

Acute acetaminophen ingestion improves performance and muscle activation during maximal intermittent knee extensor exercise

Original investigation

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1 **Abstract**

2 **Aim:** Acetaminophen is a commonly used medicine for pain relief and emerging evidence
3 suggests that it may improve endurance exercise performance. This study investigated some
4 of the physiological mechanisms by which acute acetaminophen ingestion might blunt
5 muscle fatigue development. **Methods:** Thirteen active males completed 60×3 s maximum
6 voluntary contractions (MVC) of the knee extensors with each contraction separated by a 2 s
7 passive recovery period. This protocol was completed 60 min after ingesting 1 g of
8 maltodextrin (placebo) or 1 g of acetaminophen on two separate visits. Peripheral nerve
9 stimulation was administered every 6th contraction for assessment of neuromuscular fatigue
10 development, with the critical torque (CT), which reflects the maximal sustainable rate of
11 oxidative metabolism, taken as the mean torque over the last 12 contractions. Surface
12 electromyography was recorded continuously as a measure of muscle activation. **Results:**
13 Mean torque (61 ± 11 vs. 58 ± 14 % pre-exercise MVC) and CT (44 ± 13 vs. 40 ± 15 % pre-
14 exercise MVC) were greater in the acetaminophen trial compared to placebo (both $P < 0.05$).
15 Voluntary activation and potentiated twitch declined at a similar rate in both conditions
16 ($P > 0.05$). However, the decline in electromyography amplitude was attenuated in the
17 acetaminophen trial, with electromyography amplitude being greater compared to placebo
18 from 210 s onwards ($P < 0.05$). **Conclusion:** These findings indicate that acute acetaminophen
19 ingestion might be ergogenic by increasing CT and preserving muscle activation during high-
20 intensity exercise.

21

22 **Key words:** Analgesic; critical torque; electromyography; neuromuscular fatigue; single leg
23 exercise

24 **Abbreviations**

ACT	Acetaminophen (paracetamol)
CT	Critical torque (i.e., asymptote of the P-T _{lim} relationship)
EMG	Electromyography
EMG_{RMS}	EMG amplitude using root mean square method
M-wave	M-wave amplitude
M_{max}	Maximal m-wave amplitude
MVC	Maximal voluntary contraction
PL	Placebo
pTw	Potentiated twitch
RMS	Root mean square
sTw	Superimposed twitch (onto an MVC)
VA	Voluntary activation (%)
W'	Curvature constant of the hyperbolic Torque-T _{lim} relationship

25

26 **Introduction**

27 Neuromuscular fatigue is defined as a decrease in skeletal muscle force production capacity
28 (Gandevia, 2001). This neuromuscular fatigue development can arise from physiological
29 perturbations within the central nervous system, termed central fatigue, or within, or distal to,
30 the neuromuscular junction, termed peripheral fatigue (Gandevia, 2001). Exercise-induced
31 fatigue is a complex, multi-factorial process, the physiological bases of which are still widely
32 debated (Enoka & Duchateau, 2008; Gandevia, 2001; Hureau et al. 2016; Place et al. 2010;
33 Taylor et al. 2016). Indeed, recent research suggests that peripheral fatigue and central
34 fatigue develop inter-dependently and are likely to interact in a coordinated manner to
35 determine neuromuscular fatigue development (Hureau et al. 2016).

36

37 Following the onset of muscle contractions, group III and IV afferents discharge in response
38 to mechanical and metabolic stimuli, contributing to the sensation of muscle pain during
39 exercise (O'Connor & Cook, 1999; McCord and Kaufmann, 2010; Pollak et al. 2014). Group
40 III/IV muscle afferent feedback, and the associated sensation of pain, appears to play a role in
41 neuromuscular fatigue development through modulating both central and peripheral fatigue.
42 Indeed, when the ascending projection of group III and IV muscle afferents is attenuated via
43 intrathecal fentanyl administration, central motor drive is increased (as inferred via EMG)
44 and peripheral fatigue development is expedited (Amann et al. 2009, 2011; Blain et al. 2016).
45 Therefore, interventions that can modulate skeletal muscle pain sensation have the potential
46 to impact muscle activation and exercise-induced fatigue development.

47

48 Acetaminophen (ACT; or paracetamol) is a commonly used non-prescription pain reliever
49 which is considered one of the safest non-opioid analgesics at therapeutic doses (Toussaint et
50 al. 2010). Acute ACT consumption has been shown to improve exercise performance

51 concomitant with a lower pain sensation (Foster et al. 2014; Mauger et al. 2010). The
52 mechanisms that underlie the analgesic effect of ACT are not completely understood but are
53 considered to be predominantly mediated by central factors (Anderson, 2008; Graham et al.
54 2013; Smith, 2009; Toussaint et al. 2010). Conventionally, the analgesic effect of ACT has
55 been attributed to the inhibition of cyclooxygenase enzymes, which stimulate nociceptor
56 discharge through the synthesis of prostaglandins (Graham et al. 2013; Józwiak-Bębenista
57 and Nowak, 2014). There is also evidence that the analgesic effect of ACT is linked to
58 potentiation of descending serotonergic pathways (Pickering et al. 2006, 2008), and to
59 modulation of opioid and cannabinoid receptors (Graham et al. 2013). These mechanisms
60 likely interact to lower pain sensation after acute ACT ingestion by increasing the
61 nociceptive stimuli required to evoke a given pain sensation. Although ACT ingestion has
62 been reported to increase resting cortico-spinal excitability, as inferred from an increased
63 motor evoked potential amplitude (Mauger & Hopker, 2013), the neuromuscular bases for
64 blunted exercise-induced fatigue development following ACT ingestion have yet to be
65 investigated.

66

67 In addition to altering pain sensation and neuromuscular function, ACT ingestion might
68 attenuate fatigue development and enhance performance by modulating aspects of the power
69 or torque-duration relationship. The asymptote of the hyperbolic torque-duration relationship
70 is termed the critical torque (CT) and can be estimated from the mean torque produced over
71 the last 12 maximum voluntary contractions (MVCs) of a 60 MVC knee extension protocol
72 (Burnley, 2009). The CT reflects the maximal sustainable rate of oxidative metabolism (Jones
73 et al. 2008; Jones et al. 2011 *for review*) and represents a critical threshold for neuromuscular
74 fatigue development (Burnley et al. 2012). The curvature constant of the torque-duration
75 hyperbola is termed the W' and represents a fixed amount of torque-impulse (surrogate

76 measure of 'total work') that can be completed above CT (Burnley and Jones, 2016; Jones et
77 al. 2008; Poole et al. 1988). Together, CT (or critical power, critical speed for other forms of
78 exercise) and W' (or its equivalent) can be used to accurately predict exercise performance
79 (Black et al. 2014, Burnley et al. 2012). An intervention that enhances high-intensity
80 exercise tolerance or performance, or attenuates fatigue development, would be expected to
81 enhance CT or W' (or their exercise-modality-specific equivalents) For example, critical
82 power, but not W' , is increased following endurance training concomitant with improved
83 endurance exercise performance (Vanhatalo et al. 2008). Therefore, the reported
84 improvement in exercise performance with acute ACT ingestion (Foster et al. 2014; Mauger
85 et al. 2010, 2014) would be expected to be linked to improvements in CT and/or W' , but this
86 has yet to be investigated.

87

88 The purpose of this study was to test the hypotheses that, compared to placebo, acute
89 consumption of 1 g ACT would reduce the rate of neuromuscular fatigue development, and
90 increase total impulse (torque-time integral), CT and muscle activation during a 5 min single-
91 leg intermittent MVC test.

92

93 **Materials and methods**

94 *Participants*

95 Thirteen healthy male volunteers (mean \pm SD: age 31 ± 7 years, height 1.76 ± 0.08 m, body
96 mass 75 ± 11 kg) provided written, informed consent to participate in the present study,
97 which was approved by the Ethics Committee of Sport and Health Sciences (University of
98 Exeter). After being informed of the experimental procedures and associated risks, all
99 participants completed a medical health questionnaire to ensure it was safe to consume ACT
100 prior to performing exhaustive exercise. Subjects were not consumers of any 'pain relief'

101 medication (prescription or non-prescription) over the course of the study. None of the
102 subjects had a history of motor or neurological disorders.

103

104 *Experimental Design*

105 Subjects visited the laboratory on three occasions over a 3- to 4-week period with all tests
106 separated by at least 1 week and conducted at a similar time of day (± 90 min) to limit
107 changes in quadriceps strength and to account for diurnal variations in neuromuscular
108 excitability. The first laboratory visit was used to familiarise subjects to the measurements
109 and experimental protocol described below. During these sessions, the settings and placement
110 of the dynamometer and electromyography (EMG) and peripheral nerve stimulation
111 electrodes were recorded for each subject. Subsequently, subjects performed the fatiguing
112 protocol under two conditions (*see 'Experimental Protocol'*): placebo (PL) and ACT.

113

114 *Experimental Protocol*

115 Subjects completed a preliminary trial for familiarisation to the measurement techniques and
116 experimental protocol. An isokinetic dynamometer (Biodex System 3, Shirley, NY, USA)
117 was used in all tests and adjusted so that the axis of rotation of the lever arm was in line with
118 the lateral epicondyle of the right femur. Subjects were seated with the hip and knee joints at
119 relative angles of 155° and 90° , respectively. The remainder of the chair settings were
120 recorded and replicated in all subsequent trials to ensure an identical body position was
121 assumed throughout the experimental trials. The semi-supine position was employed as it
122 permitted better access to the superficial femoral nerve for peripheral nerve stimulation as
123 verified during pilot testing. Inelastic padded Velcro straps were fastened at the ankle,
124 quadriceps, hip and shoulders to maintain a stable body position. The procedures adopted
125 during the familiarisation trial were replicated in all experimental trials.

126

127 The trials on visits 2 and 3 were completed in a double-blind, randomised fashion using a
128 cross-over experimental design. Prior to each visit, subjects were required to refrain from
129 caffeine (for at least 12 h), strenuous exercise and alcohol (for at least 24 h), analgesics and
130 any form of anti-inflammatory drug (for the duration of the experimental trial) and to arrive
131 in a fully rested, hydrated state. Subjects were instructed to maintain their usual diet and
132 exercise regime during the study. 1 g of maltodextrin (placebo) or 1 g of acetaminophen was
133 ingested orally, 60 minutes prior to the exercise bout such that the start of the exercise trial
134 was expected to coincide with attainment of the peak plasma [acetaminophen] (Anderson et
135 al. 2008; Forrest, Clements & Prescott, 1982). Following oral ingestion, ACT is rapidly
136 absorbed from the gastrointestinal tract and its bioavailability ranges from 70 to 90%
137 (Forrest, Clements & Prescott, 1982). The trials started with a standardised isometric warm-
138 up routine (10 isometric contractions for 3 s at 50% of pre-exercise MVC as determined
139 during familiarisation testing) and testing of the optimal EMG electrode, anode, and cathode
140 placement and stimulation intensity for peripheral nerve stimulation. Neuromuscular function
141 was assessed pre-trial, during and (< 10 s) post-trial. Single peripheral nerve stimulation
142 pulses were manually triggered at rest to determine pre-exercise neuromuscular function,
143 namely the characteristics of the M-wave response (M-wave amplitude; M_{max}) to supra-
144 maximal nerve stimulation, contractility of the muscle (i.e. maximal rate of force
145 development, half relaxation time and contraction time), voluntary activation (VA) and
146 potentiated twitch force (pTw). During maximal voluntary contractions (MVCs), peripheral
147 nerve stimulation pulses were triggered to occur as soon as a peak torque was achieved
148 (typically 1.5 s into a 3 s contraction), each separated by a 40 s rest. The stimuli were also
149 delivered 1-2 s after the cessation of the contraction to provide a resting pTw. Identical

150 measurements were repeated as soon as possible (<10 s) after the fatiguing exercise to
151 determine post-exercise neuromuscular function (see Fig. 1).

152

153 The intermittent isometric contraction protocol used in this study had a similar design to that
154 presented by Burnley (2009). The protocol consisted of repeated brief MVCs (3 s contraction,
155 2 s rest), timed via a visual prompt to ‘go’ and ‘relax’ (separate to the torque output screen),
156 accompanied by the same verbal instructions from the experimenter. The protocol was
157 terminated when the subjects completed the 60th MVC. Every 6th contraction was
158 accompanied by peripheral nerve stimulation during and post MVC (as described for pre- and
159 post-trial measurements). Subjects were not given a visual representation of the torque
160 produced during each MVC or made aware of the number of MVCs elapsed during the
161 protocol. Subjects were instructed to continue to perform maximal contractions throughout.
162 The coefficient of variation for MVC performance (mean torque through 60 MVCs) and
163 electromyography root mean square (EMG_{RMS}) amplitude using this experimental set up was
164 2.9% and 4.8%, respectively as calculated during pilot testing.

165

166 *Measurements*

167 *Torque*

168 Knee-extensor torque from the Biodex dynamometer was sampled at 1,000 Hz and low-pass
169 filtered at 40 Hz, before being displayed on a wide screen monitor using Spike2 (CED,
170 Cambridge, UK). Torque was expressed throughout as a percentage (%) of pre-exercise
171 MVC.

172

173 *Electromyography*

174 Surface EMG activity was recorded from vastus lateralis, vastus medialis and rectus femoris
175 of the quadriceps and biceps femoris of the hamstring of the right leg using active bipolar bar
176 electrodes single differential configuration (DE2.1, DelSys Inc, Boston, MA, USA). The
177 electrodes were placed over the respective muscle bellies (SENIAM guidelines). Double-
178 sided adhesive tape and a hypoallergenic medical tape were used to ensure the EMG sensor
179 stability for recording electrodes. The skin area underneath each EMG electrode was shaved,
180 then exfoliated and cleaned with alcohol to minimise the skin impedance. The EMG and
181 torque signals were pre-amplified (1,000 x), band-pass filtered (20–450 Hz, Bagnoli-8,
182 DelSys Inc, Boston, MA, USA), and then transferred to a computer with a sampling
183 frequency of 2 kHz and high-pass filtered at 10 Hz. EMG and torque data were recorded
184 continuously and digitised synchronously with 16 bit resolution via an A/D converter (± 5 V
185 range, CED 1401 power, