# 1 LOCAL ADAPTATION OF REPRODUCTIVE PERFORMANCE DURING

# 2 THERMAL STRESS

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#### 25 Abstract

Considerable evidence exists for local adaptation of critical thermal limits in 26 ectotherms following adult temperature stress, but fewer studies have tested for local 27 adaptation of sub-lethal heat stress effects across life history stages. In organisms 28 with complex life cycles, such as holometablous insects, heat stress during juvenile 29 30 stages may severely impact gametogenesis, having downstream consequences on reproductive performance that may be mediated by local adaptation, although this is 31 rarely studied. Here, we tested how exposure to either benign or heat stress 32 temperature during juvenile and adult stages, either independently or combined, 33 influences egg-to-adult viability, adult sperm motility and fertility in high and low 34 latitude populations of Drosophila subobscura. We found both population- and 35 temperature-specific effects on survival and sperm motility; juvenile heat stress 36 decreased survival and subsequent sperm motility and each trait was lower in the 37 northern population. We found an interaction between population and temperature 38 on fertility following application of juvenile heat stress; while fertility was negatively 39 impacted in both populations, the southern population was less affected. When the 40 adult stage was also subject to heat stress, the southern population exhibited 41 positive carry-over effects whereas the northern population's fertility remained low. 42 Thus, the northern population is more susceptible to sub-lethal reproductive 43 consequences following exposure to juvenile heat stress. This may be common in 44 other organisms with complex life cycles and current models predicting population 45 responses to climate change, which do not take into account the impact of juvenile 46 heat stress on reproductive performance, may be too conservative. 47 Keywords: spermatogenesis, climate change, range margin, phenotypic plasticity, 48

49 developmental acclimation, carry-over effects, evolution, inversions

## 50 Introduction

While a variety of abiotic factors are changing due to anthropogenic impacts on the 51 climate, perhaps the most critical are increases in the mean and variability of 52 temperature (IPCC, 2014). This is important because temperature has unique and 53 profound effects on organismal biochemistry and physiology. Selection acts on the 54 ability of organisms to function optimally within the range of temperatures that they 55 routinely encounter and to minimize fitness costs during exposure to suboptimal 56 temperatures (Angilleta, 2009). Ectotherms may be especially vulnerable to thermal 57 stress given that their basic physiology is limited by ambient temperatures 58 (Hochachka & Somero, 2002). Ectotherms also represent the vast majority of 59 terrestrial biodiversity (Wilson, 1992) and so understanding how they will respond to 60 61 increasing temperatures is of critical importance (Deutsch et al., 2008).

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For wide-ranging species that experience different thermal gradients in different 63 environments, populations may exhibit variation in thermal adaptations and 64 responses to suboptimal temperatures (e.g., Gardiner et al., 2010; Yampolsky et al., 65 2014). This may lead to local adaptation which predicts that populations inhabiting 66 warmer climes will function better at higher temperatures relative to populations from 67 cooler locations and vice versa (Angilletta et al., 2003). Many studies have examined 68 69 responses to temperature stress in terrestrial ectotherms and quantified local adaptation. In particular, critical thermal limits (CTL) - the temperatures at which the 70 ability of an organism to remain active under extreme conditions breaks down - have 71 received much attention. CTLs are thought to provide insight into species 72 distributions, ecology and evolution, with upper thermal limits positively related to 73 optimal performance temperatures (Terblanche et al., 2007). However, recent 74

reviews have emphasised that studies of local thermal adaptation have not focused 75 on the most important life history traits, such as maintenance of fertility (Hoffmann, 76 2010). In previous studies, most of the traits measured to determine physiological 77 failure are related to lethal effects (e.g. survival) or only indirectly related to fitness 78 because they are general measures of mobility such as thermal knock-down. 79 However, the latter may not be correlated with subsequent reproductive success. 80 81 Moreover, reproductive processes, such as gamete production, are likely to be negatively affected at less extreme temperatures than loss of mobility and 82 83 organismal death (Jørgensen et al., 2006). The consequences of sub-lethal effects on reproductive performance may be exceptionally important in short lived 84 organisms (i.e. many insects) because reversing thermal damage to reproductive 85 processes may require a large proportion of total lifespan (Sinclair & Roberts, 2005). 86 Thus, examining the consequences of thermal stress on reproductive performance is 87 important in understanding how organisms will respond to climate change. 88

