, (C): MeOH containing 12% (v/v) H ₂ O | Kidmose et al., 2007 |
| Sweet potato | β -Carotene and isomers | LC-PDA: C30 column (250 \times 4.6 mm, 5 μ m), isocratic elution, mobile phase: MeOH/MTBE (80:20 v/v) | Sancho et al., 2011 |
| Coriander (<i>Coriandrum sativum</i> L) | β -Carotene | LC-PDA: RP C18 column (259 \times 4.6 mm), mobile phase: ACN:MeOH:Ethylacetate (80:10:10 v/v/v) | Divya et al., 2012 |
| Papaya | β -Carotene; β -Cryptoxanthin; Lycopene | LC-PDA-APCI (+)/MS: C30 column (150 mm \times 4.6 mm, 3 μ m), mobile phase: (A) MTBE (B) MeOH: gradient elution 0 to 100% | Divya et al., 2012 |
| Orange juice | Lutein; Zeaxanthin; β -cryptoxanthin; α -cryptoxanthin β -carotene; α -carotene | LC-PDA: RP C18 column (250 \times 4.6 mm, 5 μ m), mobile phase: (A) ACN/DCM/MeOH 70:5:25 (v/v/v), (B) MeOH/water 75:25 v/v | Ndolo and Beta, 2013 |
| Cereal grains | 13-cis lutein; 9-cis lutein; 90-cis lutein; Zeaxanthin; β -cryptoxanthin; Lutein | LC-PDA: S-3 column (4.6 \times 100 mm, 3 μ m), mobile phase: MeOH/MTBE/Milli Q water (81:15:4, v/v/v) and MeOH/MTBE (90:19, v/v) | Ndolo and Beta, 2013 |
| Saffron | Trans and cis-isomers of crocin | LC-PDA-ESI (+)/MS: SB-C18 column (4.6 \times 150 mm, 3.5 μ m) linked to an SB-C18 guard column, mobile phase: (A) 0.15% formic acid in MeOH, and (B) 0.15% formic acid in water | Lech et al., 2009 |
| Frozen carrot and spinach | Lutein Phytofluene Phytoene; β -Carotene; α -Carotene | LC-PDA: Column ODS3 (250 \times 4.6 mm 5 μ m), mobile phase: (A) ACN/n-hexane/MeOH and DCM (2:1:1:1, v/v/v/v), and (B) ACN | Mazzeo et al., 2011 |

ACN: Acetonitrile; APO: Trans-b-apo-80-carotenal; BHT: Butylhydroxytoluene; DCM: Dichloromethane; MeOH: Methanol; MTBE: Methyl tert-butyl ether; THF: Tetrahydrofuran; SFC: Supercritical fluid chromatography; RP: Reversed phase; HPTLC: High-performance thin-layer chromatography; UPLC: Ultra-high-performance liquid chromatography.

range of activities in humans (Alvarez et al., 2014). It is required for cell differentiation, reproduction, immunity, eyesight, epithelial tissue maintenance, proliferation, apoptosis, brain function, and/or embryonic growth and development, among other things. Vitamin A deficiency (VAD) (serum retinol values 0.7 mol/L) is a worldwide public health concern, especially in developing nations. VAD is divided into different categories, with 10–20 g/dL (0.35–0.7 mol/L) circulation levels considered low and 10 g/dL (0.35 mol/L) circulation levels considered inadequate. VAD causes a wide range of clinical symptoms, including xerophthalmia (night blindness that develops into softening ulceration and corneal necrosis) and adverse effects on growth, foetal development, and susceptibility to severe infection. VAD is believed to account for a significant fraction of child mortality rates (about 1–2.5 million per year) in poor nations (Alvarez et al., 2014).

Numerous studies conducted over the last 40 years have discovered that carotenoids and their derivatives may be beneficial to health and reduce the risk of a variety of ailments. In the traditional carotenoid publications as well as more recent modifications of the

original studies, carotenoids have been linked to possible improvements in immune function and illnesses such as ovarian, breast, cervical, prostate, and colorectum, cardiovascular problems, skeletal, epidermis, and eye concerns (Eggersdorfer and Wyss, 2018; Hayhoe et al., 2017; Van Hoang et al., 2018).

Long-term supplementation with 50 mg of β -carotene every other day for 18 years has also been shown to provide psychological beneficial results to an otherwise healthy population (n = 4,052 males, Physicians' Health Study II) (Grodstein et al., 2007). The role of carotenoids and their variants in diabetes, obesity, and some other metabolic diseases is also becoming more widely recognized (Mullan et al., 2017; Leermakers et al., 2016). Recently, there has been a surge of motivation in studying more about the positive advantages of carotenoids through pregnancy and childhood (Leermakers et al., 2016). Furthermore, studies have been performed to assess the relationship between carotenoids and the risk of developing various disorders, such as respiratory disorders and Parkinson's disease (van Lent et al., 2016). The development of cardiovascular disease is linked to oxidative stress, inflamma-

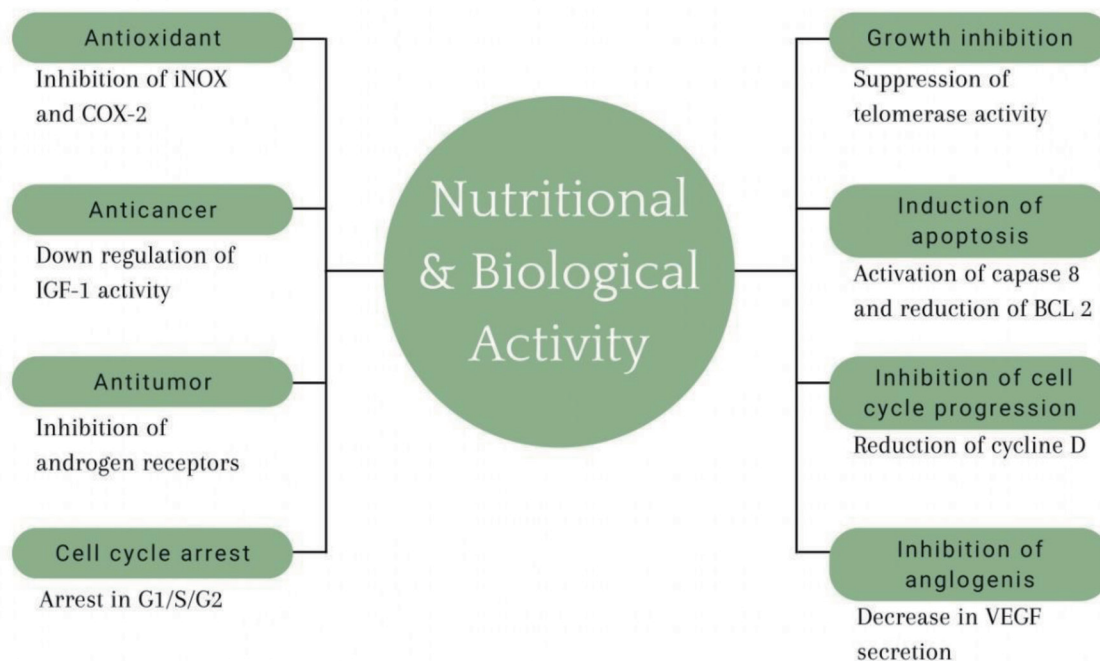


Figure 4. Nutritional and biological activity of carotenoids.

tion, dyslipidemia, and thrombosis, and there are indications that carotenoids may assist with some of these aspects. Astaxanthin, a carotenoid produced by marine creatures, has been proven to lower low-density lipoprotein peroxidation as well as enhance blood lipids and blood flow capacity. Low-density lipoprotein (LDL) oxidation susceptibility is recognized to lead to atherosclerotic. 3.6 mg and above lowered the vulnerability of LDL to oxidation, as indicated by a 5.0, 26.2, 42.3, and 30.7 per cent, respectively, prolongation of the LDL oxidation lag phase in a 14-day investigation into alleged 24 healthy adults who took astaxanthin at doses of 1.8, 3.6, 14.4, and 21.6 mg/day (Eggersdorfer and Wyss, 2018).

Lycopene is a non-provitamin A carotenoid, with remarkable antioxidant activity. It has grabbed the attention of many researchers recently exploring its effectiveness in preventing prostate cancer. Based on the *in vitro* and *in vivo* evidence, lycopene has anticancer, anti-progressive and apoptotic activities (Mirahmadi et al., 2020). This compound has been effective in suppressing progression, proliferation, arresting the in-cell cycle, and inducing apoptosis of prostate cancer cells. Nevertheless, systematic reviews and meta-analyses of clinical trials have concluded that the current evidence does not prove the potential of lycopene in preventing prostate cancer. A recent systematic review and meta-analysis evaluating the evidence of the effect of lycopene supplementation on prostate-specific antigen (PSA) reported that no significant differences in PSA levels in individuals supplemented with lycopene or tomato extract containing lycopene (WMD = -0.12 ng/ml; 95% CI: -0.62, 0.38 ng/ml; P = 0.64) compared to individuals who did not receive any supplementation (Sharifi-Zahabi et al., 2022). Similarly, another systematic review and meta-analysis reported that lycopene did not influence PSA levels in men with non-metastatic prostate cancer (WMD = -0.60, 95% CI = -2.01, 0.81 µg/L) (Sadeghian et al., 2021).

Many carotenoids included in a healthy diet concentrate in the skin, where they provide adequate protection against sunburn, ageing, and UV-induced damage. Carotenoids have a significant abil-

ity to scavenge reactive oxygen species such as peroxide radicals or singlet oxygen molecules due to their unique structure of ten or more conjugated double bonds. Several medical investigations have found that β -carotene can help prevent sunburn, also known as erythema (Stahl and Sies, 2012). A summary of key nutritional and biological activities of carotenoids are shown in Figure 4 (Milani et al., 2017).

