The timing and ecological consequences of Pleistocene megafaunal decline in the eastern Andes of Colombia



Declaration of work

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ABSTRACT

Examining the ecological consequences of Late-Quaternary megafaunal extinctions within biodiversity hotspots is crucial for our understanding of the potential consequences of contemporary extinctions. This research presents the first multi-species record of spores of coprophilous fungi (SCF) from Monquentiva and the high Andean forests of Colombia to reconstruct the Late-Pleistocene and Holocene megafaunal abundance. Fossilised pollen and charcoal are used to examine the consequences of megafaunal declines on the history of the study region. The SCF record indicated the presence of megafauna since at least 30,290 BP; with two periods of megafauna decline at ca. 22,900 and 10,990 BP. At Monquentiva, megafaunal decline in the Early-Holocene resulted in transitional non-analogue vegetation, a loss of some herbivore-dispersed taxa, an encroachment of some palatable and woody flora, and a rise in fire activity. Overall, this research shows that ecosystems in the eastern Colombian Cordillera were highly sensitive to the decline of megafaunal populations. Under the current biodiversity crisis, management and conservation efforts must account for the effect of local herbivore declines on the dispersal of certain plant species, on fire activity and the potential loss of ecosystem services.

Keywords: Pleistocene-Holocene transition; Palaeoecology; Colombian Andes; Coprophilous fungi spores; Megafaunal decline; *Sporormiella*; Multiproxy; Vegetation; Fire.

TABLE OF CONTENTS

SECTION	HEADING	PAGE
	TITLE PAGE WITH DECLARATION	1
	ABSTRACT	2
	TABLE OF CONTENTS	3
	LIST OF FIGURES	5
	LIST OF TABLES	6
	LIST OF ACCOMPANYING MATERIAL	6
	ACKNOWLEDGEMENTS	7
	AUTHORS DECLARATION	10
	DEFINITIONS, ABBREVIATIONS AND SYMBOLS	11
1.	INTRODUCTION	12
	The record of extinction in South America	13
	The ecological consequences of extinction	13
	Spores of coprophilous fungi (SCF)	14
	Research questions and objectives	17
2.	STUDY AREA	18
3.	MATERIAL AND METHODS	21
	Core collection and sampling	21
	Laboratory analysis	23
	Qualitative and quantitative analysis	25
4.	RESULTS	26
	Chronology and stratigraphy	26
	Spores of coprophilous fungi (SCF) pollen and charcoal records	26
	Zone M1 (30,300-22,900 BP)	29
	Zone M2 (22,900-17,890 BP)	29
	Zone M3 (17,890-10,990 BP)	31
	Zone M4 (10,990-6700 BP)	31
	Zone M5 (2150-1150 BP)	32
	Fungal spore correlations	34
	Multivariate ordinations	34

5.	DISCUSSION	37
	The timing of megafaunal extinctions in the northern tropical Andes	37
	Climate, vegetation and human backdrop of megafaunal declines	40
	Ecological consequences of megafaunal collapse	42
	Extant Holocene megafauna, climate and human activity	47
6.	CONCLUSIONS	48
	FUNDING	49
	APPENDICES	50
	REFERENCES	60

LIST OF FIGURES

FIGURE	SECTION AND TITLE	PAGE
	INTRODUCTION	12
1	South American Pleistocene megafauna	12
2	Generalised recreation of the lifecycle of	12
£	spores of coprophilous fungi	10
3	Validation map of existing fungal spore studies from North and South America used to analyse the extinction of Pleistocene megafauna	16
	STUDY SITE	18
4	Location of the study site	19
	MATERIAL AND METHODS	21
5	Bayesian age-depth model and stratigraphic descriptions of the Monquentiva core	22
	RESULTS	26
6	Identifiable fungal spore morphotypes recovered from the Monquentiva record	28
7	Summary diagram of the most abundant spores of coprophilous fungi and pollen taxa recorded from the sediments of Monquentiva	30
8	Monquentiva charcoal record and concentration of key spores of coprophilous fungi	33
9	Canonical correspondence analysis (CCA) results for the pollen, charcoal and spores of coprophilous fungi records from Monquentiva	35
	DISCUSSION	37
10	A conceptual landscape drawing of the changes in megafaunal presence, vegetation and fire activity reconstructed for Monquentiva during the Late-Pleistocene and Holocene	43

LIST OF TABLES

SECTION	SECTION AND TITLE	PAGE	
1	MATERIAL AND METHODS Accepted radiocarbon dates (¹⁴ C) with Bacon output calibrated ages (cal yr BP) for the Monquentiva core	21 23	
2	RESULTS Correlations of variation in spore concentration amongst key spores of coprophilous fungi (key SCF) from the Monquentiva core	26 34	

APPENDICES

ТҮРЕ	TYPE SECTION AND TITLE			
Figures		50		
Appendix A	Complete fossil pollen and charcoal diagram for all pollen taxa and charcoal particles recorded from the sediments of Monquentiva	50		
Appendix B	Complete macroscopic and microscopic charcoal fractions expressed as counts per cubic centimetre (cm ³) and area (cm ² /cm ³)	51		
Appendix C	Complete fossil spore diagram for all identifiable fungi spore morphotypes recovered from the sediments of Monquentiva	52		
Tables		53		
Appendix D	Oxcal v.4.4 outputs of a general outlier model run on the Monquentiva ¹⁴ C radiocarbon ages	53		
Appendix E	Coefficient correlations between all identifiable fungi spores recorded from the sediments of Monquentiva	54		

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AUTHORS' DECLARATION

This thesis manuscript is comprised of a supplemented publication (Pym *et al.*, 2022). The following declaration summarises a statement from the thesis author regarding the contributions made to the thesis and to the co-authored publication. We certify that the manuscript author led the completion of the supplemented thesis. The contributions made in the development of the publication are as followed:

Felix Pym: Conceptualization, Methodology, Software, Validation, Formal analysis, Investigation, Writing – Original Draft, Writing – Review & Editing. Felipe Franco-Gaviria: Methodology, Software, Formal analysis, Writing – Review & Editing. Ismael G. Espinoza: Investigation, Visualisation, Writing – Review & Editing. Dunia H. Urrego: Supervision, Funding acquisition, Conceptualization, Methodology, Validation, Writing – Review & Editing. All authors supported the production process and approved the ideas for its publication.

DEFINITIONS, ABBREVIATIONS AND SYMBOLS

Term	Definition
SCF	Spores of coprophilous fungi
Key SCF	All identifiable spores of coprophilous and semi-coprophilous fungi
	grouped to create a sum of spores associated with megafaunal
	abundance
Abbreviation	Meaning
Cal yr BP	Radiocarbon dated (¹⁴ C) years before present
ТРА	Total pollen assemblage
ITCZ	Intertropical Convergence Zone
ENSO	El Niño-Southern Oscillation
STARS	Sequential t-test analysis of regime shifts
RSI	Regime shift index
CCA	Canonical correspondence analysis
CHAR	Interpolated charcoal concentration
BP	Calendar years before present
Symbol	Meaning
ha	hectares
km	Kilometre
masl	Metres above sea level (m)
cm	Centimetre
μm	Micrometres

1. INTRODUCTION

Megafauna include mammals that weigh over 45 kg and as ecological keystone species, they can have an important ecological role in shaping habitats and entire ecosystems (Martin and Klein, 1984; Mills et al., 1993; Gill et al., 2009). Megafauna can modify vegetation composition (Gordon et al., 2021), influence seed-dispersal (Peres et al., 2016), maintain flora coexistence (Sankaran et al., 2005), shape soil nutrients and fertility (Wigley et al., 2020) and suppress fire activity (Waldram et al., 2008). In modern savannas, antelope-induced grazing suppresses grass and tree growth (Staver and Bond, 2014) while rhinos can limit fire activity by feeding on grasses and controlling fuel build-up (Waldram et al., 2008). A large proportion of megafaunal species went extinct on all continents during the Pleistocene (Barnosky et al., 2004), and given their ecological importance the consequences of these extinctions are thought to be significant (Johnson, 2009; Malhi et al., 2016). The cause of extinction remains debated; however, studies have linked climatic changes, human activity and even an extraterrestrial impact to their decline (Firestone et al., 2007; Cooper et al., 2015; Raczka et al., 2019). South America lost an estimated 80% of its megafauna, including ground sloths (Megatherium americanum) and glyptodons (Glyptodon clavipes) (Barnosky et al., 2004; Wroe et al., 2004; Lima-Ribeiro and Diniz-Filho, 2013) (Fig. 1). Despite experiencing one of the most significant declines of megafaunal species, information about South American megafaunal extinctions remains limited.



Figure 1. An artist illustration of some South American Pleistocene megafaunal species that inhabited Colombia (Carreño, 2017).

The record of extinction in South America

The timing of the Pleistocene megafaunal extinction in South America is yet to be determined as records are scarce (e.g. Elias and Schreve, 2007; Rozas-Dávila et al., 2016; Raczka et al., 2018; Raczka et al., 2019). In North America, robust archaeological evidence suggests the timing of megafaunal extinction between 11,500 and 11,000 years ago (Barnosky et al., 2004). However, extinctions may have occurred earlier, between 14,000 - 12,700 cal yr BP (calibrated years before present, hereafter BP) (Robinson et al., 2005; Gill et al., 2009; 2012; Perrotti, 2018). Perhaps in relation to overhunting resulting from humans arriving in North America as early as 15,500 years ago (Waters *et al.*, 2011). In South America, the extinction window has been suggested to have occurred between 15,000 and 10,000 years ago (Barnosky et al., 2004). The limited records available suggest variability in megafaunal extinction timings. In the Peruvian Andes, researchers found the local extinction occurring as early as 15,800 BP (Rozas-Dávila et al., 2016). Whereas in the Ecuadorian Andes extinction was recorded much later, by 12,800 BP (Raczka et al., 2019). This timing is similar to the extinction recorded in the Brazilian cerrado lowlands, reached by 12,000 BP (Raczka et al., 2018). However, in Argentina and Brazil fossilised megafaunal bones have been dated to the early-Holocene as late as 6560 BP (Borrero et al., 1998; Steadman et al., 2007). Thus, suggesting the early-Holocene survival of some megafauna (Neves and Piló, 2003). The available information in South America leaves a broad range, between 15,800 - 6560 BP, on the timing of Pleistocene megafaunal decline in the region and within different ecosystems.

The ecological consequences of extinction

Late-Pleistocene megafaunal extinctions likely had significant ecological consequences on the ecosystems we know today but available records vary. The extinction of the megafauna in North America led to the formation of flora without modern analogues (hereafter non-analogue) and increased fire activity (Gill *et al.*, 2009, 2012). In the Peruvian Andes, the collapse of megafauna was followed by a rise in fire activity and species turnover (Rozas-Dávila *et al.*, 2016). In the Pampas lowlands, megafaunal loss was apparently not associated with the expected rise in shrubby vegetation and fire activity was not observed in the

record (Prieto, 2000). In the Brazilian cerrado, the ecosystem remained largely stable but experienced an increase in some palatable species (Raczka *et al.,* 2018). Despite these recent advancements, it remains unclear what the ecological consequences of extinctions were in other high biodiversity regions of South America or whether these consequences were habitat specific.

Spores of coprophilous fungi (SCF)

Research of the Pleistocene megafaunal extinction and their impact on the Quaternary landscape in South America has used radiocarbon-dated bone remains recovered from, for example, archaeological sites (Borrero, 2009; Barnosky and Lindsey, 2010; Lima-Ribeiro and Diniz-Filho, 2013). The collagen extracted from fossilised bones provides accurate dating, however, they are rare, isolated and often, geographically concentrated (Johnson et al., 2015; van Asperen et al., 2020). The use of spores of coprophilous fungi (SCF) to reconstruct megafaunal presence can reduce these limitations because they preserve well in stratigraphic units where chronological frameworks can be built using several ages (van Geel et al., 2007; 2011; Lee et al., 2022). SCF grow and reproduce on herbivore faeces, they are spread to surrounding vegetation where they can be ingested and excreted by megafauna (Davis and Shafer, 2006; Perrotti and van Asperen, 2019) (Fig. 2). Therefore making SCF a reliable indicator for the presence/absence and abundance of megafauna (Wood and Wilmshurst, 2013; Johnson et al., 2015). SCF such as Sporormiella can also reproduce in the dung of smaller herbivores (Feranec et al., 2011). However, SCF reproduction is limited to the size of the faeces (Halbwachs and Bässler, 2020). Therefore, due to the comparative volume of dung produced, the contribution made to the production of SCF by megafauna overrules any signal associated with these smaller fauna (Gill et al., 2013; Lee et al., 2022). SCF can be found in conjunction with other palaeoecological proxies including pollen and charcoal allowing an understanding of the consequences of megafaunal extinctions on vegetation and fire activity (Raczka et al., 2018; van Asperen et al., 2020).

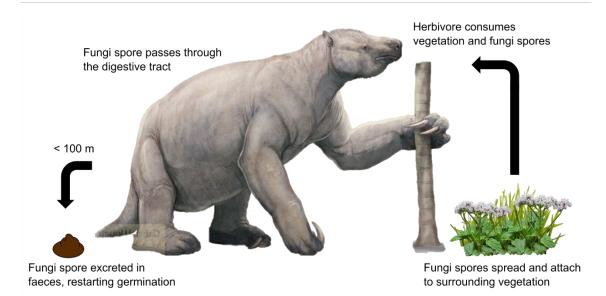


Figure 2. Generalised recreation of the lifecycle of spores of coprophilous fungi (SCF). The depiction is based on descriptive spore lifecycle analysis (Davis and Shafer, 2006; Perrotti and van Asperen, 2019). The giant ground sloth (*Eremotherium*) concept art sourced from Ugueto (2020).