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Not only should reproductive fitness traits be a focus, but different life history stages 90 may experience different selection on these reproductive traits. Holometabolous 91 insects typically experience radically different juvenile and adult thermal niches and 92 therefore are expected to exhibit different susceptibilities to thermal stress (Angilleta, 93 94 2009; Hoffmann, 2010; Kingsolver et al., 2011). Vagile stages will have the opportunity to take advantage of environmental heterogeneity via behavioural 95 flexibility whereas developmental stages that are physically restricted have little or no 96 97 ability to move to less stressful environments.

The effects of thermal stress on reproductive performance may be particularly 99 relevant for males. For example, in *Drosophila*, spermatogenesis is predicted to be 100 more sensitive to thermal stress than oogenesis (David et al., 2005). Given that 101 sperm production begins during the non-vagile juvenile stage in many insects 102 (Nijhout, 1998), such that mature sperm have already left the testes at the time of 103 emergence from the pupa (Wigglesworth, 1965), thermal stress during this time may 104 represent a profound episode of selection subject to local adaptation. Relatively few 105 studies in holometabolous insects have examined how thermal stress during earlier 106 107 life cycle stages could affect subsequent organism physiology (Kingsolver et al., 2011). While recent work has begun to examine this, studies tend to measure egg-108 to-adult viability (Kristensen et al., 2015; Rohde et al., 2016) but only rarely the 109 110 survivors' subsequent reproductive performance (Chirault et al., 2015; Zhang et al., 2015a; b) and even more rarely in the context of local adaptation (Fragata et al., 111 2016). Some studies assess physiological responses following relatively short heat 112 shock (Chirault et al., 2015; Zhang et al., 2015a; b) but others ask what the fitness 113 consequences are after longer term exposure to mildly stressful temperatures during 114 development and whether this changes after laboratory adaptation (Fragata et al., 115 2016). 116

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Understanding the extent to which exposure to sub-lethal thermal extremes during
early life-history stages subsequently impacts survival and reproduction later in life is
critical to understanding the evolutionary potential and putative constraints on
thermal adaptation (Bowler & Terblanche, 2008). Positive carry-over effects (also
known as developmental acclimation) are one way that the non-vagile stages could
mitigate the cost of thermal stress in subsequent life-cycle stages, and may

represent an important component of thermal tolerance adaptation (Sgrò et al., 124 2016). Such effects occur when exposure to thermal stress at an earlier life cycle 125 stage results in increased resistance/tolerance to extreme temperatures at later 126 stages (Angilleta, 2009) and may vary across geographic gradients, suggesting local 127 adaptation (de Jong et al., 2010; Scharf et al., 2010). In contrast, negative carry-over 128 effects occur when stress during development negatively impacts fitness 129 components at the adult stage (Bacigalupe et al., 2007; Gaston & Spicer, 2013; 130 Schiffer et al., 2013; O'Connor et al., 2014). Two knowledge gaps exists in this 131 132 literature, however. Most studies examining carry-over effects have ignored reproductive traits, which may be more sensitive to thermal stress. Moreover, when 133 performed, these studies have examined response to cold stress (Sgrò et al., 2016). 134 135 However, capacity to deal with thermal changes is likely to be reduced as upper critical limits are reached. This is because the range of variation in resistance to 136 thermal stress is considerably less at high temperatures than at cold, both between 137 species (Gaston & Chown, 1999; Addo-Bediako et al., 2000) and within species 138 (Gaston & Chown, 1999; Terblanche et al., 2007). 139