6. Absorption and metabolization of carotenoids

Carotenoids and other fat-soluble micronutrients track the destiny of lipids in the human upper gastrointestinal tract. Their digestion begins with their disintegration during the fat phase of the food. This phase is emulsified in the stomach and duodenum into lipid droplets. It was hypothesized that carotenoids *cis*-isomerization may occur during gastrointestinal digestion, however a human investigation ultimately disproved this theory. Recent *in vitro* digestion tests simulating duodenal circumstances shown that lycopene, β -carotene, and lutein did not undergo any appreciable isomerization (Reboul, 2019). Absorption efficiency of marked β -carotene is considerably diverse across clinical investigations, changing from \approx 3 to 80%, but generally ranging from 10 to 30%. This may partially be related to the varied bioavailability of β -carotene, but it may also represent its modest absorption and transport via the enterocyte. It should be highlighted that β -carotene absorption capacity was generally tested after a single meal. However, the gut may retain β -carotene from a first meal to release it during later postprandial stages in humans. β -carotene absorption capacity may, consequently, be underestimated in certain experiments. Approximately 40 percent of ingested carotenoids are remain undigested. BCO1 (β -carotene oxygenase 1), a cytosolic enzyme, may convert β -carotene into retinal in a single step in the enterocyte. The generated retinal is then transformed into retinol and esterified

into retinyl esters by the lecithin:retinol acyltransferase (LRAT) and presumably by the diacylglycerol acyltransferase 1 (DGAT1), which has an acyl-CoA:retinol acyltransferase activity (Amengual et al., 2013). BCO2 may also cleave both provitamin A and non-provitamin carotenoids asymmetrically into apocarotenoids (Palczewski et al., 2014).

7. Techno-functional characteristics of carotenoids

Our food incorporates a diverse set of components with a variety of technological and functional qualities. These components interact with one another throughout the food processing process, from harvesting to consumption. The complexes formed by their interaction have qualities that differ significantly from the inherent properties of the individual ingredients in the food system. Many properties, such as water retention, gelation, viscosity, and emulsification, are altered by interactions between macromolecules, carbohydrates, proteins, and lipids, as well as micro-molecules. Many features of meals and food supplies, such as microstructure, particle size distribution (PSD), and mouthfeel, are impacted by many components and their interactions during the food manufacturing process. The connection of compounds used in nutrient content, as well as those found naturally in food, has a massive effect on the emulsification behavior, water retention characteristics, foaming and foam consistency, gelation solubility, water holding capacity (WHC), flavour characteristics, and viscosity of the food system (Bobade et al., 2021).

In vivo and *in vitro* research have revealed that the interplay of phytochemicals, particularly carotenoids and polyphenolic compounds, is linked to a lower risk of certain degenerative illnesses such as cancer and atherosclerosis. Carotenoid absorption in humans is mostly associated with a fruit and vegetable diet. In recent decades, the rise of functional ingredients has sparked substantial interest in alternative dietary factors of carotenoids. Functional foods are defined as full, fortified, enhanced, or upgraded foods that have beneficial health impacts on the human body. Furthermore, functional meals should be in the form of conventional foods rather than tablets and capsules (Conboy Stephenson et al., 2021; Dunham and Johnson, 2013).

A recent study (Márquez-Cardozo et al., 2021) compared fresh pieces to thermally processed pumpkin (*Cucurbita maxima*) completely dried at various temperatures (55, 60, 65, and 70°C). After determining the drying kinetics and powder quality parameters, the modified Page model offered the best match, with energy levels of 29.47 kJ mol⁻¹ and 16.06 kJ mol⁻¹ for drying fresh and thermal pre-treated sliced, respectively. Powders retained their techno-functional qualities; oil absorption capacity was 1.00–1.30 g/g, water content was 3.42–6.52 g/g, and the solubility in cold water varied from 5.36 to 6.46 per cent. The DPPH and ABTS techniques revealed substantial differences in the physicochemical qualities and total carotenoids of powders generated from heat-treated pumpkin pulp, but not in the techno-functional characteristics and antioxidant capacity.

8. Uses of carotenoid-rich ingredients in the development of food products

Foods with therapeutic benefits for human health have been around for over 2,500 years in various cultures. Any chemical included in food products, animals, or crops that have a function on the human body of consumes it is referred to as a bioactive compound. Bioac-

tive substances, in this context, are the ones that, when applied to foods, promote health. Such elements may be included in bioactive compounds found in natural sources, such as nutritious fiber; or in foods enriched with bioactive molecules, such as probiotics and antioxidants; or in foods fortified with bioactive substances, such as prebiotics. Proteins, probiotics, peptides, plant extracts, omega-3 phytonutrients and formalized lipids, natural fiber, mineral resources, vitamins, special antioxidants, carotenoids, and carbohydrates are the principal bioactive ingredients, all of which can be collected from a variety of sources and are provided to consumers in terms of supplements, personal care products, beverages and functional foods (Fernandes et al., 2019; Dacoreggio et al., 2021).

When developing a carotenoid-fortified food product, a range of factors must be addressed, including optimum carotenoid delivery and a tradeable food item with increased consumer value. Carotenoids' limited bioavailability, solubility, and consistency are key hurdles for the food industry when producing a fortified food product. Carotenoid levels in plasma are commonly used to calculate systemic absorption of functional meals. *In vivo* bioavailability of carotenoid-rich foodstuffs influences their bioavailability and consequent bioactivity. Identifying the barriers and factors to limited carotenoid bioavailability is essential for developing a functional meal. A recent study presented a comprehensive review of carotenoid absorption and variables limiting bioavailability, such as dietary ingredients, carotenoid-nutrient interaction, and treatment techniques. Carotenoids uptake and availability can be significantly altered by dietary components. Dietary fats increase carotenoid absorption from low-fat fruits and vegetables. While producing a fortified product for maximum bioactivity *in vivo*, dietary fat absorption, type of lipid, food composition, and carotenoid composition should all be factored (Conboy Stephenson et al., 2021).

8.1. Carotenoids in bakery products

Bakery products, particularly bread, are among the top leading staple foods in most countries and cultures due to their easy preparation, affordability, diverse tastes, and relatively prolonged shelf life (Salehi, 2019). This product is a complex multi-component system consisting of carbohydrates (*e.g.*, starch as the main form), plant proteins, lipids, fiber, B vitamins, and minerals (*e.g.*, magnesium, phosphorus, and potassium). They are usually prepared from flour or meal derived from grain (Silow et al., 2016). Bread consumption patterns are quite different between countries as turkey had the first-highest bread consumption (app. 104 kg), followed Bulgaria (app. 95 kg), and the UK (app. 32 kg). Although the bread has a key role in meeting the daily nutritional needs of people, it is not considered a complete food source alone. Therefore, enriching flour and bread with fruits and vegetables rich in functional compounds, particularly carotenoids, increases the nutritional value of bread and ultimately improves people's diet (Minarovičová et al., 2018).

Various studies have found that incorporating fruit wastes into bakery products, such as muffins, cakes, and cookies, increased their nutritional values and antioxidant activities, as summarized in Table 2. (Olawuyi and Lee (2019) studied muffins prepared by the combination of shiitake mushroom (SM) powder and carrot pomace (CP) powders (5, 10, and 15%) with rice flour. The incorporation of SM and CP increased total polyphenol, total carotenoid content (ranging from 34.68 to 66.96 µg g⁻¹), and antioxidant properties in a dose-dependent manner. Moreover, the results showed a direct link between the MP and CP concentrations and the weight and hardness of enriched muffins. Based on the sensory

Table 2. Incorporation of carotenoids in development of bakery products

| Product | Sources of Carotenoids | Carotenoids | Treatment | Product properties | Ref. |
|------------------|-----------------------------------|---|--|--|-----------------------|
| Rice muffins | Shiitake mushroom & carrot pomace | - | Incorporation of CP & MP at 5, 10, and 15% into 200 g of rice flour | The enriched muffins with 5 and 10% had better consumer acceptability and higher antioxidant capacity | Olawuyi and Lee, 2019 |
| Bread & cake | Passion fruit peel | lutein, zeaxanthin, α -carotene, β -carotene, provitamin A | Enriched bread and cake were prepared with 85:15 (wheat flour:OP) and 50:50 (wheat flour:OP) | The fortified product had about 80% acceptance score with strong antioxidant activity | Reis et al., 2020 |
| Bread | Orange-fleshed sweet potato puree | β -carotene | Enriched bread constituted of 30% puree and 70% wheat flour | The formulated cake had longer shelf life and reduced water activity | Wanjuu et al., 2018 |
| Bread and muffin | Hairless canary seed & corn | lutein, zeaxanthin, β -cryptoxanthin | HCS with corn flour at ratios of 1:1 and 1:2 (w/w) | The higher reduction of total carotenoids under dough process | Abdel-Aal, 2008 |
| Bread | Stinging nettle | lutein & carotene | The incorporation of 100 g nettle leaves with 1 kg of double zero flour, 500 mL water; 40 g extra-virgin olive oil, 30 g yeast, 20 g NaCl, and 10 g enzyme | The formulated bread possessed higher antioxidant activities and contained higher mineral compounds, fibers, and antioxidant compounds | Maietti et al., 2021 |
| Bread | Orange-fleshed sweet potato | all-trans β -carotene | The formulated bread contained 10:90, 20:80, and 30:70 w/w (OFSP: wheat flours), 3% sugar, 1% salt, 1% fat, 2% yeast, and 1% EN85 | The formulated could supply vitamin A deficiency in children | Nzawita et al., 2017 |
| Cake & cookies | Tomato waste | lycopene | The cakes were made from 1–5% lycopene extracts, 500 g wheat flour (72%), 60 ml sunflower oil, 320 ml cow milk 3.5% fat, 2 eggs, 300 g sugar, 2 g vanillin, 16 g baking powder, and 4 g coconut powder and the cookies had 500 g wheat flour (72%), 750 g butter (80% fat), 2 eggs, 150 g powdered sugar, 1 g vanillin and 8 g baking powder in addition to lycopene | No significant changes in the color and sensory attributes of lycopene-enriched cakes and cookies and a significant increasing in their antioxidant activities | Eletr et al., 2017 |
| Bread | Yellow sweet potato | β -carotene | The enriched bread was made of 3–9% yellow sweet potato with 91–97% wheat flour | Increase β -carotene intake, moisture content, water activity and more yellowish color of bread with the increase of carotenoids | Nogueira et al., 2018 |

ratings (e.g., taste), the enriched muffins had better consumer acceptability than the control muffins. Still, it changed color from cream to brown which is attributed to Maillard's reactions. It is a chemical reaction between an amino acid and a reducing sugar that usually takes place in the presence of heat during the baking process. It also reduced the pasting viscosities, which may be due to their lower carbohydrate and the presence of proteins which caused dilution of starch content in the powder blends and influenced the integrity of the starch granules (Nath et al., 2016; Singh et al., 2014). Moreover, the addition of nettle in white flour could remarkably enhance the carotenoid and mineral contents and antioxidant properties (from 372 to 597 g GAE/GFW) of bread (Maietti et al., 2021). As such, Eletr et al. (Eletr et al., 2017) reported an enhanced 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity of lycopene-added cookies and cakes with an increase in the concentration of lycopene up to 5% without considerable negative effects on sensory parameters of both groups. Reis et al. (Reis et al., 2020) studied chemical attributes and consumer acceptance of bread and cake containing different classes of carotenoids from orange passion fruit peel (OP). The increase of drying temperature in OP up to 80 °C resulted in an increase of total carotenoids by 23%. At this temperature, OP flour had good water holding (7.89 ± 0.14 g/g) and oil holding capacities (2.96 ± 0.04 g/g). Also, the lowest proteins, carbohydrates, Na and total fat values and a higher level of fiber were achieved when adding yellow passion fruit peel flour at 12.22%. The fortified flour provided bread and cakes with more than 80% acceptance rate and greater antioxidant capacity (2.64 ± 0.01 g/100 mL).

Although the wide range of synthetic colorants known as tartrazine has been widely used in bakery products, carotenoids considered a safe, natural pigment are finding their place in this industry. The total carotenoids extracted from mandarin epicarp (containing 140.70 ± 2.66 mg β -carotene/100 g) by ultrasound-assisted extraction with close to zero ΔE values is a suitable alternative to tartrazine in the production of cakes and bread (Ordóñez-Santos et al., 2021). The bread enriched with orange-fleshed sweet potato (OFSP) containing a significant amount of β -carotene could improve the shelf-life compared to the wheat bread by controlling the mould growth and lowering water quality and moisture content (Wanjuu et al., 2018). In a study, dough formation and oven baking influence the muffin and bread enriched with hairless canary seed and corn. Due to enzyme degradation and oxidation, higher total carotenoids were found in dough formation in both bakery products (Abdel-Aal, 2008).

The sensitivity of carotenoids to heating during baking is challenging. The effective strategies to overcome this issue were (Xu et al., 2022), adding egg yolks enriched with zeaxanthin and lutein in the muffin formulation and butter. Still, the incorporation of butter influenced the bioaccessibility of carotenoids. Another study (Nogueira et al., 2018) reported a maximum of 25% β -carotene loss during storage in the bread containing yellow sweet potato flour, which might control moisture content by designing a suitable packaging.

8.2. Carotenoids in extruded snack products

Extruded snack products are popular ready-to-eat foods among consumers and redesigning this type of food to enhance their nutritional characteristics, such as micro or macronutrients, phytochemical ingredients, antioxidants, and vitamins has been developed recently in food industries (Pensamiento-Niño et al., 2018). Carotenoids, as a bioactive component, which confer numerous health benefits, can be incorporated into snacks to improve not

only their nutritional value but also their sensory characteristics (McClements, 2019). Carotenoids are abundant in agricultural wastes like tomato by-products (Wang et al., 2022); therefore, those components can be exploited to fortify snacks. In this regard, recently, tomato pomace powder at varying concentrations (5, 10, 15, and 20% (w/w)) was used to formulate extruded snacks. It was reported that when the tomato pomace powder was increased in products, the bioactive compounds and their bioaccessibility were increased (Yagci et al., 2022). Beside the carotenoid-rich additives like tomato pomace powder, carotenoid-rich flours can use for enriching the extruded snacks. A study (Kolniak-Ostek et al., 2017) used amaranth, Jerusalem artichoke, and pumpkin in the formulation of extruded snacks and reported that the highest β -carotene and lutein belonged to the products made of pumpkin flour. In another study, (Ruiz-Armenta et al., 2018) evaluated the optimization of snack production added with bagasse of naranjilla (*Citrus mitis* B.) fruit as the optimizations of a natural source of carotenoids. The by-products (bagasse) of the naranjita juice industry are a good source of bioactive compounds (including flavonoids, carotenoids, and dietary fibers). In this study, the influence of moisture content (MC) (21.10–32.89%), extrusion temperature (ET) (89.8–140.2 °C), and dehydrated naranjita bagasse content (DNB) (1.12–11.88%) were evaluated. Their results showed that the significant impact on the response variables expansion index and penetration force was extrusion temperature. In contrast, the greatest effect on total carotenoids and sensory acceptability was represented by DNB and the optimum extrusion process conditions evaluated with the numerical method were MC = 23%, ET = 125 °C, and DNB = 8.03% (Ruiz-Armenta et al., 2018). The influence of physical characterizations (moisture of the feed (13–17%, wb) and temperature of the barrel (120–160 °C)) of maize grits on the carotenoid profile of expanded snacks was evaluated by Romero Rodríguez et al. (2021) and they reported that carotenoids were affected differently. The positive values for total carotenoids (except at 17%–120 °C), zeaxanthin (just at 15%–120/140 °C), β -cryptoxanthin, and β -carotene represent a higher amount of these components after extrusion, which indicates that the process is favorable for these carotenoids' extractability. In addition, they suggested a worthy process, from nutritional and technological aspects, to extrude at the processing conditions ranging between 120–132 °C barrel temperature and 13.2–13.7% feed moisture.

Extrusion process conditions, such as temperature, feed moisture content, shear rate, and screw speed, can impact the carotenoid content of the yielded snacks; consequently, these parameters should be taken into account. Thermomechanical stress exerted on extruded snacks enriched by carotenoids during the process could degrade carotenoids (Pinho et al., 2021). Different carotenoids show varying degrees of sensitivity to the process. For instance, (Cueto et al., 2017) elucidated that 60% of lutein content declined, whereas zeaxanthin decrement was 40% in extruded snacks fortified by carotenoids. As these bioactive compounds are vulnerable to the high shear conditions and temperatures of the extrusion process, protecting them from degradation using strategies such as encapsulation can be an ideal solution. In a study, encapsulation of proanthocyanidin-rich cinnamon extract via a complex conservation technique increased the carotenoid content in extruded snacks (Favaro-Trindade et al., 2020). Igual et al. (2021) evaluated the impact of rosehip (*Rosa canina*) co-product powder in the extrusion process and recorded that by increasing this powder, swelling power, water absorption, and hygroscopicity were increased. Furthermore, the expansion value, water solubility, porosity, and image factors (perimeter and area) of the extrudates decreased. The main carotenoids of the extrudates were lycopene, lutein, carotene, and zeaxanthin. While catechin, procyanidin dimmer 1, procyanidin

Table 3. Incorporation of carotenoids in development of gluten-free products

| Product | Carotenoid source | Treatment | Product properties | Ref. |
|-------------|---|---|---|-----------------------|
| Bread | Microalga: <i>Chlorella sorokiniana</i> | The gluten-free bread was enriched with 2.5 g (M2.5) or 5.0 g (M5.0) of microalga powder per 100 g of the blend of rice flour and corn starch in substitution of pea flour. | By increasing the microalga powder, protein content and carotenoids content were increased | Diprat et al., 2020 |
| Bread | Bee pollen | The effect of bee pollen at 1, 2, 3, 4, and 5% levels were evaluated the aroma, nutritional, and bioactive ingredients of technologically viable gluten-free bread | The proteins, minerals, soluble and bioaccessible polyphenols, antiradical activity, and total carotenoids were increased at almost all levels (2–5%) | Conte et al., 2020 |
| Bread | Microalga: <i>Tetraselmis chuii</i> , <i>Chlorella vulgaris</i> , and <i>Nannochloropsis gaditana</i> | 4% (w/w, total flour basis) microalgae were added to formulations based on Buckwheat flour Provida (46%) and Rice flour Espiga (31%) | Significant increase in lipids, proteins, antioxidant activity, minerals (Ca, K, Mg, P, Fe, S, Zn, Cu, Mn), and total phenol components was observed to increase different species of microalgae | Qazi et al., 2022 |
| Crackers | Cactus (<i>Opuntia monacantha</i>) | The effect of adding cactus mucilage (2%) and cladode flour (5, 10, 15%) on gluten-free cracker based on sour cassava starch (60%), wholegrain buckwheat flour (10%), and rice flour (30%) | In comparison with control sample, physical properties were not impacted (except for the darker color), higher total phenolic and antioxidant activity, and carotenoids were observed | Dick et al., 2020 |
| Fresh pasta | <i>Arthrospira platensis</i> (spirulina) | The effect of <i>Arthrospira platensis</i> (dried in different temperatures) replacement (1 to 15% w/w) with rice flour on physicochemical, sensory, and digestibility of gluten-free pasta based on rice flour and Psyllium gel in a 50/50 ratio was evaluated | Adding the <i>A. platensis</i> biomass leads to higher carotenoids, phenolic compounds, chlorophylls, and antioxidant activity in the gluten-free pastas, in comparison with control (without <i>A. platensis</i>) and wheat pasta | Fradinho et al., 2020 |

din dimmer 2, di-gallic acid, and isorhamnetin-glucuronide were the major flavonoids components. Strong Pearson correlations were observed between carotenoids, vitamin C, total flavonoids, antioxidant activity, and total folate (Iguar et al., 2021).

8.3. Gluten-free products

Celiac/coeliac disease is a lack of dietary gluten tolerance that is present in wheat, barley, and rye during the lifespan (Lindfors et al., 2019; Lebwohl and Rubio-Tapia, 2021). Therefore, the gluten-free foods' production for individuals who have celiac disease is necessary. The consumption of gluten-free foods has increased so far as it was valued at \$4.3 billion in 2019 and is predicted to have reached \$7.5 billion in 2027 (Kajzer and Diowksz, 2021). On the other hand, since gluten, as a functional protein play the main role in the structure and texture of foods, it is challenging to substitute gluten with other ingredients to compensate for the lack of this protein. Starches and flour of different botanical sources, such as corn, rice, millet, sorghum, pseudo-cereals (quinoa, buckwheat, and amaranth), teff, potatoes, tapioca, legumes, oilseeds, nuts, fruits, and vegetables can be incorporated into foods to modify the structure and texture of foods in absence of gluten (Šmídová and Rysová, 2022; Witezak et al., 2016). Generally, a mixture of these flour is applied to enhance the quality of gluten-free products. Recently, innovative ingredients, including cladode flour (CF) and cactus mucilage (CM) were utilized for gluten-free snack making. It was reported that snacks yielded by CF did not only display great technological properties but also had an enhanced carotenoid content (Dick et al., 2020).