Since its first discovery, the Sporormiella type fungi spore dominates the reconstruction of megafaunal presence and abundance. Within contemporary sites, a threshold of >2% of the total pollen assemblage (TPA) possibly represents an active abundance of megafauna (Davis and Shafer, 2006; Gill et al., 2013). This threshold has been extrapolated to longer time scales to identify functional extinction events, which is when Sporormiella declines <2% suggesting that megafauna become so rare as to cease imposing a signal on the landscape (Davis and Shafer, 2006; Raczka et al., 2018). However, the functional extinction threshold from an arid region of Western North America (Davis, 1987) is dependent on the pollen productivity of the system (Raczka et al., 2016). Thus, extrapolation between environments may not be indicative, so using other quantifications in conjunction with percentage is recommended (Raper and Bush, 2009; Perrotti and van Asperen, 2019). Furthermore, other SCF can reflect the presence of megafauna (Davis, 1987; Perrotti and van Asperen, 2019). Actually, the sum of all SCF may provide the most accurate measure for analysis and enhance the interpretation of palaeoecological spore records (Johnson et al., 2015; Conroy et al., 2020). Despite the information provided by SCF, just three studies in South America to date have used SCF to reconstruct Pleistocene megafaunal presence (Rozas-Dávila et al., 2016; Raczka et al., 2018; Raczka et al., 2019) (Fig. 3). This compares with at least fourteen studies in North America, a continent that lost less genera of megafauna (72%) (Barnosky et al., 2004).



However, the studies in South America have relied on the percentage of *Sporormiella* as the only SCF indicative of megafaunal presence.

Figure 3. Validation map of existing spores of coprophilous fungi studies from North and South America. The mango polygon shows Colombia, the country of interest in this research. The lilac dots represent published studies and the mango dot unpublished work (Pym, 2020). Published North American sites (Davis *et al.*, 1984; Davis 1987; Robinson *et al.*, 2005; Robinson and Burney, 2008; Davis and Shafer, 2006; Gill *et al.*, 2009; Gill *et al.*, 2012; Perrotti, 2018; Conroy *et al.*, 2020) and South American sites (Rozas-Davila *et al.*, 2016; Raczka *et al.*, 2018; Raczka *et al.*, 2019). Service Layer Credits^a: Esri.

Research questions and objectives

The rationale of the research presented in this thesis was to contribute to the palaeoecological literature of Pleistocene megafaunal extinctions. This research presents the first investigation on the timing and ecological consequences of megafaunal declines from Monquentiva, a diverse ecosystem in the eastern Colombian Cordillera.

In accordance with this rationale, this research looked to address three questions:

- (1) When did the Late Pleistocene megafaunal extinction occur in the Colombian northern tropical Andes?
- (2) What was the climate, vegetation and human background of megafaunal decline?
- (3) Did any ecological consequences occur that could be attributed to a megafaunal loss?

In order to answer these questions, the objectives of this study were as follows:

- (1) Analyse a well-dated sedimentary record using a multi-species SCF approach to investigate megafaunal presence
- (2) Assess the interpretative value of all identifiable SCF
- (3) Combine the SCF record with a multi-proxy record of microscopic pollen and charcoal to evaluate the ecological consequences of megafaunal declines in the eastern Andes of Colombia

2. STUDY AREA

Pantano de Monguentiva (hereafter Monguentiva) is a peat bog that lies in the high Andean slopes of the eastern Colombian Cordillera within a ca. 4.28 ha natural park (4°54"N, 73°45"W) (Fig. 4). The site is found in the jurisdiction of the Municipality of Guatavita, Cundinamarca, roughly 50 km northeast of the city of Bogotá. Monquentiva lies in an altitudinal strip between 2900-3500 metres above sea level (hereafter masl). Despite this elevation, the site is a relatively flat ovoidal area with slopes of less than 5° (Serrano, 2019; Espinoza et al., 2022). Small streams upon the eastern side of the mountain range contribute to the site's hydrology, which feeds into the Orinoco river basin (Avella-M et al., 2014). During pre-Hispanic times, Monquentiva was prone to flooding and may have contained an extensive lagoon (Avella-M et al., 2014; Espinoza et al., 2022). However, the contemporary peatbog underwent transformations through the establishment of colonial drainage systems for livestock (Gomez and Rubio, 2003; Avella-M et al., 2014; Amador and Millner, 2019). The shape and position of Monguentiva are controlled by the Río Blanco - Machetá Anticline fold and Machetá fault (Kammer et al., 2020; Espinoza et al., 2022) built up upon sedimentary Cretaceous rocks (Kammer et al., 2020).

Monquentiva is characterised by a low 11°C average annual temperature (CAR, 2007; Rojas *et al.*, 2010). The mean annual precipitation in Monquentiva is 1425 mm, with a unimodal regime of heaviest rainfall between April and November (CAR, 2000; 2007; Avella-M *et al.*, 2014). The main climatic factors influencing Monquentiva are the annual migration of the Intertropical Convergence Zone (ITCZ) bringing moisture from the Amazon and the Atlantic Ocean and the irregular periodic variation in winds and sea surface temperatures associated with the El Niño-Southern Oscillation (ENSO) (Poveda *et al.*, 2005; Poveda *et al.*, 2006). The topography of the Andean mountain range also plays an influential role. This orographic barrier interrupts the easterly airflows from the Amazon basin forcing the air to rise, covering the slopes of the eastern Andean flank with fog (Poveda *et al.*, 2005).

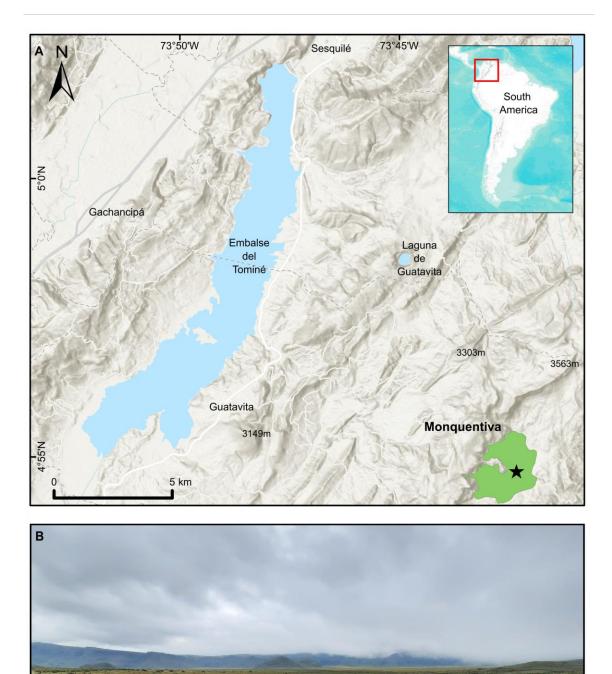


Figure 4. Location of the study site. (A) Map of Pantano de Monquentiva (green polygon), Colombia (red square). The black star shows the location of the coring site. (B) Field photograph of the Monquentiva peat bog; photo taken by Pym, 2022.

The climate, geographical location and topography of Monquentiva favour a mosaic of vegetation and concentration of biodiversity along a narrow altitudinal gradient in which the High Andean Forest, Subpáramo and Páramo are the most abundant vegetation types (Langebaek, 1985; Avella-M *et al.*, 2014; Espinoza *et al.*, 2022). High Andean forest is the most represented vegetation type surrounding Monquentiva today (76%), in which *Weinmannia* and *Clusia* dominate (Avella-M *et al.*, 2014). Other abundant arboreal genera include *Brunellia, Clethra, Drimys, Hedyosmum, Ternstroemia*, and *Vallea* (Rangel-Ch, 2000; Avella-M *et al.*, 2014). The Subpáramo follows in representation and is made up of shrubby species including *Aragoa, Arcytophyllum, Diplostephium, Hypericum, Gaultheria, Paepalanthus* and *Puya* (Avella-M *et al.*, 2014). The Páramo vegetation represents >20% of the flora in Monquentiva and contains a varied number of vegetation types dominated by grasses and herbs (Avella-M *et al.*, 2014). Poaceae is the largest component of the Páramo including species such as *Calamagrostis* and *Espeletia* (Rangel-Ch, 2000; Avella-M *et al.*, 2014).

Monquentiva's ecological richness and mosaic of vegetation make the site an attractive habitat for extant megafauna. This includes species of megafauna that survived the Pleistocene extinction, such as the white-tailed deer (Odocoileus virginianus), Brocket deer (Mazama pita) and Andean bear (Tremarctos ornatus) (Cooke, 1998; García-Rangel, 2012; Martínez-Polanco, 2019). Macrofossil evidence for extinct Pleistocene megafauna is limited in Colombia (Barnosky and Lindsey, 2010). However, skeletal records of Mastodons (Haplomastondon) have been dated between ca. 40,000 and 15,000 B.P in Bogotá (van der Hammen and Correal-Urrego, 1978). Megafaunal remains have also been found in Tocaima, an archaeological site at 340 masl (Correal-Urrego, 1993; Correal-Urrego, 2005; Alfonso-Rojas et al., 2021). In Tocaima, massive bones of gomphotheres (Notiomastodon platensis) and isolated bone fragments associated with bones of giant ground sloths (Eremotherium) have been recovered. These findings alongside lithic artefacts associated with the genus gomphothere (Cuvieronius and Stegomastodon) recovered from Tibitó at ca. 2500 masl provides evidence for an ecosystem rich in megafauna in Bogotá during the Late-Pleistocene and the diversity of species found in Colombia (Correal-Urrego, 1981; Prado et al., 2005; Borrero, 2009; Alfonso-Rojas et al., 2021). However, at higher elevations such as Monquentiva, no macrofossil remains are yet reported in the literature.

3. MATERIAL AND METHODS

Core collection and sampling

A 3.65-m sediment core labelled MAR19-II was recovered from Monquentiva using a modified Livingston piston corer (Colinvaux *et al.*, 1999) in 2019. The core was transported in sealed aluminium tubes to the Tropical Palaeoecology Laboratory at the University of Exeter. Subsequently, the cores were open lengthwise and stored at 4°C. The Troels-Smith method (Kershaw, 1997) was used to describe stratigraphic units in the four 1-m core sections. Sixteen bulk sediment samples were extracted and sent to the Radiocarbon dating facility at the Queen's University Belfast for ¹⁴C dating (Table 1). The intervals for ¹⁴C dating were selected by visual analysis of the sediments' stratigraphic changes and to give equitable dating of the whole core. A chronological model was built using the Bacon package v.2.3.9.1 (Blaauw and Christen, 2013) within RStudio (RStudio Team, 2020) (Fig. 5).

This study focused on the upper two drives, Drive 1 (from 40-77 cm) and Drive 2 (78.1-177 cm) of the MAR19-II sediment core. Volumetric samples of 1 cm³ were extracted for fungal spore quantification between 40 and 137.1 cm to cover the Late-Pleistocene and early-Holocene. Samples of 1 cm³ for charcoal and 0.5 cm³ for pollen were also extracted between 40 and 137.1 cm core depth.

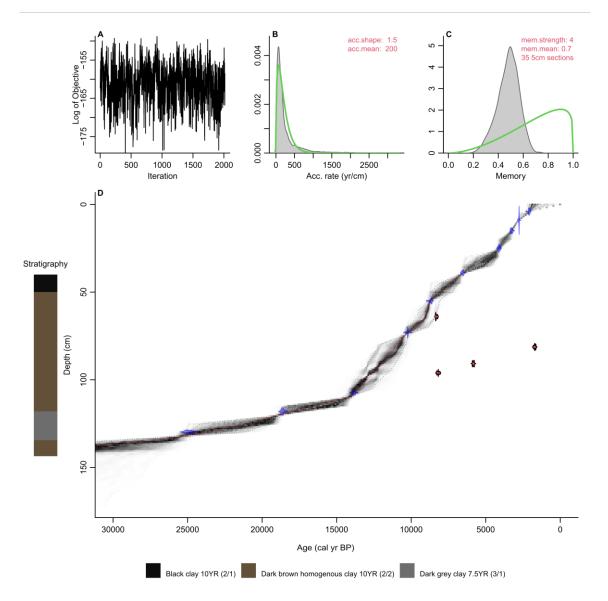


Figure 5. Bayesian age-depth model and stratigraphic descriptions for the Monquentiva core. The inset figures show the models: (A) Markov Chain Monte Carlo (MCMC) iterations; (B) distributions for the accumulation rate (Bacon MCMC iterations estimates the accumulation rate in years/cm; so more correctly, sedimentation times); (C) the memory of variability; and (D) the calibrated ¹⁴C dates (transparent blue) and the age-depth model; red curve shows the 'best' model based on the weighted mean age for each depth. The filled red radiocarbon dates represent the outliers identified using Oxcal analysis (Ramsey, 2009a, b). Stratigraphic descriptions include Munsell colour notations (Munsell Colour, 1992).

CorelD	LabID	Drive	Depth (cm)	Description	Age (BP)	Error (±)	Age (calibrated range, 2σ)
MAR19-II	UBA-45475	1	4	Organic lake mud	2115	26	1990-2160
	UBA-44495	1	9	Organic lake mud	2647	25	2720-2760
	UBA-45476	1	15	Organic lake mud	3047	30	3110–3380
	UBA-44496	1	25	Organic lake mud	3750	28	3890–4270
	UBA-42144	1	39	Organic lake mud	5734	36	6300–6700
	UBA-43589	1	55	Organic lake mud	7868	37	8480–9000
	UBA- 44497*	1	64	Organic lake mud	7499*	37	8990-10090
	UBA-44498	1	73	Organic lake mud	9077	46	10000-10550
	UBA- 45477*	2	81.3	Organic lake mud	1786*	30	10520-12100
	UBA- 44499*	2	90.8	Organic lake mud	5068*	31	11120-13190
	UBA- 45478*	2	96.1	Organic lake mud	7367*	40	11600-13620
	UBA-44500	2	107.4	Organic lake mud	11981	63	13300-14400
	UBA-45479	2	118	Organic lake mud	15347	69	16990–18740
	UBA-43590	2	129.7	Organic lake mud	20707	103	21090–25270
	UBA-45480	2	142.5	Organic lake mud	36818	598	30710–41500
	UBA-42145	2	162.7	Organic lake mud	42623	162.7	36310–48260

Table 1: Radiocarbon ages (¹⁴C) from organic lake sediments and calibrated ages (cal yr BP, IntCal20, Reimer et al., 2020) for the Monquentiva core. Calibrated ranges are rounded to the nearest 10 years.

Laboratory analysis

Fungal spores were prepared following a modified version of Faegri *et al.* (1989). This modification removed acetolysis, which can cause spore deterioration (van Asperen *et al.*, 2016). A known number of exotic *Lycopodium* spores (Batch 1031; 20,848 spores/tablet) was added for SCF and pollen analysis before processing. Percentages were calculated based on the total pollen assemblage (hereafter %TPA) and the *Lycopodium* spores were used to calculate concentrations per volume (Stockmarr, 1972). SCF treatment included 10% HCl and 10% KOH, warming in a 90°C ultrasonic bath, sieving at 180 µm, density separation using sodium metatungstate Na₆(H₂W₁₂O₄₀) at 1.9-2.0 grams/litre and suspending in glycerol (van Asperen *et al.*, 2016). Spores and pollen were identified using a Zeiss Axioskop light microscope at 400x and 1000x magnification. All fungal spores were quantified until 300 *Lycopodium* were counted per sample (Etienne and Jouffrey-Bapicot, 2014).

The fungal spore quantification included the commonly-used *Sporormiella* taxon as well as all identifiable coprophilous, semi-coprophilous and non-coprophilous spores. The spore identification was conducted using published descriptions, fungal keys and images (van Geel *et al.*, 2003; Cugny *et al.*, 2010; Guarro *et al.*, 2012; Revelles *et al.*, 2016; Loughlin *et al.*, 2018). Where identification at the taxa level became difficult, spores were assigned to their closest morphotype (Perrotti and van Asperen, 2019; van Asperen *et al.*, 2020). Spore morphotypes were assigned to their primary substrate following existing associations (Perrotti and van Asperen, 2019; van Asperen *et al.*, 2020).

The pollen and charcoal records were used to reconstruct vegetation changes and past fire activity. Pollen and charcoal samples were prepared following their respective standard protocols (Faegri et al., 1989; Whitlock and Larsen, 2002). A pollen sum of 300 pollen grains per sample were identified and counted using keys and descriptions for Andean palynology (Hooghiemstra, 1984; Colinvaux et al., 1999; Bush and Weng, 2006; Velásquez et al., 2022) and modern reference material archived in the tropical pollen reference collection at University of Exeter. The ecological and habitat affinity of pollen taxa were extracted from the literature (e.g. van der Hammen and González, 1960; Hooghiemstra, 1984; Marchant et al., 2002; Bernal et al., 2019; Espinoza et al., 2022). Charcoal particles were sieved to retain fragments >180 µm (macroscopic) and 106 – 63 µm (microscopic) (Whitlock and Larsen, 2002). The retained particles were manually separated and photographed using a Zeiss Stemi stereo microscope at 50x magnification. Subsequently, charcoal area and raw counts (Clark and Patterson, 1997; Halsall et al., 2018) were extracted using image analysis in ImageJ software (Abràmoff et al., 2004). The macroscopic fraction were extracted at a 0.8x zoom using a set calibration scale of 106 pixels/µm and the microscopic fraction at a 1.25x zoom and 170 pixels/µm calibration scale.

Qualitative and quantitative analysis

Stratigraphic diagrams of selected pollen taxa were plotted using C2 v.1.7.7 (Juggins, 2007). Confidence limits were calculated for the key spore concentration data to assess uncertainty (Maher, 1981: 179). A Pearson correlation coefficient was calculated to test correlations between key spore taxa (Johnson *et al.*, 2015) using SPSS (IBM Corp, 2020). SCF that showed positive

and significant correlations were grouped to create a multi-species group of key SCF associated with megafaunal abundance (Dix and Webster, 1995; Johnson et al., 2015; Perrotti and van Asperen, 2019). A sequential t-test analysis of regime shifts (STARS) (Rodionov, 2004) was performed using key SCF concentration to identify statistically significant deviations in megafaunal abundance. The analysis utilised a regime shift index (hereafter RSI) to quantify the size of the regime shift (Rodionov and Overland, 2005) and was calculated using a cut-off length l = 4 and probability level p = 0.05. STARS analysis was deployed using the Package rshift (Room et al., 2022). Canonical correspondence analysis (CCA) (Oksanen, 2009) was performed to test the influence of constraining variables (megafaunal abundance and fire activity) on the distribution of vegetation. CCA analysis was performed on key SCF concentration, macroscopic and microscopic charcoal counts, and pollen percentages. The CCA was performed using the R package vegan (Oksanen et al, 2007), and included every pollen taxon quantified with an abundance over 1% TPA to reduce the weighting of rare taxa (Urrego et al., 2009; Urrego et al., 2013). Both STARS and CCA were performed within RStudio (RStudio Team, 2020).

To reconstruct local fire history and identify significant fire events near Monquentiva (Clark *et al.*, 1996; Higuera *et al.*, 2009), raw charcoal counts were converted into interpolated charcoal concentration (hereafter CHAR, particles/cm³) and separated into peak magnitude and peak frequency using CharAnalysis v.1.1 within Matlab (Higuera, 2009, MathWorks, 2019). Zone division for charcoal in CharAnalysis used the whole record of samples with depth data from the youngest to oldest age (6,600–32,000 BP). CHAR values were interpolated to 200-year intervals based on rates of sediment accumulation and median sample resolution (Espinoza *et al.*, 2022). Charcoal frequency (C_{back}) was estimated using a LOWESS smoother method with a 1000-year smoothing record (Higuera, 2009; Espinoza *et al.*, 2022).

4. RESULTS

Chronology and stratigraphy

The 97.5-cm sediment section analysed provided a record of sediment accumulation from 30,290 to 6700 BP (Table 1, Fig. 5). Before age model construction, four of the sixteen-¹⁴C ages were detected as outliers in Oxcal v.4.4 (Ramsey, 2009a), using the general outlier model (Ramsey, 2009b, Appendix D). These outliers were excluded from the age-depth model as they likely represented age reversals (Trachsel and Telford, 2017). The chronological model for MAR19-II (Fig. 5) used acc.shape=1.5 and acc.mean=200 as priors. The radiocarbon dates were calibrated using the IntCal20 calibration curve (Reimer et al., 2020). Five samples at the top of drive 2 in the core were realigned after ¹⁴C ages revealed they represented Holocene sediment. Locked pistons generally do not slip when pushed through lake muds (Colinvaux et al., 1999) and sediment mixing is rare. However, maintaining an efficient core seal is complex and in this case pressure may have released the rubber seal of the piston (Colinvaux et al., 1999; Last and Smol, 2002) and Holocene sediments were collected at the top of drive 2. In the basal part of the drive, between 143.5 and 135 cm depth (ca. 38,820-27,740 BP) the sediment is characterised by dark brown homogenous clay (Munsell colour 10YR 2/1) (Fig. 5). A sharp boundary from 135 cm is followed by a 17 cm layer of very dark grey clay (7.5YR 3/1) sediment up to 118 cm (ca. 17,900 BP). From 118 to 92.5 cm depth (17,900-12,350 BP) the sediments returned to a dark brown homogenous clay (Munsell colour 10YR 2/2). This section is overlain at the top of drive 2 with a layer of black homogenous clays (10YR 2/1) that extends to 50 cm depth (ca. 8080 BP) in MAR19-II drive 1.

Spores of coprophilous fungi (SCF) pollen and charcoal records

The SCF record was composed of 28 identifiable taxa, including eight key SCF (Fig. 6). SCF were found preserved in 36 of the 52 subsamples analysed (van Geel *et al.*, 2007). In total, 1062 fungal spores were counted, with an average of 20 spores per sample (minimum 0; maximum 118). Confidence intervals calculated for key SCF concentrations showed that the uncertainties attributable to count size were minimal as the intervals are narrow (Fig. 7), this suggested the sampling effort was sufficient to provide statistical validity to the

Page | 27

key spore data (Maher *et al.*, 2012). The SCF and pollen record was divided into five palynological zones based on changes in the core stratigraphy, key SCF regime shifts, and prominent changes in SCF and pollen assemblages (Fig. 7 and see Appendix A for complete pollen record). This includes zone M5, a chronologically floating zone dating between ca. 2,150 and 1,170 BP. Charcoal counts and area showed no observable differences in the Late Holocene ca. 2150 - 1170 BP (Appendix B). The CHAR concentration, peak magnitude and frequency of fires in Monquentiva were greater in the Early to Mid-Holocene samples (Fig. 8). CHAR analysis could not be run on the charcoal data from zone M5 due to the low number of samples.

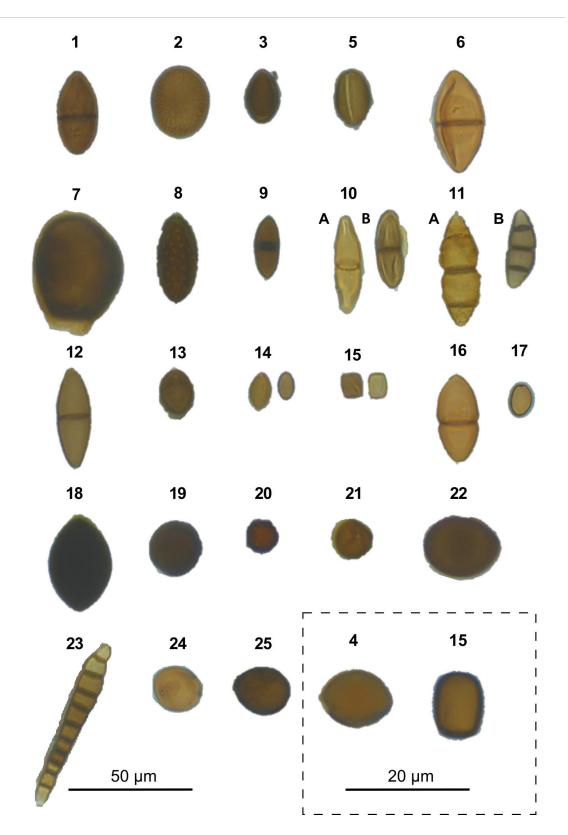


Figure 6. Identifiable fungal spore morphotypes recovered from the Monquentiva record. Due to its size, morphotype 4 (*Chaetomium*) is photographed at 1000-x magnification. Three taxa were not included in the figure due to image quality. *Sporormiella* (morphotype 15) is photographed at both magnifications. 1, *Arnium imitans* (*A.imitans*); 2, *Achaetomium*; 3, *Cercophora*; 4, *Chaetomium*; 5, *Coniochaeta*; 6, *Delitschia*; 7, *Endophragmiella*; 8, *Gelasinospora*-types; 9, 0U-5; 10, 0U-18 AFF; 11, OU-100 AFF; 12, OU-108; 13, *Podospora*; 14, *Sordaria*. 15, *Sporormiella*; 16, *Trichodelitschia*; 17, UAB-1; 18, UAB-2; 19, UAB-7; 20, UAB-15; 21, UAB-30A; 22, UAB-32; 23, UAB-40; 24, UAB-48; 25, UAB-50. Morphotypes OU- and UAB- identified using keys from Loughin *et al.* (2018) and Revelles *et al.* (2016).

Zone M1 (30,300-22,900 BP)

In this zone, the highest abundance of SCF was observed (Fig. 7). *Sporormiella* peaked at 417 spores/cm³ (< 1%), *Sordaria* at 1737 spores/cm³ (3%), and *Podospora* at 139 spores/cm³ (< 1%). The record of key SCF also included *Arnium imitans* (1946 spores/cm^{3,} 3%), *Coniochaeta* (695 spores/cm^{3,} 1%), *Delitschia* (208 spores/cm^{3,} < 1%) and *Cercophora* (139 spores/cm^{3,} < 1%). The most abundant pollen taxa were Poaceae (49%), Asteraceae (16%) and *Alnus* (15%) (Fig. 7). Other prominent tree and shrub taxa occurring at < 5% are *Hedyosmum, Hypericum*, Melastomataceae, *Podocarpus* and *Weinmannia*. Before 25,000 BP, macroscopic and microscopic charcoal concentrations were low (<20 particles/cm³), but increased to 91 and 157 particles/cm³, respectively at ca. 24,240 BP (Fig. 8). Peak frequency for both charcoal fractions remained below <2 fires per 1000 yr. The STARS analysis identified a key SCF regime shift at 24,240 BP reaching 2.3 RSI (Fig. 7).

Zone M2 (22,900-17,890 BP)

Zone M2 is characterised by the near absence of SCF (Fig. 7). OU-100 and UAB-2 were the only fungal spores recovered ca. 19,900 and 17,890 BP (69 spores/cm³) (Appendix C). Asteraceae (36%) and *Alnus* (25%) were the most abundant pollen taxa, while Poaceae remained relatively low (20%) (Fig. 7). *Hedyosmum* (4%), *Podocarpus* (4%), *Valeriana* (2%) and *Weinmannia* (1%) were all present. Arboreal taxa such as *Morella* and *Myrsine* increased slightly (1-2%). Macroscopic (<2 particles/cm³) and microscopic (0-29 particles/cm³) charcoal remained low (Fig. 8). Peak frequency for both charcoal fractions increased in this zone (between 0 and 2.5 fires per 1000 yr). One key SCF regime shift was detected in the zone, reaching 1.23 RSI at 17,890 BP (Fig. 7).

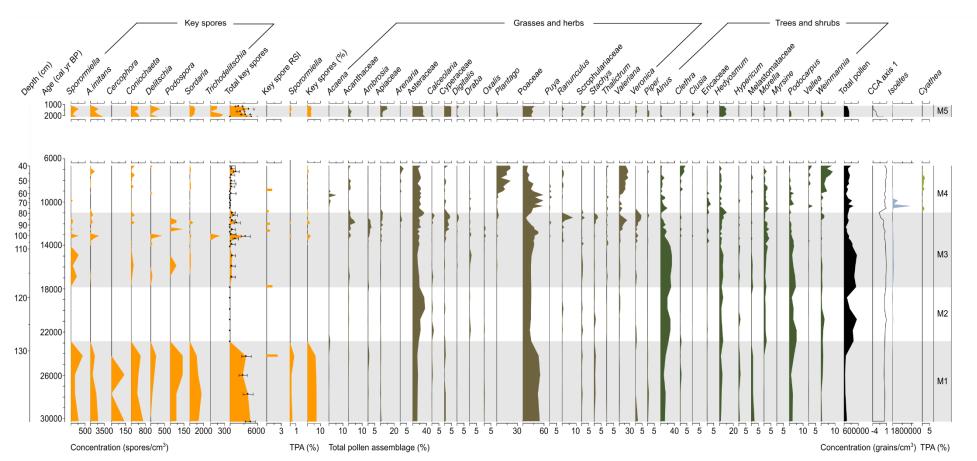


Figure 7. Summary diagram of the most abundant spore and pollen taxa recorded from the sediments of Monquentiva. All spores are expressed as a concentration per cubic centimetre (spores/cm³). Key SCF concentration shows the 0.95 confidence interval (Maher, 1981). The key SCF RSI shows the regime shift analysis and plots the statistically significant shifts in key SCF concentration.

Zone M3 (17,890-10,990 BP)

In zone M3 SCF rebounded to levels comparable to those recorded in zone M1 (Fig. 7). Key SCF concentration fluctuated between 0 and 2154 spores/cm³ (0-3%). Sporormiella increased within this zone (278 spores/cm³, < 1%), along with Arnium imitans (3336 spores/cm³, 2%), Delitschia (416 spores/cm³, < 1%) and Sordaria (695 spores/cm³, < 1%). Podospora (139 spores/cm³, <1%), Coniochaeta (208 spores/cm³, <1%) and Trichodelitshia $(208 \text{ spores/cm}^3, < 1\%)$ were also present. Herbaceous taxa such as Asteraceae (20%) and Poaceae (27%) dominated in this zone (Fig. 7). Acanthaceae (1-5%), Ambrosia (0-3%), Apiaceae (0-4%), Cyperaceae (0-4%), Draba (0-2%), Ranunculus (0-8%) and Valeriana (0-24%) all increased in abundance. Alnus (29%) and Podocarpus (4%) increased again up to ca. 14,150 BP before declining to 2% and 0% respectively by 10,990 BP. Hedyosmum (4%) and Melastomataceae (1%) were present whilst Ericaceae (0-2%) and Morella (1-4%) increased. Macroscopic charcoal was absent at 17,890 BP but increased to 59 particles/cm³ by 11,200 BP alongside the largest macroscopic charcoal peak recorded (25 particles/cm⁻² peak⁻¹) (Fig. 8). Peak frequency steadily increased (from 2 to 3 fires per 1000 yr). Microscopic charcoal increased from 0-65 particles/cm³ across the zone. By 14,200 BP microscopic peak frequency decreased to <1 fires per 1000 yr before rising to >3 fires per 1000 yr by 12,000 BP. The regime shift analysis recorded two key SCF changes at 12,770 and 12,140 BP (0.681 and 0.621 RSI) (Fig. 7).

Zone M4 (10,990-6700 BP)

Zone M4 showed a reduction of all SCF (Fig. 7). Between 11,915 and 9720 BP, key SCF declined from 1459 to 0 spores/cm³ (2-0%). *Sporormiella* was found in only one sample at 9890 BP whilst *Cercophora, Podospora* and *Trichodeltischia* were absent. At ca. 7220 BP, *Arnium imitans* (1112 spores/cm³, 1%) and *Sordaria* (139 spores/cm³, < 1%) increased, aligning with a rise in key SCF to 1251 spores/cm³ (> 1%). Most grasses and herbs declined at the onset of this zone including Cyperaceae (2-0%), Acanthaceae (2-0%), *Ambrosia* (2-0%), Apiaceae (4-0%), *Ranunculus* (8-0%), *Stachys* (3-0%) and *Veronica* (3-0%) (Fig. 7). Poaceae increased up to 59% and then decreased to 15% by 6700 BP. Asteraceae remained abundant (22%) whilst *Acaena* (0-6%), *Plantago* (1-20%)

and *Valeriana* (9-27%) increased. Tree taxa such as *Alnus* (20-0%), *Hedyosmum* (5-0%) and *Podocarpus* (4-0%) decreased in the zone, whilst *Clethra* (0-4%), Ericaceae (1-3%), *Vallea* (0-3%) and *Weinmannia* (1-8%) increased. *Cyathea* (2%) was recorded for the first time at ca. 10,600 BP. This zone displayed the highest macroscopic charcoal concentration (123 particles/cm³) in the record ca. 10,990 BP (Fig. 8). Macroscopic charcoal peak frequency continued to increase across the zone (2.5-3.5 fires per 1000 yr). Microscopic charcoal and peak magnitude were highest in abundance reaching 190 particles/cm³ and 123 particles cm⁻² peak⁻¹ (ca. 10,990 BP). Between 9600-6600 BP, microscopic charcoal frequency increased then decreased from 3.5-1.4 fires per 1000 yr. The interval recorded two final key SCF regime shifts at 10,990 and 9018 BP reaching 0.41 and 1.1 RSI, respectively.

Zone M5 (2150-1150 BP)

Key SCF peaked at 4726 spores/cm³ (3%) ca. 2150 BP in zone M5 (Fig. 7). *Sporormiella* (278 spores/cm³, < 1%), *Arnium imitans* (3405 spores/cm³, 2%), *Coniochaeta* (487 spores/cm³, < 1%), *Delitshia* (347 spores/cm³, < 1%), *Sordaria* (695 spores/cm³, < 1%), and *Trichodelitschia* (278 spores/cm³, < 1%) were all present. Herbaceous taxa including Apiaceae (10%), Asteraceae (36%), Cyperaceae (6%), and Poaceae (34%) dominated the zone and were accompanied by Acanthaceae (< 3%), Scrophulariaceae (< 4%) and *Veronica* (< 2%) (Fig. 7). From the arboreal record, *Alnus* (3%) and *Hedyosmum* (8%) were the most representative. In this zone, *Rumex* appeared for the first time (1%) (Appendix A). Concentrations of both macroscopic (624-1308 particles/cm³) and microscopic (1484-3574 particles/cm³) charcoal were the highest in the record (Appendix B).

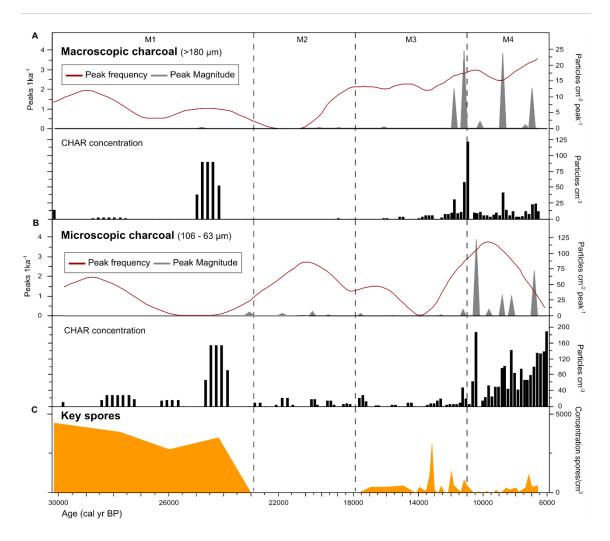


Figure 8. Monquentiva charcoal record and concentration of key spores of coprophilous fungi (key SCF). (A) Macroscopic charcoal; (B) microscopic charcoal; and (C) key SCF record. Charcoal concentration (black bars; particles/cm³), peak frequency (red line, left hand axis; peaks/ka), and peak magnitude (grey silhouettes, right hand axis; particles/cm²/peak) for charcoal data analysed in CharAnalysis using a LOWESS smoother model (Higuera et al., 2009). Vertical dashed lines show the four palynological zones (M1–M4) based on sedimentological changes in the core stratigraphy, key SCF regime shifts, and prominent changes in SCF and pollen assemblages as in Figure 7.

Fungal spore correlations

Sporormiella abundance in the Monquentiva record correlates positively with *A. imitans, Delitschia, Podospora, Sordaria, and Trichodelitschia (p-value < 0.01)* and with *Cercophora* and *Coniochaeta (p-value < 0.05)* (Table 2). This includes a high positive correlation between *Sporormiella* spores with *Sordaria (R*² = 0.633, p = 0.001), *A.imitans (R*² = 0.644, p = 0.001) and *Delitschia (R*² = 0.691, p = 0.001). Twenty other spores were recorded at Monquentiva and included 13 spores described relatively recently in the literature (Fig. 6) (Revelles *et al.,* 2016; Loughlin *et al.,* 2018). *Sporormiella* was positively correlated with three of these newly described spores OU-5 ($R^2 = 0.624$, p = 0.001), OU-100 ($R^2 = 0.770$, p = 0.001) and OU-108 ($R^2 = 0.549$, p = 0.001), but showed no significant correlation with OU-18, UAB-1, UAB-2, UAB-7, UAB-15, UAB-30A, UAB-32, UAB-40, UAB-48 and UAB-49 at the 0.01 level of significance (Appendix E). The significant correlations between *Sporormiella* and the other SCF were used to support the key SCF sum to represent megafaunal abundance changes over time (Fig. 7).

Monquentiva	Key SCF						
	Sporormiella	A.imitans	Cercophora	Coniochaeta	Delitschia	Podospora	Sordaria
A.imitans	.644**						
Cercophora	.312*	.225					
Coniochaeta	.672*	.714**	.530**				
Delitschia	.691**	.722**	.069	.435**			
Podospora	.468**	.176	.485**	.462**	.138		
Sordaria	.633**	.574**	.562**	.797**	.392**	.586**	
Trichodelitschia	.364**	.749**	067	.402**	.605**	133	.206

Table 2. Pearson correlations analysis amongst key spores of coprophilous fungi (Key SCF) from the Monquentiva record. Statistical significance of correlations are highlighted at p-value 0.01 (**) and 0.05 (*) levels.

Multivariate ordinations

The CCA analysis of the fossil pollen record grouped taxa along two axes which explained a cumulative 87% of the total variance in the dataset (Fig. 9). CCA Axis 1 explained 63% of the variance (eigenvalue: 0.0879) of pollen data, and CCA Axis 2 explained 24% of variance (eigenvalue: 0.0337). CCA Axis 1 ordinated taxa including *llex*, *Croton*, Gesneriaceae, and *Oxalis* at the positive end, and *Rumex*, *Alternanthera*, *Aragoa*, and Apiaceae at the negative end (Fig.

Page | 35

9a). The pollen taxa with the highest scores in Axis 2 were *Alternanthera*, *Rumex*, *Vicia* and *Dodonaea* at the positive extreme of the Axis, and *Carex*, *Escallonia*, Myrtaceae and Rosaceae were distributed at the negative end. The CCA ordination displayed a clear separation between samples from the Holocene at the positive extreme of Axis 2 and the Pleistocene at the negative end (Fig. 9b). Pollen from taxa found in the centre of the ordination such as Asteraceae, Poaceae, *Alnus, Hedyosmum* and *Podocapus* are more constant throughout the record thus are not strongly grouped to any of the environmental variables. The CCA also showed that the key SCF covaried with the negative end of Axis 2 and explained 74% of the variance along this axis. Macroscopic and microscopic charcoal covaried with the negative end of Axis 1 and both explained 98% of the variance along Axis 1.

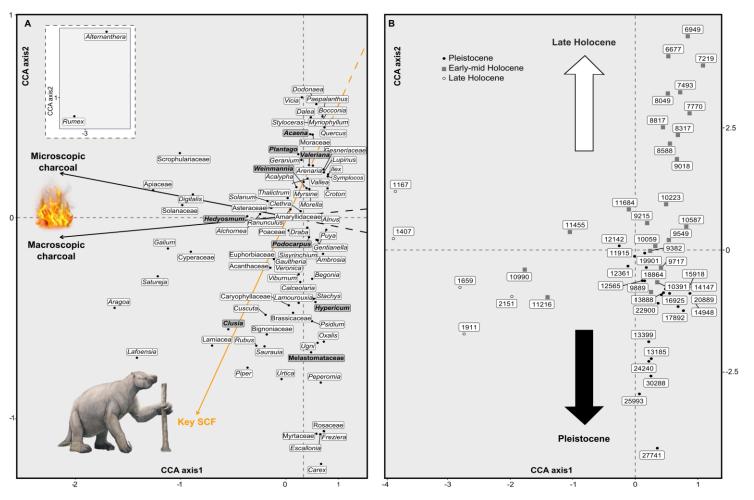


Figure 9. Canonical correspondence analysis (CCA) results for the pollen, charcoal, and spores of coprophilous fungi records from Monquentiva. (A) Species scores for the pollen record of taxa >1% the total pollen sum, using key SCF and macroscopic and microscopic charcoal as constraining variables. Bold taxa in grey boxes are discussed in detail in the text. The inset shows pollen species with high CCA Axis 1 and Axis 2 scores. The giant ground sloth (*Eremotherium*) concept art sourced from Ugueto (2020). (B) Sample scores labelled with their corresponding ages in calibrated years before present (BP).

5. DISCUSSION

The timing of megafaunal decline in the northern tropical Andes

We found robust evidence for megafaunal presence and its decline at Monguentiva (Table 2, Fig. 7). In our record, Sporormiella remained below the 2% functional megafaunal extinction threshold as proposed from an arid region of western North America (Davis and Shafer, 2006). However, that threshold may be inappropriate for other environments (Perrotti and van Asperen, 2019). Sporormiella percentages below 2% are not uncommon in the literature (e.g. Burney et al., 2003; Perrotti, 2018). The use of Sporormiella concentrations (Perrotti, 2018; Chepstow-Lusty et al., 2019; Conroy et al., 2020) instead of percentages, has been shown to be effective to infer megafaunal extinctions (Johnson et al., 2015; Raczka et al., 2018; Raczka et al., 2019). In lakes located in southeastern Brazilian, Sporormiella concentrations between 0 and 339 spores/cm³ have indicated low to medium contemporary cattle presence around the lakes (Raczka et al., 2016). We recovered Sporormiella at concentrations between 0 and 417 spores/cm³, falling in the upper-end of those ranges and suggesting Pleistocene megafaunal presence. We also found statistically significant positive correlations between Sporormiella abundance and the other key SCF (Table 2). This allowed us to create a multi-species key SCF group to estimate megafaunal presence and track megafaunal declines (Fig. 7) (Conroy et al., 2020). We combine the concentration of key SCF and statisticallysignificant regime shifts to identify megafaunal declines in Monguentiva.

The recovery of SCF can also be influenced by biotic and abiotic conditions related to fungal spore germination and growth. Dung fungi face interspecific competition (Kuthubutheen and Webster, 1986; Dix and Webster, 1995) and have genus- or species-specific responses to micro-environmental factors including temperature and limnological conditions (Dickinson and Underhay, 1977; Krug *et al.*, 2004). Our understanding of the role of environmental conditions on fungal spore germination is focussed on temperature zones (Lee *et al.*, 2022). However, fluctuating water levels have been shown to skew *Sporormiella* (Raper and Bush, 2009; Raczka *et al.*, 2016) and other spore records (Ponel *et al.*, 2011). This can increase or decrease the transportation and accumulation of spores in lakes (Raper and Bush, 2009;

Etienne *et al.*, 2013; Lee *et al.*, 2022). At Monquentiva, we used multi-species key SCF concentration declines to minimise the effects of environmental conditions on individual SCF germination and growth experienced by individual SCF. We also used our pollen record to infer environmental conditions such as water levels during periods of low spore concentrations to infer the presence and declines of megafauna.

The multi-species key SCF record indicates the presence of megafauna in Monquentiva and the eastern Colombian Cordillera since at least 30,290 BP (Fig. 7). Existing evidence of megafaunal presence in the tropical Andes has been derived from macrofossil remains and spore records. Megafauna macrofossil remains are scarce (Barnosky and Lindsey, 2010) but have been dated in Venezuela back to 47,990 BP (Jull et al., 2004), Ecuador back to 20,980 BP (Coltorti et al., 1998), Peru back to 10,150 BP (Marshall et al., 1984) and Brazil back to 20,060 BP (Neves and Piló, 2003). The limited spore records produced to date provide evidence for megafaunal presence in Ecuador back to 12,000 BP (Raczka et al., 2019), Peru back to 25,000 BP (Rozas-Dávila et al., 2016) and Brazil back to 23,000 BP (Raczka et al., 2018). In Colombia, macrofossil remains have been dated back to 40,000 BP (van der Hammen and Correal-Urrego, 1978) and although limited (Barnosky and Lindsey, 2010) have been found in Tocaima, an archaeological site at 340 masl (Correal-Urrego, 1993; Correal-Urrego, 2005; Alfonso-Rojas et al., 2021) and at Tibitó at 2500 masl (Correal-Urrego, 1981; Borrero, 2009). Megafaunal presence have also been inferred from rock shelter cave paintings from Serranía de la Lindosa (ca. 500 km from Monquentiva) that date back to ca. 12,600 BP (Bischler and Pinto, 1959; Correal et al., 1990). However, this evidence has been highly criticised (Baena et al., 2004). Our SCF record reveals that megafauna also roamed the higher altitudes of Colombia during the Pleistocene and were present at Monquentiva and the Colombian Andes dating back to at least 30,290 BP.

Our data indicated the first wave of megafaunal decline in Monquentiva at 22,900 BP, followed by a recovery ca. 16,900 BP and a second decline by ca. 10,990 BP (Fig. 7). Such SCF fluctuations could also result from differential preservation due to water level changes (Wood and Wilmshurst, 2013; Raczka *et al.*, 2019) and water-body size (Raper and Bush, 2009; Lee *et al.*, 2022). However, the pollen record shows that *Isoëtes* was low during the onset of both

Page | 39

megafaunal declines and did not coincide with changes in SCF (Fig. 7). *Isoëtes* is an aquatic taxon, common at the bottom of lakes, marshes and bogs in the high Páramo (van der Hammen and Gonzalez, 1960). *Isoëtes* did increase in the Monquentiva record suggesting the formation of a small water body, but ca. 600 years after the second SCF decline. Consistently low water levels and independence of the SCF and *Isoëtes* records at Monquentiva suggest the SCF concentration decreases are representative of megafaunal declines and are not due to water-level fluctuations.

The first megafaunal decline at ca. 22,900 BP was identified as a statistically significant regime shift in Monquentiva (Fig. 7). Three further key SCF regime shifts were identified at ca. 12,770, 12,140 and 10,990 BP. The shifts at 12,140 and 10,900 BP are linked to a decline in key SCF concentration from 1459 to 0 spores/cm³. We interpret these shifts as the local extinction of some Pleistocene megafauna at Monquentiva. This record is consistent with a twowave megafaunal decline from Lake Pacucha in the high Andes of Peru (Rozas-Dávila et al., 2016). However, at Pacucha the first megafaunal decline started at 21,000 BP and resulted in an extinction by 15,800 BP (Rozas-Dávila et al., 2016). In Ecuador the extinction of Pleistocene megafauna is recorded at 12,800 BP (Nascimento et al., 2020) while in Lagoa Santa, Brazil is reached by 12,000 BP and recorded no megafaunal recovery (Raczka et al., 2018). At Monquentiva and the Colombian Andes the second decline of megafauna (ca. 10,990 BP) was recorded later than other South American sites. However, macrofossil remains from species such as Doedicurus have been dated as late as 6560 BP in Argentina (Borrero et al., 1998). Ecosystems are complex and highly-diverse across South America and this heterogeneity could be linked to variability in megafaunal extinction timings (Gill et al., 2009; 2012; Rozas-Dávila et al., 2016). Thus, we suggest that continent-wide assessments could overwrite this variability and the complexity of factors that shaped these habitats after the Pleistocene megafaunal extinctions.

Climate, vegetation and human backdrop of megafaunal declines

Páramo vegetation dominated Monguentiva between 30,000 to 22,900 BP (Fig. 10). The dominance of Poaceae up to 24,240 BP, the low abundance of arboreal elements and low total pollen concentration at Monguentiva indicated a cool and dry climate. The tree line, defined as the upper altitudinal boundary of continuous forests (Urrego et al., 2011a), was lowered and open vegetation was found not just at Monquentiva but also across the high elevations in the eastern Colombian Andes (van Geel and Van der Hammen, 1973). This period corresponded with the coldest phase of the Last Glacial period (van der Hammen, 1974, Bogotá-A et al., 2011). High concentrations of key SCF during the Last Glacial period at Monquentiva indicate that the open vegetation structure was favourable for megafaunal presence. Between 25,000 and 24,200 BP, CHAR increased but the charcoal magnitude and frequency remained low suggesting a period of high-intensity but low-frequency fires (Fig. 8). These high-intensity fires resulted from relatively dry conditions indicated by the low abundance of aquatic and wetland taxa including *Isoëtes* and Cyperaceae (Fig. 7). However, fire activity was not sustained during the first wave of megafaunal decline ca. 22,900 BP. This could result from the cold and dry climate at the end of the Late Glacial (Bogotá-A et al., 2011; Urrego et al., 2011b) and a reduction in fuel-load by the decline of Poaceae from 24,240 BP (Fig. 7).

The first wave of megafaunal decline at 22,900 BP took place during a relative glacial temperature and humidity increase at Monquentiva as evidenced by displacements of Páramo by SubPáramo vegetation and increases in arboreal elements (Fig. 7 and 7). The increase of Andean forest taxa including *Alnus* and *Podocarpus* indicated a relative increase in temperatures and the absence of *lsoëtes* indicated low water levels. *Alnus* and *Podocarpus* are cold tolerant and generous wind-pollinated pollen producers (anemophilous), and their high abundance could imply over-representation from lower altitudes (van der Hammen and Gonzalez, 1960; Colinvaux *et al.*, 1999). However, we suggest the forest tree line moved upwards, as *Weinmannia* and *Podocarpus* also increased, two taxa indicative of a local signal of upper montane forest in the Tropical Andes (Gomez *et al.*, 2007; Hagemans *et al.*, 2019). This relative warming was short-lived and temperatures decreased again as indicated by a further rise in

Subpáramo taxa such as Asteraceae and a decline of arboreal elements at ca. 20,890 BP. This fluctuation of the upper forest line is consistent with changes from lower altitudes of Colombia (Bogotá-A *et al.*, 2011) suggesting temperature changes as megafaunal populations declined during the Late-Pleistocene. By 16,930 BP, environmental conditions once again provided a favourable habitat for megafauna at Monquentiva.

The second wave of megafauna decline occurred at ca. 10,990 BP during the El Abra stadial (Fig. 10) (Hooghiemstra and van der Hammen, 1993). This period coincided with an increase in herbaceous, arboreal and shrub taxa from the high Andean forest and Páramo vegetation, suggesting a novel structure and transitional vegetation (see ecological consequences discussion below). Nearby vegetation records from Lake Fuquene (2580 masl, Bogotá-A *et al.*, 2011) and Cienaga del Visitador (3300 masl, van der Hammen and González, 1965) also show rises in *Alnus* and a vegetation mosaic of Andean forest, Subpáramo and Páramo species. The decline of megafauna at Monquentiva during the cool El Abra stadial contrasts with literature suggesting that megafaunal extinctions tracked warm interstadials (Cooper *et al.*, 2015), but it does point to declines occurring during periods of climate instability.

The decline of megafauna at Monquentiva from 10,990 BP also coincided with the earliest evidence of hunter-gatherers in Colombia (Fig. 10). Hunter-gatherer groups are reported since the Guantiva interstadial ca. 12,500 BP in the Bogotá highlands (2600 masl) and downslope from Monquentiva at 340 masl since 11,740 BP (van der Hammen and Correal-Urrego, 1978; Correal-Urrego, 1993; Triana-Vega *et al.*, 2019; Alfonso-Rojas *et al.*, 2021). Megafaunal extinctions have been linked to overhunting by humans worldwide (Martin and Klein, 1984; MacPhee and Sues, 1999) and suggested as a major driver of extinction in North America and Australia (Martin and Klein, 1984; Stuart, 1999; Rule *et al.*, 2012). In South America, our understanding of the role of overhunting in the decline of Pleistocene megafauna is less clear due to the paucity of records. Records from coastal Ecuador suggest a stratigraphic correlation between megafaunal remains and stone tools of suspected Holocene age (Ficcarelli *et al.*, 1997). However, megafaunal hunting by humans is not thought to have been determinant in their extinction at this site (Ficcarelli *et al.*, 1997).