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In this study, we test for local adaptation of survival and reproductive traits in a 141 southern and a northern population of the holometabolous ectotherm, Drosophila 142 143 subobscura, manipulating heat stress during either juvenile or both juvenile and adult stages. We focus on this species as it is a model for research on local adaptation to 144 increasing temperatures (Balanyá et al., 2006). We predict that the southern 145 population will have greater reproductive fitness after continuous application of heat 146 stress during the juvenile stage whereas the northern population will have either 147 greater or similar fitness to the southern population at permissive temperatures. 148

Moreover, because traits related to reproduction should exhibit thermal impacts before more frequently measured traits, such as survival, we predict the signal of local adaptation will be stronger for reproductive traits compared to survival following heat stress. Subsequent application of adult heat stress is predicted to affect reproductive performance differently between the populations with the southern population exhibiting positive, and the northern population exhibiting negative, carryover effects.

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## 157 Material and Methods

158 Fly stocks

Stocks of D. subobscura originated from wild-caught individuals collected in August-159 160 September 2011 from Uppsala, Sweden (59°85'N, 17°63'E) and from Valencia, Spain (39°51'N, 0°42'W), in the same season, with outbred laboratory populations 161 established in mesh cages (50cm<sup>3</sup>) at 18°C under a 12h light/dark cycle. 162 Subsequently, isofemale lines were established in August 2012 from these two 163 populations (representing ca. 8 overlapping generations) from a single female per 164 line and kept at a population size of greater than 50 at 18°C. The 30 year historical 165 average maximum temperature of the hottest month at each site is 23°C in Uppsala 166 and 31°C in Valencia and previous work across the distribution of this species has 167 168 suggested that 18°C is a non-thermally stressful temperature for all populations (Santos, 2007; Castañeda et al., 2013). Previous work found that males become 169 sterile at 25°C or even lower (Krimbas, 1993) and our preliminary experiments 170 suggested that this temperature elicited substantial egg-to-adult mortality. 171

A maximum of twelve isofemale lines from the Valencia population and four 173 isofemale lines from the Uppsala population were used in the experiments, 3-7 174 months after they were established (representing 3-6 generations), as Valencia 175 exhibits a more diverse set of chromosomal inversion polymorphisms (Krimbas, 176 1993) and we wanted to capture as much genetic variation as possible. While 177 isofemale lines may produce inbreeding effects, levels of fertility and sperm motility 178 under benign conditions are relatively high (see Results) and our statistical models 179 specified overdispersion to account for isofemale line variation. Flies were kept in 180 181 vials containing a maize, sugar and agar food mix with additional live dry yeast for all experiments, and kept under a 12h light/dark cycle. In all experiments, light CO<sub>2</sub> 182 anaesthesia was used to collect and sex flies. Flies were collected as virgins, stored 183 for 6 days to become reproductively mature (Holman et al., 2008) and then used in 184 experiments. 185

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## 187 Egg-to-adult viability

Male and female pairs from each isofemale line were put in separate vials and kept 188 at 18 °C. Vials were inspected each day, and when more than 30 eggs had been laid 189 (ca. 2 days), the pair was removed, the final number of eggs counted, and vials 190 maintained at either 18°C or 23.5°C. We hereafter refer to the 18°C treatment as 191 192 benign (B) and the 23.5°C treatment as stressful (H). We aimed for a minimum of ten vials per isofemale line per population per temperature treatment (total sample sizes: 193 B, Upp = 37 vials; H, Upp = 67; B, Val = 102; H, Val = 192). The vials were kept at 194 195 one of these two constant temperatures through development from egg to imago, and the numbers of eggs that emerged as adults were counted. 196

All analyses were performed using R (v. 3.2.2, the R Foundation for Statistical 198 Computing, 2016). For each trait, the maximal model was simplified by step-wise 199 removal of non-significant factors, as judged by change in Akaike Information 200 Criterion. For ease of comparison among traits, data in figures are presented relative 201 to the mean performance of the Valencia population at the benign temperature, after 202 back transformation from the logit scale. For viability we used a generalized linear 203 models (GLM) of the proportion of survivors, with quasibinomial error structure to 204 account for over-dispersion and a logit link function. Temperature [B, H], population 205 206 [Val, Upp]) and their interaction, to detect local adaptation (Kawecki & Ebert, 2004), were included in the analysis. The number of eggs was log transformed and treated 207 as a covariate in order to account for density effects. 208

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## 210 Sperm motility

Spermatogenesis is predicted to be more strongly affected by thermal stress than 211 oogenesis. Male sterility is frequently quantified in *Drosophila* as lack of motile sperm 212 (Coyne & Orr, 1989) and here we examined the seminal vesicles for the presence of 213 motile sperm following either benign or stressful developmental temperatures. Flies 214 were generated as described above. Male flies were collected on the day of eclosion 215 and kept in vials of less than 20 flies for six days at their developmental temperature 216 (B or H) to reach sexual maturity. Seminal vesicles of a minimum of ten males per 217 line per population per temperature treatment were dissected (Snook, 1998), gently 218 squashed on a slide under a cover slip in phosphate buffered saline, and sperm 219 motility assessed (total sample sizes: B, Upp = 46; H, Upp = 73; B, Val = 126; H, Val 220 = 223). Males were considered sterile (coded as 0) if no sperm, or very few motile 221 sperm, were present; males were considered fertile (coded as 1) if motile sperm 222

were present. This response was analysed using a quasibinomial GLM as forviability (but with no covariate).

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226 Fertility

An ultimate measure of fertility is the ability to produce progeny. Here we were 227 interested to measure the consequences of the thermal environment at both 228 developmental and adult stages on fertility. We had four treatments for each 229 population: both juvenile and adult temperatures were benign (BB); both were 230 231 stressful (HH); juvenile temperature was stressful but adult temperature was benign (HB); or juvenile temperature was benign and adult temperature stressful (BH). Flies 232 were generated as described above, being reared throughout development at either 233 234 the benign or stressful temperature and upon eclosion, virgin males and females were collected and separated by sex into vials of less than 20 individuals for six days 235 to reach sexual maturity and were randomly assigned to either remain as adults at 236 their developmental temperature or to switch to the alternative temperature. At 6 237 days post-ecolsion, a male and female were placed in a vial and monitored for larvae 238 production (we aimed for a minimum of 10 pairs per line per population per 239 temperature treatment; total sample size: for Uppsala, BB = 14, HH = 19, HB = 22, 240 BH = 21; for Valencia, BB = 50, HH = 48, HB = 89, BH = 73). Pairs were transferred 241 242 from the first vial after five days and placed in another vial for a further five days. Pairs were considered fully sterile if no larvae were produced in either vial (coded as 243 0); pairs were considered fully fertile if larvae were produced in both vials (coded as 244 2); and pairs were considered partially fertile if larvae were produced in only one of 245 the two vials (coded as 1). This response was analysed using a binomial GLM (0, 1 246 or 2 successes out of 2 trials) because no over-dispersion was detected. The 247

temperature treatment in this case had four levels (BB, BH, HB, HH). We tested for
the predicted carry-over effects by making the appropriate contrast: the effect of HH
relative to HB was expected to depend on population (HH>HB in Val, HB>HH in
Upp).

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## 253 **Results**

Temperature and population, but not their interaction, significantly influenced viability from egg to adult (Fig. 1a) and sperm motility (Fig. 1b; Table 1; interaction term, viability: df = 1, deviance = 0.065, p = 0.98; interaction term, sperm motility: df = 1, deviance = 1.042, p = 0.31). Higher temperatures resulted in decreased survivorship and sperm motility compared to lower temperature. Uppsala had lower viability and sperm motility than Valencia at both temperatures.

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In contrast, temperature, population and their interaction significantly influenced 261 fertility (Fig. 2, Table 1). Stressful temperatures experienced only during 262 development (HB, Fig. 2) had a greater consequence on fertility than only 263 experiencing stressful temperatures as an adult (BH, Fig. 2) and this affect was 264 stronger on the Uppsala population than the Valencia population. We predicted that 265 the southern population would exhibit positive, whereas the northern population 266 would exhibit negative, carry-over effects. To test these predictions, we performed 267 two directional (and thus, one-tailed) t-tests, contrasting the effect of HH relative to 268 HB (HH>HB in Val, HB>HH in Upp). There was no difference in fertility for Uppsala 269 between stress in the juvenile stage only compared with stress in both stages ( $t_{328}$  = 270 0.99, P = 0.16). However, in Valencia, while heat stress negatively impacted fertility 271 during any stage, fertility was higher in individuals that experienced heat stress in 272

both juvenile and adult stages compared to heat stress only during the juvenile stage 273  $(t_{328} = 1.74, P = 0.041).$ 274

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#### Discussion 276

In this study, we aimed to reveal local adaptation of reproductive performance traits 277 under sub-lethal heat stress, with a particular emphasis on heat stress during the 278 juvenile stage. Furthermore, we aimed to evaluate whether acclimation to heat stress 279 during development impacted the late life response to these temperatures. We found 280 281 juvenile heat stress and population impacted egg-to-adult viability and sperm motility, with the northern population having lower fitness than the southern population. We 282 found fertility exhibited local adaptation; although both populations were negatively 283 284 impacted during juvenile heat stress, the southern population was less effected. We also found positive carry-over effects in fertility. Heat stress exposure during the 285 adult stage, after juvenile heat stress exposure, increased fertility in the southern, but 286 not northern, population. Overall, the northern population was significantly more 287 susceptible to juvenile and adult thermal stress across all fitness traits measured. 288 Our work fills a gap in research on temperature effects that has largely been 289 dominated by experiments focusing on CTLs of non-reproductive traits, primarily 290 following adult exposure to thermal, particularly cold, stress. In this work, we have 291 292 linked reproductive performance based responses to chronic sub-lethal heat stress, particularly at the juvenile stage, to local adaptation. This work will provide novel 293 insights of how populations may respond to such thermal variation and may be 294 applicable across a variety of other animals. The generality of the results is likely 295 because many animals will lose fertility at temperatures much lower than fatal 296

thermal limits (Jørgensen *et al.*, 2006) and because, for many invertebrates, gamete
production begins during juvenile stages.

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As predicted, we found no evidence of local adaptation in juvenile survival although 300 northern males had lower egg-to-adult viability. Likewise, sperm motility was 301 negatively impacted primarily by juvenile temperature and weakly by population, with 302 northern males having lower sperm motility overall than southern males but no 303 evidence of local adaptation. Sperm motility is a standard measure of male sterility 304 305 but is frequently measured as a binary trait, and thus may not reveal subtle changes in sperm performance following thermal stress. The lower general fitness of the 306 Uppsala population may reflect range margin effects (Kawecki, 2008), consistent 307 308 with the decreased variation in chromosomal inversion polymorphisms as latitude increases (Krimbas, 1993). Decreased genetic variation has been suggested to 309 occur at range margins as a consequence of some combination of Allee effects. 310 genetic drift and the lack of sufficient gene flow into populations at range limits to fuel 311 adaptive potential (Hoffmann & Blows, 1994; Bridle & Vines, 2007). Alternatively, 312 local adaptation may be decreased due to gene swamping, with dispersal from 313 (typically larger) more central populations continually introducing alleles into range 314 margins that are more appropriate for maximising fitness elsewhere (Hoffmann & 315 316 Blows 1994). The situation in *D. subobscura* is more complicated given the presence of extensive variation in chromosomal inversion polymorphisms that prevent 317 recombination in inversion heterozygotes. While there is gene flow within the same 318 inversion arrangement (Simões et al., 2012; Pegueroles et al., 2013; Pratdesaba et 319 al., 2015), there is substantial evidence that inversion polymorphisms themselves 320 are under selection (Balanyá et al., 2006; Santos et al., 2016). 321