In another study, microalgae were exploited as a functional composition to raise the nutritional characteristics of gluten-free

bread. The addition of microalga improved the protein content, omega-3 content, and lutein content in the fatty acids (Diprat et al., 2020). Further, it was proved that adding 5% of freeze-dried purple and red potatoes to gluten-free bread could increase the amount of phenolic and carotenoid contents of the product (Gumul et al., 2017). The number of studies that focused on gluten-free products was summarized in Table 3.

8.4. Carotenoids in meat products

Carotenoids are considered tetraterpene pigments with orange, red, yellow, and purple colors, which are formed in photosynthetic bacteria, plants, algae, some types of archaea and fungi, and animals (Mori, 2020). Hence, natural resources with carotenoids can be used in meat product ingredients to improve their color during storage or process. The by-products of tomato manufacturing industries, such as peels and seeds, contain a high number of active components including carotenoids (e.g., lycopene, phytoene, β -carotene, lutein, and phytofluene) (Alahakoon et al., 2015). In a study, tomato pomace extract, as a natural source of β -carotene, was spread on the lamb meat surface to improve the shelf-life due to its antioxidant properties. The redness in lamb meat covered with tomato pomace extract was, therefore, higher in comparison to the control sample after 7 days of storage at 2 °C (Andres et al., 2017). Also, the tomato by-products are used in other meat products, such as patties and burgers, dry-fermented sausage, frankfurter and cooked sausages, minced meat, Luncheon roll, and Mortadella to decrease lipid oxidation and improve sensory properties (Domínguez et al., 2020). In addition, adding tomato extracts into meat product formulations has shown a reduction increases the nitrite contents involved in the curing reactions when added together

with nitrite and thus reducing the residual nitrite percentage (Alahakoon et al., 2015).

Moreover, the attendance of carotenoids in meat products can inhibit color alteration induced by some conservation methods like irradiation and modified atmosphere (Calderón-Oliver and López-Hernández, 2022). A study recorded that using fiber microparticles (2.0% w/w level) evaluated from pulp and peel of Japanese plum by-product, which contained coextracted β -carotene, α - and γ -tocopherols, and lutein as well as polyphenols (quercetin derivatives, cyanidins, and pentameric proanthocyanidins) reduced in 50% the formation of thiobarbituric acid reactive substances in raw patties during storage at 4.0 °C for 10 days (Basanta et al., 2018). Similar results were reported by Another study investigated the influence of berry leaves and their inhibitory activity on lipid peroxidation of broiler chickens' thigh meat. They also recorded that by increasing the doses of vitamin E, zeaxanthin, lutein, and β -carotene, as well as polyphenols, total antioxidant capacity and inhibiting meat lipid peroxidation was increased (Varzaru et al., 2020).

However, if some specific carotenoids, will be utilized in high dosage, they lost their beneficial impact and instead act as pro-oxidants (Bolognesi and Garcia, 2018). Accordingly, finding a proper portion of carotenoids in food formulations is crucial. Recently, papaya epicarp flour, which includes various carotenoids such as β -cryptoxanthin, zeaxanthin, β -carotene, α -carotene, and lycopene, was incorporated into the beef burger to produce a nitrite-reduced burger. The optimal color-maximization in the beef burger was obtained when the nitrite/flour ratio was 150 mg/kg nitrite/40 mg/kg flour, and the maximum values for lycopene, β -cryptoxanthin, and zeaxanthin in the sample were 0.251, 0.451, and 0.447 mg/100 g, respectively (Velasco-Arango et al., 2021). It is worth noting that carotenoids that are intrinsically present in some meats can play a protective role. In this regard, the protein-bound carotenoid present in salmon may preserve unsaturated lipids from tyrosine (Radünz et al., 2021). In addition to fortifying foods with natural sources of carotenoids, there is another strategy to increase the carotenoids in meat. In this technique, animals are fed by plants enriched with carotenoids, and consequently, desired characteristics appear in their meat. For instance, it is claimed that a high carotenoid diet of chicken possessed better bioavailability of provitamin A carotenoids and higher yellowness scores in their meat (Nabi et al., 2020). In Table 4, studies investigating the effect of adding citrus by-products as carotenoids source to meat products are summarized.

8.5. Incorporation of carotenoids in other products

In addition to the abovementioned food categories, carotenoid sources were used to products promote the functional properties of other food products, like pasta, dessert, and sauces. For example, ripe and unripe gac fruit (*Momordica cochinchinensis* Spreng) enrichment (5–15% w/w) of wheat flour pasta increased antioxidant activities (%) and undigested starch (%) in the past. The replacement of gac powder (GP) in the flour could increase the cooking loss but reduce swelling and water absorption. The enriched flour with the 10% powder significantly enhanced the pasta's hardness, adhesiveness, and cohesiveness. The modified pasta with unripe GP at 5% GP had the highest scores in all tested sensory attributes (Chusak et al., 2020). A comparative study Porto Dalla Costa et al. (2016) confirmed higher total carotenoids and antioxidant capacity of pasta enriched by 20% waste than that produced with 30% synthetic β -carotene. No carotenoid loss was observed after cooking at a moderate temperature. In another study, stinging nettle (3.5%

w/w) enrichment of egg pasta (20% w/w) and semolina improved the lutein and β -carotene contents by 44 and 60% compared to non-enriched pasta. The cooking process caused about 20% carotenoid loss and the maximum bioaccessibility of both carotenoids lasted for two h of colonic fermentation (Marchetti et al. 2018). The addition of 15% tomato peel in wheat enhanced the lycopene and β -carotene to 12.94 and 7.93 mg/100 g, respectively. But the quality of enriched pasta was not acceptable when it comes to adhesive, hardness, cooking loss, and sensory paymasters. Using hydrocolloids in this formula was not compensated for these adverse effects (Padalino et al., 2017). The impact of *S. cumini* pulp (from 0 to 40%) on pasta's sensorial, phytochemical, nutritional, and functional attributes were assessed. This formulation doubled and increased the antioxidant activities and β -carotene up to more than five times, respectively, compared to the non-enriched samples. The most acceptable formula belonged to the pasta with 30% pulp due to the highest nutritional compounds and the most acceptable color, taste, and texture (Panghal et al., 2018). A study by Waqas et al. 2017 used 1–5% carotenoids extracted from tomato peel containing 690 mg/100g carotenoids for wheat flour spaghetti and found 55% enhanced antioxidant capacity and 102 higher score accessibility in compression to no tomato-added spaghetti. In another investigation (Agustini et al., 2017), 9% *S. platensis* paste added to the noodle improved β -carotene (from 0.06 ± 0.007 to 1.51 ± 0.305 mg/100 gr), protein (from $8.88 \pm 0.39\%$ to $28.603 \pm 0.33\%$), and moisture contents with acceptable sensory. A study by Nour (2021) demonstrated the inhibition of peroxide values and lipid peroxidation by 64% and 45%, respectively, in a functional property developed by fortifying soymilk mayonnaise with sea buckthorn juice containing 14.66 mg/100 g total carotenoids. Adding sources of carotenoids to snacks can boost the nutritional values of these products for children. In a study another study (Hashim et al., 2021), *Manilkara zapota* was used in the formulation of pastel at different concentrations from 10–20%, and the results were demonstrated. In Table 5, studies investigating the effect of adding carotenoids to a variety of products are summarized.

9. Conclusions

Carotenoids as tetraterpene pigments can be derived from various plant, alga, and bacteria species. They offer a wide variety of functional effects, namely anti-inflammatory, anti-cancer, and antioxidant activities and protect our body from degenerative disorders. The most common methods applied for carotenoids' characterization and quantification in food matrix are thin-layer chromatography, colorimetric, HPLC, and spectrophotometric, among which HPLC is predominant. The main obstacle to carotenoid uses in different types of food products includes poor water solubility, poor stability under different pH ranges, sensitivity to oxidation, high extraction cost, and low bioaccessibility. Encapsulation and nano delivery systems are effective strategies for such issues, but more research is still needed to confirm their efficiency and safety. The incorporation of carotenoids into diverse food products, namely bakery, snacks, gluten-free, meat, and dairy products, can not only improve their nutritional values like antioxidant activities but also improve their physicochemical behaviors such as color, flavor, texture, and crispness.

Funding

This research received no funding.

Table 4. Incorporation of citrus by-products in meat and/or meat products

| Product | By-product | Experiment | Effect on Meat and/or meat product | Ref. |
|--|---|---|---|----------------------|
| Beef muscle | Orange Peel Extract | Effect of 2, 4, and 4% of orange peel extract on fresh and preserved beef for 0, 15, 30, and 60 days of the storage period evaluated | Color, flavour, tenderness, juiciness, and overall acceptability at varied concentrations of orange peel extract were almost like control but decreased remarkably with increased storage time | Haque et al., 2020 |
| Alternatively cured all-pork Bologna sausage | Citrus fiber | Sodium tripolyphosphate replacement with citrus fiber (0.50, 0.75, and 1.00%) | Citrus fiber treatments resulted in bologna sausage with excellent technological properties, as indicated by similar emulsion stability and cook/chill yields | Powell et al., 2019 |
| Low-fat beef hamburgers | Lemon fiber | Evaluation of replacing fat with lemon fiber (2, 4, and 6% levels) on physico-chemical, organoleptic, and textural properties of low-fat beef hamburgers | Lemon fiber increased cooking yield and moisture content due to its better water-binding characteristics, redder, lighter, and more yellow color; hardness, springiness, gumminess, and chewiness values decreased when the concentration of lemon fibers increased | Soncu et al., 2015 |
| Beef patties | Citrus orange or lemon peels powder | Quality properties and shelf-life evaluation of the beef patties during refrigeration storage at 4 ± 1 °C for 15 days by adding citrus peels | Thiobarbituric acid reactive (TBA) substances, microbial count, pH value, total volatile basic nitrogen, and shrinkage decreased; increase in cooking yield, water and oil holding capacity, and moisture and fat retentions by adding 1 or 2% of both powders. The color, texture and sensory properties, and radical scavenging activity affected in resulted of adding powders | Ibrahim et al., 2018 |
| Beef burger | Orange Peel Powder | Evaluated the lipid stability of beef burger with varied percentages (2.5, 5.0, 7.5, and 10.0%) of orange peel | The lowest TBA value was determined in the burger with 10% | |
| Turkey sausage | Citrus paradisi fruit barks (CPF) extract | Assessment of different solvents, including ethyl acetate, methanol and water on antimicrobial and antioxidant activities of CPF extract and turkey sausage shelf life. | CPF retard lipid oxidation in sausages was evaluated at the level of 2.5 g kg^{-1} turkey sausage, by the CPF water extract addition, the number of microorganisms decreased significantly | Sayari et al., 2015 |