The Pleistocene collapse of megafauna in Brazil occurred during a window of human occupation (Raczka *et al.*, 2018), but in Peru, megafaunal populations collapsed before human presence (Rozas-Dávila *et al.*, 2016). At Monquentiva megafaunal populations were unable to recover after 10,990 BP suggesting that a synergy between humans and climate could have contributed to their decline and absence. Overall, our record allows us to infer the timing of megafaunal decline and ecological consequences but we have little evidence to explore the cause of extinctions.

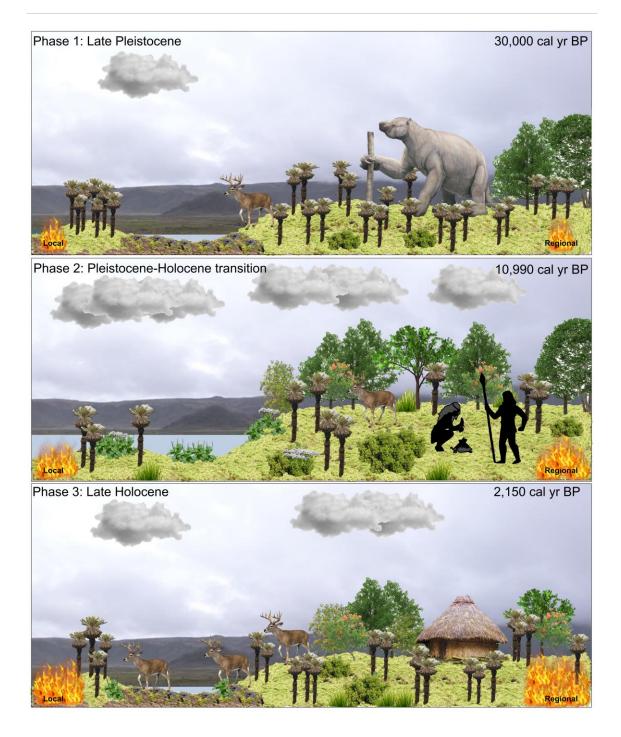


Figure 10. A conceptual landscape drawing of the changes in megafaunal presence, vegetation and fire activity reconstructed for Monquentiva during the Late Pleistocene and Holocene. Produced using Inkscape (Inkscape Project, 2022). Megafauna are depicted by the giant ground sloth (*Eremotherium*; concept art sourced from Ugueto, 2020) and a white-tailed deer representation (*Odocoileus virginianus*) (https://www.allwhitebackground.com/?p=4845; viewed 04 January 2022). The flora clipart not cited are sourced from Microsoft clipart 2016. Trees and shrubs include representations of *Alnus* and *Morella* (3d_moiler, 2014; Sharpj99, n.d.). Herbaceous taxa include *Espeletia* (Asteraceae), *Stachys* and *Valeriana* (Mergili, 2007; Vancouverislandgrows, 2018; Zwerver, n.d). Local and regional fire activity represented by relative size of flame. Silhouettes indicate hunter-gatherer groups (Vlasiuk, 2020). Kogi hut represents occupation by the pre-Hispanic indigenous group Muiscas (Benedek, 2015). Clouds and lake (blue polygon) represent humidity and lake level at the site (Garcia, 2022). Photographic backdrop of Monquentiva taken by J.W. Oughton in 2019.

Ecological consequences of megafaunal collapse

We found that some tree and shrub species became absent from the record during both waves of megafaunal decline, increasing in abundance during partial recovery of megafauna ca. 16,930-10,990 BP (Fig. 7). the Melastomataceae, Clusia and Hypericum all increase at Monquentiva when megafaunal abundance recovers. These pollen taxa can be dispersed by herbivores (Grabandt, 1980) suggesting a correlation that is corroborated by the CCA analysis (Fig. 9a). Across the contemporary tropics, pollination and dispersal of these families by megafauna have been recorded directly, as with the case with species of Melastomataceae, Clusiaceae, and Hypericaceae (Lemes-Martins et al., 2007; Bufalo et al., 2016; Escribano-Avila et al., 2018). Melastomataceae and Clusiaceae are also known to be dispersed by birds (Cronquist, 1988; Sánchez-Sánchez et al., 2015) however, if birds at Monquentiva dispersed species such as Clusia from the Clusiaceae family, we suggest this could be an indirect relation to the presence of megafauna attracting birds to the site (Galetti et al., 2018). Similarly, we found that several grass and herb taxa, including Acanthaceae, Apiaceae, Ranunculus and Veronica, became absent from the record during the second wave of megafaunal decline ca. 10,990 BP (Fig. 7). This suggests that the decline of these taxa are likely a result of reduced meditation in seed-dispersal by megafauna, rather than climate alone. Such relations between seed-dispersal services and the loss of megafauna have been found in the Neotropics today where the loss of primates has led to dispersal limitation in plant species (Peres et al., 2016). As Pleistocene megafauna declined in the high Andes of Peru, grasses and forbs were also observed to decline (Rozas-Dávila et al., 2016). In the Brazilian lowlands, vegetation remained largely unchanged with only a few taxa declining in abundance (Raczka et al., 2018). These contrasting records probably relate to differences in the original vegetation and suggest the ecological consequences of megafaunal declines were habitat-specific. In the Colombian Andes however, our palaeoecological record suggests an important role of the megafauna in the reproduction and dispersion of tree, grass and herb taxa.

The pollen record also showed palatable and woody taxa increasing at Monquentiva following the second decline of megafauna ca. 10,990 BP (Fig. 7).

Plantago is a highly palatable herb (Rozas-Dávila et al., 2016) that increased following megafaunal declines. The CCA analysis shows that *Plantago* plotted on the opposite extreme of key spores indicating its sensitivity to megafaunal presence (Fig. 9a). Plantago is indicative of cold Páramo vegetation (Hooghiemstra and Flantua, 2019), thus the species representation in the record under early-Holocene warming suggests their release from megafaunal grazing. The lagged increase of *Plantago* could reflect the late-successional trait of the taxon (Hooghiemstra and Flantua, 2019). At Monquentiva, Ericaceae increases after the second megafaunal decline (Fig. 7) and plots on the opposite ecological extreme of key SCF in the CCA (Fig. 9a). Some Ericaceae are woody shrubs in Andean sites (Bush et al., 2015; Rozas-Dávila et al., 2016) and this increase suggests woody species overgrowth following megafaunal declines. Such woody overgrowth is also recorded in other palaeoecological records including other Andean and North American records (Gill et al., 2009; 2012; Raczka et al., 2018, Rozas-Dávila et al., 2016). Valeriana declined after the second megafaunal decline but became more abundant in the Mid-Holocene when some megafaunal species recovered (Fig. 7). Valeriana, is also woody but commonly present in the SubPáramo-Páramo (Sylvester et al., 2018; Hooghiemstra and Flantua, 2019). The first decline of Valeriana could thus indicate Early-Holocene warming and the Mid-Holocene rise could suggest reduced woody suppression by the reduced megafaunal populations. This is supported by the CCA analysis where Valeriana plots on the opposite ecological extreme of key SCF (Fig. 9a). The lack of herbivory after the second megafaunal decline probably contributed to the encroachment of palatable and woody species at the higher altitudes of the Colombian eastern Cordillera.

At Monquentiva, a turnover in arboreal taxa and reduction of an open Poaceae-dominated grassland is recorded after the second wave of megafaunal decline ca. 10,990 BP (Fig. 10). The prominent Andean taxa were Ericaceae, *Hedyosmum, Podocarpus* and *Weinmannia,* surrounded by shrubby Subpáramo grasses and herbs including *Acaena*, Asteraceae, and *Valeriana*. The presence of *Cyathea,* a tree fern that typically inhabits gaps in closed forests suggests increases in forest cover (Crystal-Arens, 2001). This ecosystem transformation at Monquentiva suggests the replacement of open vegetation with a closed formation of non-analogue vegetation following the second decline of megafauna. This finding coincides with the consequences of defaunation reported for the Peruvian Andes (Rozas-Dávila *et al.*, 2016), other neotropical (Dantas and Pausas, 2022), and North American sites (Gill *et al.*, 2009; 2012). The flora turnover at Monquentiva occurred during the warmer Guantiva interstadial and cooler El Abra stadial 12,000-10,200 BP (Hooghiemstra and van der Hammen, 1993) thus climate oscillations could have played a role in vegetation changes observed. However, the CCA analysis showed that the abundance of some of these novel flora including *Acaena*, *Valeriana* and *Weinmannia* were ordinated on the opposite end of the constraining variable for megafauna (Fig. 9a). This suggests the transitional non-analogue vegetation was closely linked to megafaunal declines and their role in determining vegetation composition and structure at Monquentiva.

The charcoal record from Monguentiva represents a transition to higher local and regional fire activity during the early-Holocene megafaunal decline (Fig. 8). The reduction of herbivory during the early-Holocene probably led to fuel buildup of woody and grass biomass and increased fire activity. Megafaunal presence and fire activity on our CCA show an independent relationship (Fig. 9a), highlighting the link between rising fire activity and megafaunal absence. Andean megafauna likely acted as fuel-load suppressors in the tropical Andes as species including the white-tailed deer are browsers, thus trample on vegetation and feed on palatable flora (Martínez-Polanco, 2019). Our record is consistent with increased fire activity after megafaunal collapse reported in other systems after megafaunal declines (Burney et al., 2003; Gill et al., 2009; 2012; Rule et al., 2012; Rozas-Dávila et al., 2016), but it contrasts with the record from the Brazilian Cerrado where the collapse of megafaunal populations did not result in increased fire activity (Raczka et al., 2018). Thus, we suggest that the complexity and diversity of South American ecosystems lead to heterogeneity and habitat specificity regarding the expected relationship between Pleistocene megafaunal decline and increasing fire activity.

Extant Holocene megafauna, climate and human activity

Monquentiva witnessed a recovery in key SCF concentrations from ca. 8800 BP (Fig. 7 and Fig. 10) that could represent the presence of extant megafauna (Perrotti, 2018; Conroy et al., 2020). This probably included those that inhabit Colombian ecosystems today such as the Brocket and white-tailed deer (Cooke, 1998; Martínez-Polanco, 2019), and the Andean spectacled bear (García-Rangel, 2012). This is consistent with surveys of white-tailed deer bone remains dated back to 2320 ± 50 14C yr BP (Becerra, 1995). The recovery of megafauna under human occupation could relate to a change away from huntergatherer societies towards sedentism (Triana-Vega et al., 2019; Archila et al., 2021). In the Late-Holocene, Poaceae and Asteraceae were dominant suggesting an increase in open vegetation at Monguentiva. Increases in Hedyosmum, Weinmannia and Podocarpus indicated an expansion of forest on the surrounding slopes (Bogotá-A et al., 2011). Rumex appeared in the record at low but consistent percentages (Appendix A) and has been linked to degraded soils in Monquentiva (Espinoza et al., 2022). The dominance of Hedyosmum could also suggest human disturbance and be linked to disturbed areas (Gentry, 1993). These vegetation changes could reflect disturbance by pre-Hispanic Muisca groups (Langebaek, 1985; Langebaek, 2001). Increased humidity could have resulted in a shallow water body as suggested by the rise in Cyperaceae and low abundance of *Isoëtes* (Hooghiemstra and van der Hammen, 1993). The Late-Holocene vegetation cover and environmental conditions at Monquentiva likely provided habitat, water and food for extant megafauna, allowing some species to persist in this part of the Colombian Andes until today.

During the Late-Holocene, fire activity escalated at Monquentiva (Appendix B, Fig. 10). High charcoal concentrations likely reflected increased fires linked to agriculture (Gómez *et al.*, 2007) and other anthropogenic activities (Espinoza *et al.*, 2022). Thus, the escalated fire record under Late-Holocene megafaunal presence is likely to have been induced by human activities and facilitated by climate, which coupled to overrule the suppressive effect of megafauna on the vegetation.

6. CONCLUSIONS

This study is the first in the eastern Andes of Colombia, and probably across the continent, to use a multi-species SCF record to investigate the decline of Late-Pleistocene megafauna. We showed that the combined record of coprophilous and semi-coprophilous fungi provided more robust evidence to infer megafaunal presence and declines. The multi-species SCF record from Monquentiva indicated megafaunal presence in the eastern Colombian cordillera since at least 30,290 BP. At Monquentiva, megafaunal populations declined in two waves. The first decline was reached by 22,900 BP during a relative glacial temperature and humidity increase. By 10,990 BP the second megafaunal decline was reached, a period coeval with the EI Abra interstadial and the presence of hunter-gatherer groups. The extinction of some megafauna at Monquentiva also points to declines occurring during periods of climate instability. However, our data does not allow us to infer the causes of megafaunal extinctions or the potential role of overhunting.

One ecological consequence of the megafaunal declines at Monquentiva was the formation of novel vegetation assemblages. The representation of some plant species decreased whilst other palatable and woody taxa increased. This highlighted the role of megafauna in seed-dispersal and herbivory. Another ecological consequence was the increase in fire activity after megafauna declined. Pleistocene megafauna likely acted as fuel-load suppressors in the tropical Andes as they trampled on vegetation and fed on palatable flora.

Overall, we show that ecosystems in the high eastern Andes of Colombia were highly sensitive to the decline of megafaunal populations. Under the current biodiversity crisis, our understanding of the ecological consequences of megafaunal declines at Monquentiva suggest that current and future extinctions could have devastating consequences for biodiversity. This knowledge is fundamental for contemporary management and conservation efforts. To effectively manage biodiversity, these efforts must account for the effect of local herbivore declines on the dispersal of certain plant species, on fire activity and the potential loss of ecosystem services.

FUNDING

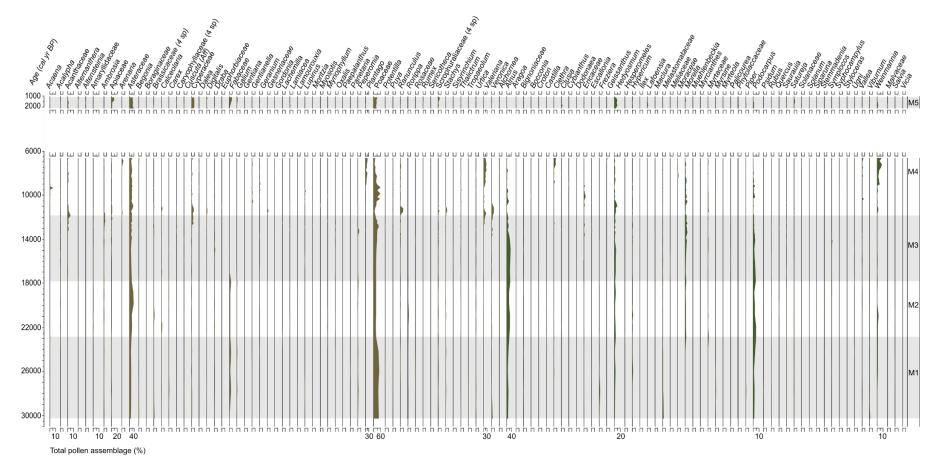
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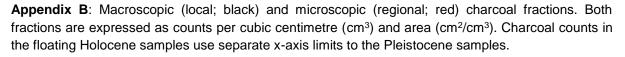
Page | 50

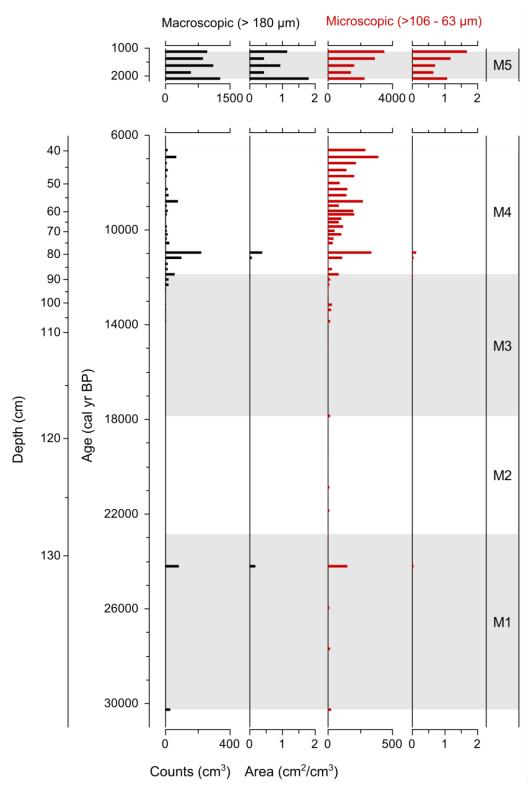
APPENDICES

Appendix A: Complete fossil pollen and charcoal diagram for all pollen taxa and charcoal particles recorded from the sediments of Monquentiva.

The pollen taxa are split into their ecological affinity, colour codes: Grasses and herbs (Brown-green); Trees and shrubs (Green). Malvaceae, *Salvia* and *Vicia* grouped at the end of the diagram as they can be found as trees, shrubs and herbs (Marchant *et al.*, 2002). Species are expressed as a percentage of the pollen sum (%TPA). Pollen types without visible value have an x-axis maximum of 5%. Pollen taxa unidentifiable at species level are grouped in their family and include the number of morphotypes.

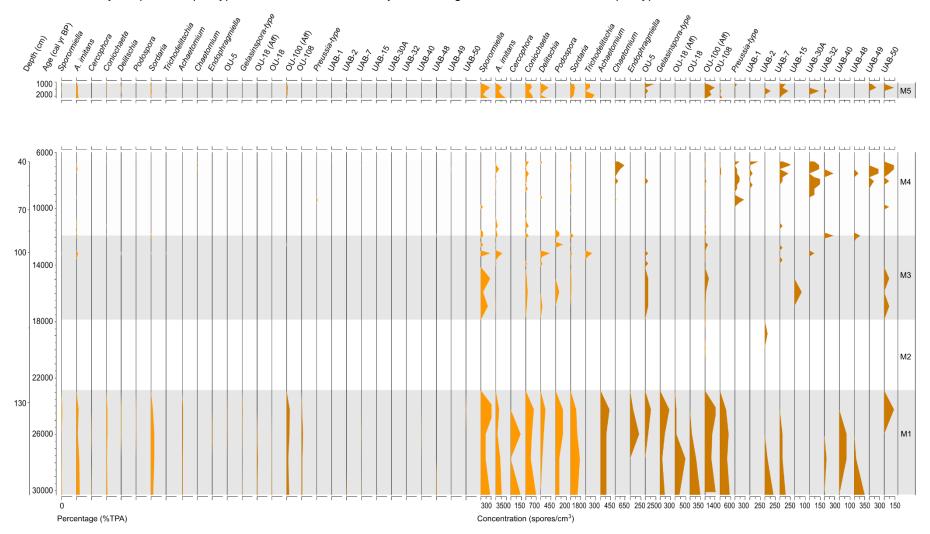






Appendix C: Complete fossil spore diagram for all identifiable fungi spore morphotypes recovered from the sediments of Monquentiva.

All coprophilous spores are expressed as a percentage of the total pollen assemblage (%TPA) and as concentration of counts per cubic centimetre (cm³). A 1x scale factor exaggeration is shown for all spores in the percentage quantification. Key spores (light orange) other identified fungi spores (dark orange). AFF refers to the affinity of spore morphotypes and indicates when they were assigned to their most akin morphotype.



Appendix D: Adapted outputs of a general outlier model run on the Monquentiva ¹⁴C radiocarbon ages, created using Oxcal v.4.4 (Ramsey, 2009a; Ramsey, 2009b).

The prior probability for each measurement being an outlier was set to 0.05. The posterior column provides the probability for each measurement being an outlier.

Core	Drive	LABID	Ok 📃	Outlier		Prior	Posterior	Model	Туре
MAR19II	D2	UBA-42145				5	5	General	t
		UBA-45480				5	5	General	t
		UBA-43590				5	5	General	t
		UBA-45479				5	5	General	t
		UBA-44500				5	5	General	t
		UBA-45478				5	100	General	t
		UBA-44499				5	99	General	t
		UBA-45477				5	100	General	t
	D1	UBA-44498				5	16	General	t
		UBA-44497				5	73	General	t
		UBA-43589				5	31	General	t
		UBA-42144				5	4	General	t
		UBA-44496				5	4	General	t
		UBA-45476			A	5	3	General	t
		UBA-44495				5	2	General	t
		UBA-45475				5	4	General	t

Colour codes: Accepted age (Blue); Likely outlier (Red).

Appendix E: Coefficient correlation outputs between all identifiable fungi spores recorded from the sediments of Monquentiva.

Correlations are calculated using Pearson correlation coefficients using the concentration of fungi spore counts per cubic centimetre (cm^3), at both the 0.01 (**) and 0.05 (*) level of significance.

	•		,	,.		()		()	0			
	Podosp Endoph UAB1	Spororn ora oragmiella UAB2	Sordaria Gelasin		<i>Trichod</i> OU-5	OU-18 t	<i>Achaet</i> o type	OU-100	<i>Chaetor</i> type	OU108	nia Preussi	a-type
Spororn			n Correla -0.163 0.021		1 .507** 0.002	.644** .624** 0.171	.312* .351* 0.249	.672** .770** 0.018	.691** .549** .412**	.468** -0.246	.633** -0.119	
	Sig. (2-1 0.249 0.883	tailed) 0.046 0.460	0.000 0.989	0.000 0.000 0.226	0.024 0.011 0.075	0.000 0.000 0.902	0.000 0.000 0.002	0.000 0.078	0.000 0.402	0.008 0.019	0.000 0.513	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
A.imitar	ns .749** .327*	Pearsor .334* 0.195	n Correla -0.020 -0.083	tion 0.069 0.020	.644** 0.152 0.035	1 .290* 0.168	0.225 .362** 0.251	.714** .562** 0.174	.722** .453** 0.181	0.176 -0.180	.574** -0.090	
	Sig. (2-1 0.888 0.560	tailed) 0.627 0.887	0.000 0.282 0.804	0.037 0.234	0.108 0.008 0.073	0.000 0.000 0.216	0.000 0.001 0.200	0.213 0.201	0.000 0.528	0.000 0.018	0.016 0.165	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Cercopi 0.067	hora .598** 0.234	Pearsor -0.059 -0.028	n Correla .659** -0.093	tion 0.094 -0.049	.312* .544** .480**	0.225 .523** .536**	1 .527** -0.064	.530** .781** -0.086	0.069 -0.095	.485** -0.043	.562** .525**	-
	Sig. (2-1 0.680 0.844	tailed) 0.000 0.510	0.024 0.510 0.730	0.108 0.000 0.000	0.000 0.000	0.000 0.000 0.654	0.626 0.000 0.546	0.000 0.504	0.000 0.763	0.637 0.000	0.000 0.095	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Coniocł	naeta .402** .459**	Pearsor .673** .410**	0.027 0.027	tion .363** -0.103	.672** .364** 0.167	.714** .648** .325*	.530** .529** .481**	1 .829** 0.121	.435** .733** 0.216	.462** -0.105	.797** 0.076	
	Sig. (2-1 0.758 0.852	tailed) 0.008 0.466	0.000 0.008 0.237	0.000 0.000 0.019	0.000 0.000 0.000	0.000 0.392	0.001 0.000 0.124	0.001 0.460	0.000 0.594	0.003 0.001	0.000 0.003	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Delitsch	nia .605** 0.032	Pearsor 0.246 0.061	Correla -0.098 -0.061	tion 0.236 0.033	.691** .374** -0.049	.722** 0.269 0.148	0.069 0.045 -0.101	.435** .423** 0.127	1 0.236 0.251	0.138 -0.114	.392** 0.007	
	Sig. (2-1 0.490 0.667	tailed) 0.092 0.818	0.000 0.006 0.728	0.000 0.054 0.297	0.626 0.750 0.475	0.001 0.002 0.370	0.092 0.072	0.330 0.421	0.004 0.960	0.000 0.823	0.079 0.666	

	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Podosp 0.133 0.003	oora .716** 0.208	Pearsor -0.117 -0.186	n Correla .600** 0.133	tion .281* .485**	.468** .706** .276*	0.176 .403** -0.127	.485** .593** 0.077	.462** .644**	0.138 -0.189	1 -0.085	.586** 0.175	-
	Sig. (2- 0.411 0.139	tailed) 0.000 0.186	0.000 0.044 0.347	0.213 0.000 0.000	0.000 0.003 0.047	0.001 0.000 0.370	0.330 0.000 0.585	0.181	0.000 0.548	0.347 0.214	0.000 0.982	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Sordari	a-type 0.206 .544**	Pearsor .809** .347*	n Correla -0.076 -0.023	tion .460** -0.028	.633** .337* 0.167	.574** .685** .670**	.562** .773** .446**	.797** .853** 0.047	.392** .840** 0.166	.586** -0.173	1 -0.074	
	Sig. (2- 0.591 0.870	tailed) 0.001 0.843	0.000 0.015 0.237	0.000 0.000 0.000	0.000 0.000 0.001	0.000 0.000 0.743	0.004 0.000 0.239	0.000 0.221	0.602	0.144 0.000	0.000 0.012	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Trichod	<i>lelitschia</i> -0.095 0.077	Pearsor -0.098 -0.047	n Correla -0.060 -0.061	tion 0.215 -0.082	.364** -0.086 -0.067	.749** -0.077 -0.091	-0.067 0.248 0.090	.402** -0.037 0.010	.605** -0.103	-0.133 -0.072	0.206 -0.087	1
	Sig. (2- 0.490 0.742	tailed) 0.675 0.669	0.008 0.127 0.564	0.000 0.543 0.637	0.637 0.588 0.520	0.003 0.077 0.524	0.000 0.794 0.946	0.347 0.465	0.144 0.614	0.542	0.504 0.586	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Achaete 0.095	o <i>mium</i> 1 0.136	Pearsor -0.083 -0.040	n Correla .605** -0.132	tion .324* 0.011	.596** .958** .598**	.334* .705** .369**	.598** .776** -0.090	.673** .919** 0.202	0.246 -0.134	.716** -0.061	.809** .411**	-
	Sig. (2- 0.559 0.780	tailed) 0.000 0.350	0.000 0.019 0.940	0.016 0.000 0.000	0.000 0.000 0.007	0.000 0.000 0.525	0.079 0.000 0.151	0.000 0.343	0.000 0.670	0.504 0.002	0.335	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Chaeto 0.098	<i>mium</i> -0.083 .536**	Pearsor 1 -0.041	n Correla -0.052 .591**	tion -0.117 0.058	-0.163 -0.075 -0.059	-0.020 -0.067 -0.020	-0.059 -0.113 .384**	-0.044 -0.048 .593**	-0.098 0.222	-0.117 0.249	-0.076 -0.076	-
	Sig. (2- 0.714 0.000	tailed) 0.408 0.683	0.249 0.595 0.680	0.888 0.636 0.890	0.680 0.427 0.005	0.758 0.734 0.000	0.490 0.114	0.411 0.076	0.591 0.594	0.490 0.000	0.559 0.773	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52