We did, however, demonstrate local adaptation to the thermal environment in our 323 other measure of fertility, as predicted. Fertility was negatively impacted by 324 application of juvenile heat stress, particularly for Uppsala compared to Valencia 325 pairs. Juvenile heat stress resulted in positive carry-over effects on fertility in 326 Valencia, but not Uppsala, when adults also experienced heat stress. These results 327 328 indicate that northern populations are more susceptible to sub-lethal reproductive consequences following exposure to juvenile heat stress, and that such exposure 329 330 does not buffer reproductive performance against subsequent adult heat stress.

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The effects of thermal stress on male and female reproductive function in the fertility 332 experiment cannot be separated. Now that we have demonstrated local adaptation in 333 fertility, future work should decompose the sex-specific effects of juvenile thermal 334 stress on gamete performance in more physiological detail, such as quantifying 335 apoptosis of oocytes and the effect of thermal stress on sperm number and length. 336 Female D. subobscura are monogamous (Smith, 1956; Holman et al., 2008; Fisher 337 et al., 2013) but, as with other obscura group species, males are sperm 338 heteromorphic, with males producing short, non-fertile sperm (parasperm) and 339 longer, fertile sperm (eusperm; Snook & Karr, 1998). In work on the closely related 340 341 polyandrous species, *D. pseudoobscura*, parasperm protect brother eusperm from a hostile female reproductive tract (Holman & Snook, 2008). Sperm precedence 342 favours the last male, but after lengthy exposure to cold stress, sperm precedence 343 reverses to favour the first male (Giraldo-Perez et al., 2016). Whether, and the extent 344 to which intra-ejaculate interactions and ejaculate-female reproductive tract 345

interactions change as a consequence of either juvenile or adult heat stress and
 whether this is impacted by mating system is unknown.

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We have previously found, across six *D. subobscura* populations from a 20° 349 latitudinal cline in Europe (of which the Valencia and Uppsala populations represent 350 the southern- and northern-most population respectively), a gradient in gene 351 expression in which adult males from southern populations have higher expression 352 of spermatogenesis genes than males from northern populations, under benign 353 354 (18°C) conditions ((Porcelli *et al.*, 2016). Given that females are monandrous, sperm competition cannot explain increased investment in spermatogenesis in lower 355 latitude populations. As investment in spermatogenesis is less in Uppsala than 356 357 Valencia, it may be that northern males are not able to compensate as much as southern males following juvenile heat stress. In support, we found evidence of 358 developmental acclimation in Valencia, but not Uppsala, populations. 359

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If fertility reduction following juvenile heat stress is male-driven, then population 361 fitness can recover only if females remate with a fertile male. While females are 362 naturally monogamous, previous work has shown that *D. subobscura* females will 363 remate if they do not receive a functional ejaculate during copulation, a phenomenon 364 termed pseudo-polyandry (Fisher et al., 2013). Thus, mitigating male sterility and 365 recovering population fitness may be possible if females can find a fertile mate, 366 although whether females will remate after receipt of a partially-fertile ejaculate is 367 unknown. Moreover, finding a fertile mate may be more difficult in northern, more 368 susceptible populations, because of the lower density of range margin populations 369 (including the *D. subobscura* Uppsala population; Krimbas, 1993). However, if 370

oogenesis is also damaged during juvenile heat stress, then even pseudo-polyandry 371 cannot rescue population fitness. The combination of higher latitude populations 372 being more susceptible to juvenile heat stress, decreasing both survival and 373 measures of reproductive performance (sperm motility and fertility), northern 374 populations investing less in spermatogenesis, and having low population density, 375 makes high latitude *D. subobscura* populations susceptible to extinction as both 376 average temperature and the frequency of extreme heat events increase (IPCC, 377 2014). 378

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While this work is specific for *D. subobscura*, such results may be quite common. 380 Many organisms spend a portion of their life cycle as relatively immobile juveniles, so 381 382 thermal stress during this life cycle stage may have profound negative fitness consequences beyond survival because gamete production typically starts during 383 juvenile stages. Such subsequent negative fertility effects on adults should 384 compound reductions in population size, which can then have substantial 385 downstream effects via, for example, genetic drift (Polechova & Barton, 2015). As 386 range margin populations tend to have less genetic diversity to begin with, and the 387 range of variation in resistance to thermal stress is considerably less at high 388 temperatures than at cold (Gaston & Chown, 1999; Addo-Bediako et al., 2000; 389 Terblanche et al., 2007), sub-lethal effects on reproductive traits may make these 390 populations even more susceptible as the climate continues to get warmer and the 391 frequency of extreme heat events increases. Local adaptation may provide some 392 resilience to climate change and we found such local adaptation for juvenile heat 393 stress and reproductive performance in a southern population. Variation in 394 organismal tolerance throughout the life cycle should be incorporated in models that 395

predict impacts of climate change. To date, heavy reliance has necessarily been 396 placed on predictions based on latitudinal trends in upper and lower critical or lethal 397 thermal limits (Lancaster, 2016) derived from data compilations (e.g. Addo-Bediako 398 et al., 2000; Sunday et al., 2011). A recent study on lizards, with a sessile embryonic 399 stage, developed a life cycle model for demographic consequences based on 400 microclimates at high spatio-thermal resolution and experimentally derived 401 embryonic thermal tolerances (Levy et al., 2015). Levy and colleagues found that 402 models that ignored embryonic consequences (egg-to-adult viability) severely 403 underestimated the demographic effects of (previously unrecognised) thermal 404 events. Such detailed analysis that also incorporates any subsequent fertility effects 405 remains to be done. Given that we show substantial effects beyond egg-to-adult 406 407 survival, models without this consideration may prove even more unduly conservative. 408

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Table 1. Generalized linear model of the effects of temperature, population and their
interaction on (a) egg-to-adult viability, (b) sperm motility and (c) fertility. *p* values are
reported either exactly or < 0.001. For egg-to-adult viability, the starting number of</li>
eggs was also included in the model. Non-significant terms have been dropped.
Residual deviance is included where the quasibinomial model was used.

source	d.f.	Deviance	Deviance ratio	p
(a) egg-to-adult viability				
log(eggs)	1	67.232	6.1864	0.01
Temperature	1	302.528	27.8375	< 0.001
Population	1	59.993	5.5203	0.01
Residual	394	4937.1		
(b) sperm motility				
Temperature	1	76.95		<0.001
Population	1	4.49		0.03
(c) fertility				
Temperature	3	148.731	28.77	<0.001
combination				
Population	1	11.224	6.5128	0.01
Temperature*Population	3	14.657	2.8348	0.038
Residual	328	587.43		

569 Figure legends.

Figure 1. The effect of juvenile heat stress and population influence a) egg-to-adult
viability and b) sperm motility. Mean ± SE relative to Valencia (southern population)
at the benign temperature, following back transformation from the logit scale.

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Figure 2. Heat stress during the juvenile life cycle stages negatively impacts fertility 574 in Valencia (southern population) pairs significantly less than Uppsala (northern 575 population) pairs and subsequent adult heat stress improves fertility in Valenica, but 576 not Uppsala, pairs. B= benign and H = heat stress; BB = benign temperature for both 577 juvenile and adult stages, BH = benign temperature during juvenile stages but heat 578 stress as adults, HB = heat stress during juvenile stages but not during the adult 579 stage; HH = heat stress during juvenile and adult stages. Mean  $\pm$  SE relative to 580 Valencia at the benign temperature for BB, following back transformation from the 581 logit scale. 582