Table 5. The effects of carotenoid-rich sources to a variety of other products

| Products | Sources of carotenoids | Types of carotenoids | Treatment | Outcomes | Ref. |
|---------------------|--------------------------------|---|---|---|--------------------------------|
| Pasta | Gac fruit | lutein, β -carotene, zeaxanthin, β -cryptoxanthin, α -carotene, lycopene, and α -carotene | The incorporation of 5–15 g gac fruits powder, 85–95 g wheat flour, 1 g salt, 50 g whole egg, and 5 g vegetable oil | Significant increase dietary in fiber, hardness, cohesiveness and decrease in the total carotenoids during cooking and water absorption and swelling index | Chusak et al., 2020 |
| Dry pasta | Carrot flour | β -carotene, lutein, zeaxanthin, cryptoxanthin, α -carotene | From 10% to 20% carrot flour, 100% wheat flour, 37.5% egg powder and 1.13 and 1.35% commercial β -carotene, 25–50% water | Improvement of sensory and nutritional values, carotenoid levels, antioxidant capacity, total fibers, and color | Porto Dalla Costa et al., 2016 |
| Pasta | <i>Urtica dioica</i> L. | β -carotene & lutein | The mixture of 3.5% (w/w) dried nettle leaves, semolina and 20% (w/w) egg, 30% (w/w) water | The highest carotenoid content belonged to flowering stage of nettle which increased the formulated egg pasta and the highest bioaccessibility of nettle-based product at the early stage of colon fermentation | Marchetti et al., 2018 |
| Pasta | Tomato | lycopene & β -carotene | 10 and 15% (w/w) tomato peel flour, 2% hydrocolloids, water, and 85 and 90% (w/w) whole meal flour | The addition of tomato flour enhanced carotenoid and fiber contents but caused slight increase in cooking loss leading to make the pasta more adhesive and firmer with unpleasant odor | Padalino et al., 2017 |
| Pasta | <i>Syzygium cumini</i> L. pulp | β -carotene | The effects of from 0- 40% fruit pulp's addition into pasta formulation on its antioxidant activities, functional compounds, and sensory attributes | It improved antioxidant capacity along with the improvement of nutritional components and caused an attractive appearance, but prolonged the cooking time | Panghal et al., 2018 |
| Spaghetti | Tomato peel | β -carotene, phytoene, phytofluene, cis-lycopene, lutein, cis- α -carotene, α -carotene, γ -carotene | The tomato-based spaghetti contained 72% extract wheat flour, 35% water and 1% salt 1%. | the enriched product had higher carotenoid contents and antioxidant capacity as well as higher consumer's acceptability in terms of taste, color, and texture | Waqas et al., 2017 |
| Dried noodle | <i>S. platensis</i> | β -carotene | <i>S. platensis</i> powder and basil extract at 1:3 ratio, 200 g wheat flour, 30 g egg, 1 g salt, and 1 g baking powder | This incorporation increased protein, B-carotene and moisture contents and improved elasticity, and consumer acceptance of noodle | Agustini et al., 2017 |
| Soy milk mayonnaise | Sea buckthorn juice | - | Soy milk 30.8% (w/w), sunflower oil 61.5% (w/w), salt 1.5% (w/w), mustard 3.1% (w/w) and lemon juice 3.1% (w/w), and 3 and 6% (w/w) sea buckthorn juice | No significant decrease in all sensory attributes except color, inhibition of lipid peroxidation, peroxide value during eight weeks of storage | Nour, 2021 |

Conflict of interest

The authors declare no conflict of interest.

References

- Abdel-Aal, E.-S. (2008). Effects of baking on protein digestibility of organic spelt products determined by two in vitro digestion methods. *LWT - Food Sci. Technol.* 41(7): 1282–1288.
- Agócs, A., Nagy, V., Szabó, Z., Márk, L., Ohmacht, R., and Deli, J. (2007). Comparative study on the carotenoid composition of the peel and the pulp of different citrus species. *Innov. Food Sci. Emerg. Technol.* 8(3): 390–394.
- Agustini, T., Ma'ruf, W., and Wibowo, B. (2017). Study on the effect of different concentration of *Spirulina platensis* paste added into dried noodle to its quality characteristics. IOP Conference Series: Earth and Environmental Science. IOP Publishing, p. 012068.
- Alahakoon, A.U., Jayasena, D.D., Ramachandra, S., and Jo, C. (2015). Alternatives to nitrite in processed meat: up to date. *Trends Food Sci. Technol.* 45(1): 37–49.
- Alvarez, R., Vaz, B., Gronemeyer, H., and de Lera, A.R. (2014). Functions, therapeutic applications, and synthesis of retinoids and carotenoids. *Chem. Rev.* 114(1): 1–125.
- Ambati, R.R., Siew Moi, P., Ravi, S., and Aswathanarayana, R.G. (2014). Astaxanthin: Sources, extraction, stability, biological activities and its commercial applications—A review. *Mar. Drugs* 12(1): 128–152.
- Amengual, J., Widjaja-Adhi, M.A.K., Rodriguez-Santiago, S., Hessel, S., Golczak, M., Palczewski, K., and Von Lintig, J. (2013). Two Carotenoid Oxygenases Contribute to Mammalian Provitamin A Metabolism. *J. Biol. Chem.* 288(47): 34081–34096.
- Amorim-Carrilho, K.T., Cepeda, A., Fente, C., and Regal, P. (2014). Review of methods for analysis of carotenoids. *TrAC - Trends Anal. Chem.* 56: 49–73.
- Andres, A.I., Petron, M.J., Delgado-Adamez, J., Lopez, M., and Timon, M. (2017). Effect of tomato pomace extracts on the shelf-life of modified atmosphere-packaged lamb meat. *J. Food Process. Preserv.* 41(4): e13018.
- Anunciato, T.P., and da Rocha Filho, P.A. (2012). Carotenoids and polyphenols in nutricosmetics, nutraceuticals, and cosmeceuticals. *J. Cosmet. Dermatol.* 11(1): 51–54.
- Bakry, A.M., Abbas, S., Ali, B., Majeed, H., Abouelwafa, M.Y., Mousa, A., and Liang, L. (2016). Microencapsulation of oils: A comprehensive review of benefits, techniques, and applications. *Compr. Rev. Food Sci. Food Saf.* 15(1): 143–182.
- Basanta, M.F., Rizzo, S.A., Szerman, N., Vaudagna, S.R., Descalzo, A.M., Gerschenson, L.N., Pérez, C.D., and Rojas, A.M. (2018). Plum (*Prunus salicina*) peel and pulp microparticles as natural antioxidant additives in breast chicken patties. *Int. Food Res. J.* 106: 1086–1094.
- Bernstein, P.S., Li, B., Vachali, P.P., Gorusupudi, A., Shyam, R., Henriksen, B.S., and Nolan, J.M. (2016). Lutein, zeaxanthin, and meso-zeaxanthin: The basic and clinical science underlying carotenoid-based nutritional interventions against ocular disease. *Prog. Retin. Eye Res.* 50: 34–66.
- Bobade, H., Sharma, S.M., and Singh, B. (2021). Effect of Ingredient Interactions on Techno-Functional Properties. *Food Formulation: Novel Ingredients and Processing Techniques*. Wiley publisher, New Jersey, United States, pp. 71–99.
- Bohn, T. (2018). Metabolic fate of bioaccessible and non-bioaccessible carotenoids. *Non-Extractable Polyphenols and Carotenoids: Importance in Human Nutrition and Health*. Royal Society of Chemistry publisher, London, United Kingdom, pp. 165–200.
- Bolognesi, V.J., and Garcia, C.E. (2018). Annatto carotenoids as additives replacers in meat products. In: Holban, A.M., and Grumezescu, A.M. (Ed.). *Alternative and Replacement Foods*. Academic Press, pp. 355–384.
- Calderón-Oliver, M., and López-Hernández, L.H. (2022). Food vegetable and fruit waste used in meat products. *Food Rev. Int.* 38(4): 628–654.
- Cardenas-Toro, F.P., Alcázar-Alay, S.C., Coutinho, J.P., Godoy, H.T., Forster-Carneiro, T., and Meireles, M.A.A. (2015). Pressurized liquid extraction and low-pressure solvent extraction of carotenoids from pressed palm fiber: experimental and economical evaluation. *Food Bioprod. Process.* 94: 90–100.
- Chusak, C., Chantabunyawat, P., Chumnumduang, P., Chantarasrinlapin, P., Suantawee, T., and Adisakwattana, S. (2020). Effect of gac fruit (*Momordica cochinchinensis*) powder on in vitro starch digestibility, nutritional quality, textural and sensory characteristics of pasta. *LWT* 118: 108856.
- Conboy Stephenson, R., Ross, R.P., and Stanton, C. (2021). Carotenoids in milk and the potential for dairy based functional foods. *Foods*. 10(6): 1263.
- Conte, P., Del Caro, A., Urgoghe, P.P., Petretto, G.L., Montanari, L., Piga, A., and Fadda, C. (2020). Nutritional and aroma improvement of gluten-free bread: is bee pollen effective? *LWT* 118: 108711.
- Coutte, F., Lecouturier, D., Dimitrov, K., Guez, J.S., Delvigne, F., Dhulster, P., and Jacques, P. (2017). Microbial lipopeptide production and purification bioprocesses, current progress and future challenges. *Bio-technol. J.* 12(7): 1600566.
- Cueto, M., Farroni, A., Schoenlechner, R., Schleining, G., and Buera, P. (2017). Carotenoid and color changes in traditionally flaked and extruded products. *Food. Chem.* 229: 640–645.
- Dacoreggio, M.V., Santetti, G.S., Inácio, H.P., Kempka, A.P., and de Mello Castanho Amboni, R.D. (2021). A comprehensive review of eugenia pyriformis cambess: reported bioactivities and health effects. *Food Rev. Int.* 31: 1–15.
- de Freitas Santos, P.D., Rubio, F.T.V., da Silva, M.P., Pinho, L.S., and Favaro-Trindade, C.S. (2021a). Microencapsulation of carotenoid-rich materials: a review. *Int. Food Res. J.* 147: 110571.
- Dick, M., Limberger, C., Thys, R.C.S., de Oliveira Rios, A., and Flôres, S.H. (2020). Mucilage and cladode flour from cactus (*Opuntia monacantha*) as alternative ingredients in gluten-free crackers. *Food. Chem.* 314: 126178.
- Diprat, A.B., Thys, R.C.S., Rodrigues, E., and Rech, R. (2020). Chlorella sorokiniana: a new alternative source of carotenoids and proteins for gluten-free bread. *LWT* 134: 109974.
- Divya, P., Puthusseri, B., and Neelwarne, B. (2012). Carotenoid content, its stability during drying and the antioxidant activity of commercial coriander (*Coriandrum sativum* L.) varieties. *Food. Res. Int.* 45(1): 342–350.
- Domínguez, R., Gullón, P., Pateiro, M., Munekata, P.E., Zhang, W., and Lorenzo, J.M. (2020). Tomato as potential source of natural additives for meat industry: a review. *Antioxidants* 9(1): 73.
- Dunham, A., and Johnson, E.J. (2013). Fruits, vegetables, and their components and mild cognitive impairment and dementia: a review. *Food Rev. Int.* 29(4): 409–440.
- Eggersdorfer, M., and Wyss, A. (2018). Carotenoids in human nutrition and health. *Arch. Biochem. Biophys.* 652: 18–26.
- Eletr, A.A., Siliha, H.A.E., Elshorbagy, G.A., and Galal, G.A. (2017). Evaluation of lycopene extracted from tomato processing waste as a natural antioxidant in some bakery products. *Zagazig J. Agric. Res.* 44(4): 1389–1401.
- Englert, M., Hammann, S., and Vetter, W. (2015). Isolation of β -carotene, α -carotene and lutein from carrots by countercurrent chromatography with the solvent system modifier benzotrifluoride. *J. Chromatogr. A.* 1388: 119–125.
- Favaro-Trindade, C.S., Patel, B., Silva, M.P., Comunian, T.A., Federici, E., Jones, O.G., and Campanella, O.H. (2020). Microencapsulation as a tool to producing an extruded functional food. *LWT* 128: 109433.
- Feeney, J., Finucane, C., Savva, G.M., Cronin, H., Beatty, S., Nolan, J.M., and Kenny, R.A. (2013). Low macular pigment optical density is associated with lower cognitive performance in a large, population-based sample of older adults. *Neurobiol. Aging.* 34(11): 2449–2456.
- Fernandes, S.S., Coelho, M.S., and de las Mercedes Salas-Mellado, M. Bioactive compounds as ingredients of functional foods: polyphenols, carotenoids, peptides from animal and plant sources new. In: Rubi, M., and Campos, S. (Ed.). *Bioactive Compounds*. Woodhead Publishing, pp. 129–142.
- Ferreiro-Vera, C., Mata-Granados, J.M., Gómez, J.Q., and de Castro, M.L. (2011). On-line coupling of automatic solid-phase extraction and HPLC for determination of carotenoids in serum. *Talanta* 85(4):

- 1842–1847.
- Fiedor, J., and Burda, K. (2014). Potential role of carotenoids as antioxidants in human health and disease. *Nutrients* 6(2): 466–488.
- Fradinho, P., Niccolai, A., Soares, R., Rodolfi, L., Biondi, N., Tredici, M.R., Sousa, I., and Raymundo, A. (2020). Effect of *Arthrospira platensis* (spirulina) incorporation on the rheological and bioactive properties of gluten-free fresh pasta. *Algal Res.* 45: 101743.
- Gomez-Estaca, J., Comunian, T.A., Montero, P., Ferro-Furtado, R., and Fávoro-Trindade, C.S. (2016). Encapsulation of an astaxanthin-containing lipid extract from shrimp waste by complex coacervation using a novel gelatin-cashew gum complex. *Food Hydrocoll.* 61: 155–162.
- Giuffrida, D., Zoccali, M., Arigò, A., Cacciola, F., Roa, C.O., Dugo, P., and Mondello, L. (2018). Comparison of different analytical techniques for the analysis of carotenoids in tamarillo (*Solanum betaceum* Cav.). *Arch. Biochem. Biophys.* 646: 161–167.
- Grodstein, F., Kang, J.H., Glynn, R.J., Cook, N.R., and Gaziano, J.M. (2007). A randomized trial of beta carotene supplementation and cognitive function in men: the Physicians' Health Study II. *Arch. Intern. Med.* 167(20): 2184–2190.
- Gumul, D., Ziobro, R., Ivanišová, E., Korus, A., Árvay, J., and Tóth, T. (2017). Gluten-free bread with an addition of freeze-dried red and purple potatoes as a source of phenolic compounds in gluten-free diet. *Int J Food Sci Nutr.* 68(1): 43–51.
- Gupta, P., Sreelakshmi, Y., and Sharma, R. (2015). A rapid and sensitive method for determination of carotenoids in plant tissues by high performance liquid chromatography. *Plant Methods* 11(1): 1–12.
- Gurmu, F., Hussein, S., and Laing, M. (2014). The potential of orange-fleshed sweet potato to prevent vitamin A deficiency in Africa. *Int J Vitam Nutr Res.* 84(1–2): 65–78.
- Haque, F., Rahman, M.H., Habib, M., Alam, M.S., Monir, M.M., and Hosain, M.M. (2020). Effect of different levels of orange peel extract on the quality and shelf life of beef muscle during frozen storage. *IOSR J. Agric. Vet. Sci.* 13: 43–56.
- Hashim, N.H., Zamri, A.I., Rusli, N.D., Smedley, K.L., and Zainol, M.K. (2021). physicochemical properties and sensory characteristics of ciku fruit (*Manilkara zapota*) pastilles. *Food Res.* 5(2): 164–172.
- Hayhoe, R.P., Lentjes, M.A., Mulligan, A.A., Luben, R.N., Khaw, K.T., and Welch, A.A. (2017). Carotenoid dietary intakes and plasma concentrations are associated with heel bone ultrasound attenuation and osteoporotic fracture risk in the European Prospective Investigation into Cancer and Nutrition (EPIC)-Norfolk cohort. *Br. J. Nutr.* 117(10): 1439–1453.
- Helgeland, H., Sodeland, M., Zoric, N., Torgersen, J.S., Grammes, F., von Lintig, J., and Våge, D.I. (2019). Genomic and functional gene studies suggest a key role of beta-carotene oxygenase 1 like (bco11) gene in salmon flesh color. *Sci. Rep.* 9(1): 1–12.
- Honda, M. (2022). Application of e/z-isomerization technology for enhancing processing efficiency, health-promoting effects, and usability of carotenoids: a review and future perspectives. *J. Oleo Sci.* 71(2): 151–165.
- Honda, M. (2021). Carotenoid isomers: a systematic review of the analysis, biological activity, physicochemical property, and methods for isomerization. *Stud. Nat. Prod. Chem.* 68: 173–220.
- Honda, M., Maeda, H., Fukaya, T., and Goto, M. (2018). Effects of z-isomerization on the bioavailability and functionality of carotenoids: a review. *Prog. Carot. Res.* 5: 139–159.
- Ibrahim, H.M., Hassan, I.M., and Hamed, A.A. (2018). Application of lemon and orange peels in meat products: quality and safety. *Int. J. Curr. Microbiol. App. Sci.* 7(4): 2703–2723.
- Igual, M., Chiş, M.S., Păucean, A., Vodnar, D.C., Muste, S., Man, S., Martínez-Monzó, J., and García-Segovia, P. (2021). Valorization of rose hip (*Rosa canina*) puree co-product in enriched corn extrudates. *Foods* 10(11): 2787.
- Ikeogu, U.N., Davrieux, F., Dufour, D., Ceballos, H., Egesi, C.N., and Jannink, J.L. (2017). Rapid analyses of dry matter content and carotenoids in fresh cassava roots using a portable visible and near infrared spectrometer (Vis/NIRS). *PLoS One* 12(12): e0188918.
- Kajzer, M., and Diowks, A. (2021). The clean label concept: Novel approaches in gluten-free breadmaking. *Appl. Sci.* 11(13): 6129.
- Kalac, P. (2012). Carotenoids, ergosterol and tocopherols in fresh and preserved herbage and their transfer to bovine milk fat and adipose tissues: a review. *J. Agrobiol.* 29(1): 1.
- Khoo, H.E., Prasad, K.N., Kong, K.W., Jiang, Y., and Ismail, A. (2011). Carotenoids and their isomers: color pigments in fruits and vegetables. *Molecules* 16(2): 1710–1738.
- Kidmose, U., Christensen, L.P., Agili, S.M., and Thilsted, S.H. (2007). Effect of home preparation practices on the content of provitamin A carotenoids in coloured sweet potato varieties (*Ipomoea batatas* Lam.) from Kenya. *Innov Food Sci Emerg Technol.* 8(3): 399–406.
- Kolnias-Ostek, J., Kita, A., Pęksa, A., Wawrzyniak, A., Hamułka, J., Jeznach, M., Daniłchenko, H., and Jariene, E. (2017). Analysis of the content of bioactive compounds in selected flours and enriched extruded corn products. *J. Food Compos. Anal.* 64: 147–155.
- Kopec, R.E., and Failla, M.L. (2018). Recent advances in the bioaccessibility and bioavailability of carotenoids and effects of other dietary lipophiles. *J. Food Compos. Anal.* 68: 16–30.
- Lebwohl, B., and Rubio-Tapia, A. (2021). Epidemiology, presentation, and diagnosis of celiac disease. *Gastroenterology* 160(1): 63–75.
- Lech, K., Witowska-Jarosz, J., and Jarosz, M. (2009). Saffron yellow: characterization of carotenoids by high performance liquid chromatography with electrospray mass spectrometric detection. *J. Mass Spectrom.* 44(12): 1661–1667.
- Leermakers, E.T., Darweesh, S.K., Baena, C.P., Moreira, E.M., Melo van Lent, D., Tielemans, M.J., Muka, T., Vitezova, A., Chowdhury, R., Bramer, W.M., and Kieft-de Jong, J.C. (2016). The effects of lutein on cardiometabolic health across the life course: a systematic review and meta-analysis. *2. Am. J. Clin. Nutr.* 103(2): 481–494.
- Leong, H.Y., Show, P.L., Lim, M.H., Ooi, C.W., and Ling, T.C. (2018). Natural red pigments from plants and their health benefits: a review. *Food Rev. Int.* 34(5): 463–482.
- Lindfors, K., Ciacci, C., Kurppa, K., Knut, E., Lundin, A., Makharia, K.G., Mearin, M.L., Murray, J.A., Verdu, E.F., and Kaukinen, K. (2019). Coeliac disease. *Nat. Rev. Dis. Primers.* 5: 3.
- López-Igual, P., and Rodríguez-Modroño, P. (2021). Factores de desigualdad entre teletrabajadores en Europa. *Revista de economía crítica.* 31: 62–79.
- Maeda, H. (2013). Anti-obesity and anti-diabetic activities of algae. In: Domínguez, H. (Ed.). *Functional Ingredients from Algae for Foods and Nutraceuticals*. Woodhead Publishing, pp. 453–472.
- Mahmoud, M., Abou-Arab, A., and Abu-Salem, F.M. (2017). Quality characteristics of beef burger as influenced by different levels of orange peel powder. *Am. J. Food Technol.* 12(4): 262–270.
- Maietti, A., Tedeschi, P., Catani, M., Stevanin, C., Pasti, L., Cavazzini, A., and Marchetti, N. (2021). Nutrient composition and antioxidant performances of bread-making products enriched with stinging nettle (*Urtica dioica*) leaves. *Foods* 10(5): 938.
- Marchetti, N., Bonetti, G., Brandolini, V., Cavazzini, A., Maietti, A., Meca, G.m., and Mañes, J. (2018). Stinging nettle (*Urtica dioica* L.) as a functional food additive in egg pasta: Enrichment and bioaccessibility of Lutein and β -carotene. *J. Funct. Foods* 47: 547–553.
- Márquez-Cardozo, C.J., Caballero-Gutiérrez, B.L., Ciro-Velázquez, H.J.m., and Restrepo-Molina, D.A. (2021). Effect of pretreatment and temperature on the drying kinetics and physicochemical and techno-functional characteristics of pumpkin (*Cucurbita maxima*). *Heliyon.* 7(4): e06802.
- Mazzeo, T., N'Dri, D., Chiavaro, E., Visconti, A., Fogliano, V.m., and Pellegrini, N. (2011). Effect of two cooking procedures on phytochemical compounds, total antioxidant capacity and colour of selected frozen vegetables. *Food Chem.* 128(3): 627–633.
- McClements, D.J. (2019). *The science of foods: Designing our edible future. Future Foods: How Modern Science Is Transforming the Way We Eat*. Springer International Publishing, Cham.
- Mendes-Silva, T.D.C.D., da Silva Andrade, R.F., Ootani, M.A., Mendes, P.V.D., da Silva, M.R.F., Souza, K.S., dos Santos Correia, M.T., da Silva, M.V., and de Oliveira, M.B.M. (2020). Biotechnological potential of carotenoids produced by extremophilic microorganisms and application prospects for the cosmetics industry. *J. Adv. Microbiol.* 10(8): 397–410.
- Meléndez-Martínez, A.J., Mapelli-Brahm, P., Hornero-Méndez, D., and Viçario, I.M. (2019). Structures, nomenclature and general chemistry of carotenoids and their esters. Royal Society of Chemistry publisher, London, The UK, pp. 1–50.
- Méresse, S., Fodil, M., Fleury, F., and Chénais, B. (2020). Fucoxanthin, a

- marine-derived carotenoid from brown seaweeds and microalgae: A promising bioactive compound for cancer therapy. *International Journal of Molecular Sciences* 21(23): 9273.
- Milani, A., Basirnejad, M., Shahbazi, S., and Bolhassani, A. (2017). Carotenoids: biochemistry, pharmacology and treatment. *Br. J. Pharmacol.* 174(11): 1290–1324.
- Minarovičová, L., Lauková, M., Karovičová, J., and Kohajdová, Z. (2018). Utilization of pumpkin powder in baked rolls. *Potr. S. J. F. Sci.* 12: 195–201.
- Mirahmadi, M., Azimi-Hashemi, S., Saburi, E., Kamali, H., Pishbin, M., and Hadizadeh, F. (2020). Potential inhibitory effect of lycopene on prostate cancer. *Biomed. Pharmacother.* 129: 110459.
- Mori, T. (2020). Enzymatic studies on aromatic prenyltransferases. *J. Nat. Med.* 74(3): 501–512.
- Mullan, K., Williams, M.A., Cardwell, C.R., McGuinness, B., Passmore, P., Silvestri, G., Woodside, J.V., and McKay, G.J. (2017). Serum concentrations of vitamin E and carotenoids are altered in Alzheimer's disease: A case-control study. *Alzheimer's Dement: Transl. Res. Clin. Interv.* 3(3): 432–439.
- Nabi, F., Arain, M.A., Rajput, N., Alagawany, M., Soomro, J., Umer, M., Soomro, F., Wang, Z., Ye, R., and Liu, J. (2020). Health benefits of carotenoids and potential application in poultry industry: a review. *J. Anim. Physiol. Anim.* 104(6): 1809–1818.
- Nath, P., Kaur, C., Rudra, S.G., and Varghese, E. (2016). Enzyme-assisted extraction of carotenoid-rich extract from red capsicum (*Capsicum annuum*). *Agric. Sci.* 5(2): 193–204.
- Ndolo, V.U., and Beta, T. (2013). Distribution of carotenoids in endosperm, germ, and aleurone fractions of cereal grain kernels. *Food Chem.* 39(1-4): 663–671.
- Nogueira, A.C., Sehn, G.A., Rebellato, A.P., Coutinho, J.P., Godoy, H.T., Chang, Y.K., Steel, C.J., and Clerici, M.T.P. (2018). Yellow sweet potato flour: use in sweet bread processing to increase β -carotene content and improve quality. *An. Acad. Bras. Cienc.* 90: 283–293.
- Nour, V. (2021). Effect of sea buckthorn juice addition on the oxidative stability, physicochemical and sensory properties of soy milk mayonnaise during refrigerated storage. *Ukr. J. Food Sci.* 9(1): 5–17.
- Nzamwita, M., Duodu, K.G., and Minnaar, A. (2017). Stability of β -carotene during baking of orange-fleshed sweet potato-wheat composite bread and estimated contribution to vitamin A requirements. *Food Chem.* 228: 85–90.
- Olawuyi, I.F., and Lee, W.Y. (2019). Quality and antioxidant properties of functional rice muffins enriched with shiitake mushroom and carrot pomace. *Int. J. Food Sci.* 54(7): 2321–2328.
- Ordóñez-Santos, L.E., Esparza-Estrada, J., and Vanegas-Mahecha, P. (2021). Ultrasound-assisted extraction of total carotenoids from mandarin epicarp and application as natural colorant in bakery products. *LWT* 139: 110598.
- Padalino, L., Conte, A., Lecce, L., Likyova, D., Sicari, V., Pellicanò, T.M., Poiana, M., and Del Nobile, M.A. (2017). Functional pasta with tomato by-product as a source of antioxidant compounds and dietary fibre. *Czech J. Food Sci.* 35(1): 48–56.
- Palczewski, G., Amengual, J., Hoppel, C.L., and Von Lintig, J. (2014). Evidence for compartmentalization of mammalian carotenoid metabolism. *The FASEB Journal.* 28(10): 4457–4469.
- Panghal, A., Chhikara, N., and Khatkar, B. (2018). Effect of processing parameters and principal ingredients on quality of sugar snap cookies: a response surface approach. *J. Food Sci. Technol.* 55(8): 3127–3134.
- Pensamiento-Niño, C.A., Gómez-Aldapa, C.A., Hernández-Santos, B., Juárez-Barrientos, J.M., Herman-Lara, E., Martínez-Sánchez, C.E., Torruco-Uco, J.G., and Rodríguez-Miranda, J. (2018). Optimization and characterization of an extruded snack based on taro flour (*Colocasia esculenta* L.) enriched with mango pulp (*Mangifera indica* L.). *J. Food Sci. Technol.* 55(10): 4244–4255.
- Pinho, L.S., Rodrigues, C.E.C., Favaro-Trindade, C.S., and Campanella, O.H. (2021). The incorporation of carotenoids on ready to eat foods studied through their stability during extrusion processing. *Food Eng. Rev.* 13(4): 902–915.
- Porto Dalla Costa, A., Cruz Silveira Thys, R., De Oliveira Rios, A., and Hickmann Flôres, S. (2016). Carrot flour from minimally processed residue as substitute of β -carotene commercial in dry pasta prepared with common wheat (*Triticum aestivum*). *J. Food Qual.* 39(6): 590–598.
- Potrykus, I. (2003). Nutritionally enhanced rice to combat malnutrition disorders of the poor. *Nut Rev.* 61(6): S101–S104.
- Powell, M.J., Sebranek, J.G., Prusa, K.J., and Tarté, R. (2019). Evaluation of citrus fiber as a natural replacer of sodium phosphate in alternatively-cured all-pork Bologna sausage. *Meat Sci.* 157: 107883.
- Qazi, M.W., de Sousa, I.G., Nunes, M.C., and Raymundo, A. (2022). Improving the nutritional, structural, and sensory properties of gluten-free bread with different species of microalgae. *Foods* 11(3): 397.
- Qiu, D., Zhu, W.L., Tang, C.K., Shi, L.F., and Gao, H.Q. (2014). Identification of the composition of isomeric canthaxanthin sample by NMR, HPLC, and mass spectrometry. *Food Anal. Methods.* 7(3): 597–605.
- Radünz, M., Camargo, T.M., Nunes, C.F.P., Pereira, E.D.S., Ribeiro, J.A., Dos Santos Hackbart, H.C., Radünz, A.F.O., Radünz, A.L., Gularte, M.A., and Da Fonseca Barbosa, F. (2021). Gluten-free green banana flour muffins: chemical, physical, antioxidant, digestibility and sensory analysis. *J. Food Sci. Technol.* 58(4): 1295–1301.
- Rasmussen, H.M., Muzhingi, T., Eggert, E.M., and Johnson, E.J. (2012). Lutein, zeaxanthin, meso-zeaxanthin content in egg yolk and their absence in fish and seafood. *J. Food Compos. Anal.* 27(2): 139–144.
- Reboul, E. (2019). Mechanisms of carotenoid intestinal absorption: where do we stand? *Nutrients.* 11(4): 838.
- Reis, L.C.R.D., Facco, E.M.P., Salvador, M., Flôres, S.H., and Rios, A.D.O. (2020). Characterization of orange passion fruit peel flour and its use as an ingredient in bakery products. *J. Culin. Sci. Technol.* 18(3): 214–230.
- Rivera, S., and Canela-Garayoa, R. (2012). Analytical tools for the analysis of carotenoids in diverse materials. *J. Chromatogr. A.* 1224: 1–10.
- Rivera, S.M., Christou, P., and Canela-Garayoa, R. (2014). Identification of carotenoids using mass spectrometry. *Mass Spectrom. Rev.* 33(5): 353–372.
- Rodríguez-Concepcion, M., Avalos, J., Bonet, M.L., Boronat, A., Gomez-Gomez, L., Hornero-Mendez, D., Limon, M.C., Meléndez-Martínez, A.J., Olmedilla-Alonso, B., Palou, A., and Ribot, J. (2018). A global perspective on carotenoids: Metabolism, biotechnology, and benefits for nutrition and health. *Prog. Lipid Res.* 70: 62–93.
- Romero-Rodríguez, L.M., Tejedor, S., and Montealegre, M.V.P. (2021). Actitudes populistas y percepciones de la opinión pública y los medios de comunicación: Estudio correlacional comparado entre España y Colombia. *Rev. Lat. Comun. Soc.* 79: 43–66.
- Rosales, A., Crossa, J., Cuevas, J., Cabrera-Soto, L., Dhliwayo, T., Ndhlela, T., and Palacios-Rojas, N. (2022). Near-infrared spectroscopy to predict provitamin A carotenoids content in maize. *Agron* 12(5): 1027.
- Ruiz-Armenta, X.A., Zazueta-Morales, J.D.J., Aguilar-Palazuelos, E., Delgado-Nieblas, C.I., López-Díaz, A., Camacho-Hernández, I.L., Gutiérrez-Dorado, R., and Martínez-Bustos, F. (2018). Effect of extrusion on the carotenoid content, physical and sensory properties of snacks added with bagasse of naranjita fruit: optimization process. *CYTA-J Food* 16(1): 172–180.
- Saad, A.G., Pék, Z., Szuvandzsiev, P., Gehad, D.H., and Helyes, L. (2021). Determination of carotenoids in tomato products using Vis/NIR spectroscopy. *J. Microbiol. Biotechnol. Food Sci.* 2021: 27–31.
- Sadeghian, M., Asadi, M., Rahmani, S., Sadeghi, N., Hosseini, S.A., and Zare Javid, A. (2021). Lycopene does not affect prostate-specific antigen in men with non-metastatic prostate cancer: a systematic review and meta-analysis of randomized controlled trials. *Nutr. Cancer.* 73(11-12): 2796–2807.
- Santos, P.D.F., Rubio, F.T.V., Balieiro, J.D.C., Thomazini, M., and Favaro-Trindade, C.S. (2021b). Application of spray drying for production of microparticles containing the carotenoid-rich tucuma oil (*Astrocarum vulgare* Mart.). *LWT* 143: 111106.
- Salehi, F. (2019). Characterization of different mushrooms powder and its application in bakery products: A review. *Int. J. Food Prop.* 22(1): 1375–1385.
- Sancho, L.E.G.-G., Yahia, E.M., and González-Aguilar, G.A. (2011). Identification and quantification of phenols, carotenoids, and vitamin C from papaya (*Carica papaya* L., cv. Maradol) fruit determined by HPLC-DAD-MS/MS-ESI. *Int. Food Res. J.* 44(5): 1284–1291.
- Sayari, N., Sila, A., Balti, R., Abid, E., Hajlaoui, K., Nasri, M., and Bougatef, A. (2015). Antioxidant and antibacterial properties of Citrus paradisi barks extracts during turkey sausage formulation and storage. *Bio-*