Endoph 0.060	nragmiella .605** 0.058	Pearsor -0.052 -0.025	n Correla 1 -0.083	tion .315* -0.044	.278* .617** .659**	0.069 0.137 -0.049	.659** .431** -0.057	.363** .595** 0.076	0.236 -0.084	.600** -0.038	.460** -0.046	-
	Sig. (2-1 0.714 0.861	tailed) 0.557	0.046 0.023 0.759	0.627 0.000 0.000	0.000 0.335 0.732	0.008 0.001 0.690	0.092 0.000 0.591	0.000 0.552	0.001 0.789	0.675 0.745	0.000 0.683	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Gelasir	0.215 0.025	Pearsor .324* 0.168	Correla -0.117 0.187	tion .315* -0.044	.507** 1 -0.056	0.152 .394** 0.094	0.094 -0.016 -0.109	.364** .351* -0.017	.374** 0.205 0.172	.281* -0.066	.337* -0.086	
	Sig. (2-1 0.408 0.184	tailed) 0.023 0.757	0.000 0.695	0.282 0.004 0.510	0.510 0.910 0.440	0.008 0.011 0.905	0.006 0.145 0.224	0.044 0.640	0.015 0.546	0.127 0.862	0.019 0.234	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
OU-5	Pearsor .958** 0.092	n Correlat -0.075 -0.036	tion .617** -0.120	.624** .394** -0.027	.290* 1 .395**	.544** .495** .302*	.648** .761** -0.082	0.269 .820** .282*	.706** -0.122	.685** -0.055	-0.086 .319*	
	Sig. (2-1 0.595 0.799	tailed) 0.000 0.395	0.000 0.004 0.851	0.037 0.004	0.000 0.000 0.030	0.000 0.000 0.563	0.054 0.000 0.043	0.000 0.389	0.000 0.698	0.543 0.021	0.000 0.517	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
OU-18 0.077	52 52	52 52	52	52 52								
	52 52 type .705**	52 52 Pearsor -0.067 -0.032	52 52 Correla 0.137	52 52 tion -0.016	52 .351* .495**	52 .362** 1	52 .523** .552**	52 .529** .808**	52 0.045	52 .403**	52 .773**	
	52 52 type .705** 0.226 Sig. (2-1 0.636	52 52 Pearsor -0.067 -0.032 tailed) 0.335	52 52 Correla 0.137 -0.107 0.011 0.910	52 52 tion -0.016 0.083 0.008 0.000	52 .351* .495** .637** 0.000	52 .362** 1 .584** 0.000 0.000	52 .523** .552** -0.073 0.750 0.000	52 .529** .808** -0.061 0.003	52 0.045 -0.109 0.000	52 .403** -0.049 0.588	52 .773** .656** 0.000	
	52 52 type .705** 0.226 Sig. (2-1 0.636 0.821 N 52 52	52 52 Pearsor -0.067 -0.032 tailed) 0.335 0.448 52 52 52 52	52 52 Correlar 0.137 -0.107 0.011 0.910 0.558 52 52	52 52 tion -0.016 0.083 0.008 0.000 0.000 52 52 52 52	52 .351* .495** .637** 0.000 0.000 52	52 .362** 1 .584** 0.000 0.000 0.606 52	52 .523** .552** -0.073 0.750 0.000 0.668 52	52 .529** .808** -0.061 0.003 0.442 52	52 0.045 -0.109 0.000 0.729 52	52 .403** -0.049 0.588 0.000 52	52 .773** .656** 0.000 0.107 52	52 - 52
0.077	52 52 type .705** 0.226 Sig. (2-1 0.636 0.821 N 52 52 52) type 0.248	52 52 Pearsor -0.067 -0.032 tailed) 0.335 0.448 52 52 52 Pearsor .776** .299*	52 52 Correlat 0.137 -0.107 0.011 0.910 0.558 52 52 52 52 52 52 52	52 52 tion -0.016 0.083 0.008 0.000 0.000 52 52 52 52 52 tion .431**	52 .351* .495** .637** 0.000 0.000 52 52 52 .770** .351*	52 .362** 1 .584** 0.000 0.000 0.606 52 52 52 .562** .761**	52 .523** .552** -0.073 0.750 0.000 0.668 52 52 52 .527** .552**	52 .529** .808** -0.061 0.003 0.442 52 52 52 .829** 1	52 0.045 -0.109 0.000 0.729 52 52 52 .423** .737**	52 .403** -0.049 0.588 0.000 52 52 52 .593**	52 .773** .656** 0.000 0.107 52 52 52 .853**	52 - 52
0.077	52 52 type .705** 0.226 Sig. (2-1 0.636 0.821 N 52 52 0 type 0.248 .476** Sig. (2-1 0.427	52 52 Pearsor -0.067 -0.032 tailed) 0.335 0.448 52 52 52 52 Pearsor .776** .299* tailed) 0.001	52 52 0.137 -0.107 0.011 0.910 0.558 52 52 52 52 52 52 52 52 52 52 52 52 52	52 52 tion -0.016 0.083 0.008 0.000 0.000 52 52 52 52 52 tion .431** -0.102 0.000 0.000	52 .351* .495** .637** 0.000 0.000 52 52 .770** .351* -0.023 0.000 0.000	52 .362** 1 .584** 0.000 0.000 0.606 52 52 .562** .761** .373** 0.000	52 .523** .552** -0.073 0.750 0.000 0.668 52 52 .527** .552** .348* 0.002 0.000	52 .529** .808** -0.061 0.003 0.442 52 52 .829** 1 0.086 0.000	52 0.045 -0.109 0.000 0.729 52 52 52 .423** .737** .345* 0.000	52 .403** -0.049 0.588 0.000 52 52 52 .593** -0.200 0.077	52 .773** .656** 0.000 0.107 52 52 .853** -0.094 0.000	52 - 52

	Sig. (2- 0.734 0.740	tailed) 0.000 0.501	0.000 0.145 0.623	0.001 0.000 0.000	0.000 0.000 0.000	0.000 0.000 0.937	0.092 0.491	0.000 0.348	0.000 0.736	0.794 0.000	0.000 0.070	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
Preussi 0.103	<i>ia</i> -type -0.134 0.047	Pearso 0.222 -0.066	n Correla -0.084 .286*	tion -0.066 0.005	-0.246 -0.122 -0.095	-0.180 -0.109 -0.073	-0.095 -0.200 0.215	-0.105 -0.133 0.000	-0.114 1	-0.189 .320*	-0.173 -0.123	-
	Sig. (2- 0.114 0.640	tailed) 0.552 0.040	0.078 0.640 0.974	0.201 0.389 0.504	0.504 0.442 0.606	0.460 0.156 0.126	0.421 0.348 1.000	0.181	0.221 0.021	0.465 0.387	0.343 0.740	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB1 0.061 0.030	Pearso 0.249 0.206	n Correla -0.038 0.165	tion -0.086 -0.043	-0.119 -0.055 0.043	-0.090 -0.049 0.168	-0.043 -0.094 -0.018	0.076 -0.048	0.007 .320*	-0.085 1	-0.074 -0.055	-0.072 0.178	-
	Sig. (2- 0.076 0.833	tailed) 0.789 0.143	0.402 0.546 0.241	0.528 0.698 0.763	0.763 0.729 0.763	0.594 0.508 0.234	0.960 0.736 0.898	0.548 0.021	0.602	0.614 0.697	0.670 0.207	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB2	Pearso .411** .407**	n Correla -0.076 -0.036	tion -0.046 0.091	.325* 0.025 0.078	.327* .319* 0.141	.525** .656** .629**	.459** .476** -0.082	0.032 .582** -0.111	0.175 -0.123	.544** -0.055	-0.087 1	
	Sig. (2- 0.594 0.799	tailed) 0.745 0.523	0.019 0.862 0.583	0.018 0.021 0.320	0.000 0.000 0.000	0.001 0.000 0.561	0.823 0.000 0.434	0.214 0.387	0.000 0.697	0.542	0.002 0.003	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB7	Pearso 0.136 -0.073	n Correla .536** .346*	tion 0.058 .314*	0.093 0.168 0.121	0.195 0.092 .294*	0.234 0.226 .289*	.410** .299* .297*	0.061 0.254	-0.003 0.047	.347* 0.178	0.077 .407**	1
	Sig. (2- 0.000 0.608	tailed) 0.683 0.012	0.513 0.234 0.023	0.165 0.517 0.391	0.095 0.107 0.035	0.003 0.031 0.038	0.666 0.070 0.033	0.982 0.740	0.012 0.207	0.586 0.003	0.335	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB15 0.040	Pearso -0.041 -0.065	n Correla -0.025 -0.034	tion 0.187 -0.028	0.021 -0.036 -0.038	-0.083 -0.032 -0.045	-0.028 -0.028 -0.060	0.027 -0.047	-0.061 -0.066	0.208 -0.030	-0.023 -0.036	-0.047 -0.073	- 1
	Sig. (2- 0.773 0.645	tailed) 0.861 0.809	0.883 0.184 0.844	0.560 0.799 0.788	0.844 0.821 0.754	0.852 0.845 0.673	0.667 0.740	0.139 0.640	0.870 0.833	0.742 0.799	0.780 0.608	

	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB30/ 0.132 0.065	A Pearson .591** 1	n Correla -0.083 -0.044	tion -0.044 -0.093	-0.105 -0.120 -0.127	0.020 -0.107 .337*	-0.093 -0.102 .318*	-0.103 -0.095	0.033 .286*	-0.186 0.206	-0.028 0.091	-0.061 .346*	-
	Sig. (2- 0.000 0.645	tailed) 0.557	0.460 0.757 0.754	0.887 0.395 0.510	0.510 0.448 0.368	0.466 0.472 0.015	0.818 0.501 0.022	0.186 0.040	0.843 0.143	0.669 0.523	0.350 0.012	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB32	Pearson 0.011 .314*	n Correla 0.058 -0.034	tion -0.044 -0.044	0.002 -0.056 1	0.035 -0.027 0.078	-0.049 0.083 .541**	0.167 -0.023 .354**	-0.049 0.070 0.063	0.133 0.005	0.167 0.165	-0.082 0.078	
	Sig. (2- 0.683 0.809	tailed) 0.759 0.754	0.989 0.695	0.804 0.851 0.581	0.730 0.558 0.000	0.237 0.870 0.010	0.728 0.623 0.658	0.347 0.974	0.237 0.241	0.564 0.583	0.940 0.023	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB40	Pearson .598** 0.121	n Correla -0.059 -0.028	tion .659** -0.093	0.171 0.094 0.078	0.168 .395** 1	.480** .637** 0.064	.325* .373** -0.064	0.148 .668** -0.086	.485** -0.095	.670** -0.043	-0.067 0.141	
	Sig. (2- 0.680 0.844	tailed) 0.000 0.510	0.226 0.510 0.581	0.234 0.004	0.000 0.000 0.654	0.019 0.007 0.654	0.297 0.000 0.546	0.000 0.504	0.000 0.763	0.637 0.320	0.000 0.391	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB48	Pearson .369** .294*	n Correla -0.020 -0.038	tion -0.049 -0.127	0.249 -0.109 .541**	0.251 .302* 0.064	.536** .584** 1	.481** .348* 0.114	-0.101 .525** -0.039	.276* -0.073	.446** 0.043	-0.091 .629**	
	Sig. (2- 0.890 0.788	tailed) 0.732 0.368	0.075 0.440 0.000	0.073 0.030 0.654	0.000 0.000	0.000 0.011 0.422	0.475 0.000 0.784	0.047 0.606	0.001 0.763	0.520 0.000	0.007 0.035	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52
UAB49 0.090 0.045	Pearson .384** .337*	n Correla -0.057 .354**	tion -0.017 -0.064	0.018 -0.082 0.114	0.174 -0.073 1	-0.064 0.086 .590**	0.121 -0.011	0.127 0.215	-0.127 0.168	0.047 -0.082	0.090 .289*	-
	Sig. (2- 0.005 0.754	tailed) 0.690 0.015	0.902 0.905 0.010	0.216 0.563 0.654	0.654 0.606 0.422	0.392 0.543	0.370 0.937 0.000	0.370 0.126	0.743 0.234	0.524 0.561	0.525 0.038	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52

UAB50	Pearson Correlation 0.202 .593** 0.076 .297* -0.060 .318*		.412** 0.172 0.063	0.181 .282* -0.086	-0.086 -0.061 -0.039	0.216 .345* .590**	0.251 0.098 1	0.077 0.000	0.166 -0.018	0.010 -0.111		
	Sig. (2- 0.000 0.673	tailed) 0.591 0.022	0.002 0.224 0.658	0.200 0.043 0.546	0.546 0.668 0.784	0.124 0.012 0.000	0.072 0.491	0.585 1.000	0.239 0.898	0.946 0.434	0.151 0.033	
	N 52 52	52 52 52	52 52 52	52 52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52	52 52

**. Correlation is significant at the 0.01 level (2-tailed) *. Correlation is significant at the 0.05 level (2-tailed).

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