- catal. Agric. Biotechnol. 4(4): 616–623.
- Shahidi, F., and Brown, J.A. (1998). Carotenoid pigments in seafoods and aquaculture. *Crit. Rev. Food Sci. Nutr.* 38(1): 1–67.
- Sharifi-Zahabi, E., Soltani, S., Malekhamadi, M., Rezavand, L., Clark, C.C., and Shidfar, F. (2022). The effect of lycopene supplement from different sources on prostate specific antigen (PSA): A systematic review and meta-analysis of randomized controlled trials. *Complement. Ther. Med.* 11: 102801.
- Sharoni, Y., Linnewiel-Hermoni, K., Khanin, M., Salman, H., Veprik, A., Danilenko, M., and Levy, J. (2012). Carotenoids and apocarotenoids in cellular signaling related to cancer: a review. *Mol. Nutr. Food Res.* 56(2): 259–269.
- Silow, C., Axel, C., Zannini, E., and Arendt, E.K. (2016). Current status of salt reduction in bread and bakery products—a review. *J. Cereal Sci.* 72: 135–145.
- Singh, N., Shevkani, K., Kaur, A., Thakur, S., Parmar, N., and Singh Viridi, A. (2014). Characteristics of starch obtained at different stages of purification during commercial wet milling of maize. *Starch-Stärke.* 66(7-8): 668–677.
- Šmídová, Z., and Rysová, J. (2022). Gluten-free bread and bakery products technology. *Foods* 11(3): 480.
- Sommer, A. (2008). Vitamin A deficiency and clinical disease: an historical overview. *Nutr. J.* 138(10): 1835–1839.
- Soncu, E.D., Kolsarıcı, N., Cicek, N., Öztürk, G.S., and Arıcı, Y.K. (2015). The comparative effect of carrot and lemon fiber as a fat replacer on physico-chemical, textural, and organoleptic quality of low-fat beef hamburger. *Korean J. Food Sci. Anim. Resour.* 35(3): 370.
- Stahl, W., and Sies, H. (2012). β -Carotene and other carotenoids in protection from sunlight. *Am. J. Clin. Nutr.* 96(5): 1179S–1184S.
- Stephensen, C.B. (2013). Provitamin A carotenoids and immune function. *Carotenoids and Human Health.* Springer, pp. 261–270.
- Toledo-Martín, E.M., García-García, M.D.C., Font, R., Moreno-Rojas, J.M., Salinas-Navarro, M., Gómez, P., and del Río-Celestino, M. (2018). Quantification of total phenolic and carotenoid content in blackberries (*Rubus Fruticosus* L.) using near infrared spectroscopy (NIRS) and multivariate analysis. *Molecules* 23(12): 3191.
- van Chuyen, H., and Eun, J.-B. (2017). Marine carotenoids: Bioactivities and potential benefits to human health. *Crit. Rev. Food Sci. Nutr.* 57(12): 2600–2610.
- Van Hoang, D., Pham, N.M., Lee, A.H., Tran, D.N., and Binns, C.W. (2018). Dietary carotenoid intakes and prostate cancer risk: a case-control study from Vietnam. *Nutrients.* 10(1): 70.
- van Lent, D.M., Leermakers, E.T., Darweesh, S.K., Moreira, E.M., Tielemans, M.J., Muka, T., Vitezova, A., Chowdhury, R., Bramer, W.M., Brusselle, G.G., and Felix, J.F. (2016). The effects of lutein on respiratory health across the life course: A systematic review. *Clin. Nutr. ESPEN.* 13: e1–e7.
- Varzaru, I., Untea, A.E., and Saracila, M. (2020). In vitro antioxidant properties of berry leaves and their inhibitory effect on lipid peroxidation of thigh meat from broiler chickens. *Eur. J. Lipid Sci. Technol.* 122(4): 1900384.
- Velasco-Arango, V.A., Hleap-Zapata, J.I., and Ordóñez-Santos, L.E. (2021). Nitrite reduction in beef burger using papaya (*Carica papaya* L.) epicarp. *Int. J. Food Sci. Technol.* 27(4): 344–352.
- Wang, C., Li, M., Duan, X., Abu-Izneid, T., Rauf, A., Khan, Z., Mitra, S., Emran, T.B., Aljohani, A.S., Alhumaydhi, F.A., and Thiruvengadam, M. (2022). Phytochemical and Nutritional Profiling of Tomatoes; Impact of Processing on Bioavailability—A Comprehensive Review. *Food Rev. Int.* 15: 1–25.
- Wanjuu, C., Abong, G., Mbogo, D., Heck, S., Low, J., and Muzhingi, T. (2018). The physicochemical properties and shelf-life of orange-fleshed sweet potato puree composite bread. *Food Sci. Nutr.* 6(6): 1555–1563.
- Waqas, M., Shad, A.A., Bashir, O., and Iqbal, M. (2017). Extraction and utilization of tomato carotenoids as antioxidant and natural colorants in sunflower oil and spaghetti. *Sarhad J. Agric.* 33: 2.
- Witczak, M., Ziobro, R., Juszcak, L., and Korus, J. (2016). Starch and starch derivatives in gluten-free systems—a review. *J. Cereal Sci.* 67: 46–57.
- Xu, L., Echeverria-Jaramillo, E., and Shin, W.-S. (2022). Physicochemical properties of muffins prepared with lutein & zeaxanthin-enriched egg yolk powder. *LWT* 156: 113017.
- Yagci, S., Caliskan, R., Gunes, Z.S., Capanoglu, E., and Tomas, M. (2022). Impact of tomato pomace powder added to extruded snacks on the *in vitro* gastrointestinal behaviour and stability of bioactive compounds. *Food Chem.* 368: 130847.
- Zanfani, A., Corbini, G., La Rosa, C., and Dreassi, E. (2010). Antioxidant activity of tomato lipophilic extracts and interactions between carotenoids and α -tocopherol in synthetic mixtures. *LWT - Food Sci. Technol.* 43(1): 67–72.
- Zare, M., Zandi Esfahan, E., and Ghorbani, A. (2019). Forage Quality of *Sal-sola yazdiana* and *S. tomentosa* in Different Growth Stages in Saline Desert of Yazd Province, Iran. *J. Rangel. Sci.* 9: 104–113.
- Zhao, X., Liang, K., and Zhu, H. (2022). Carotenoids in cereals and related foodstuffs: a review of extraction and analysis methods. *Food Rev. Int.* 14: 1–16.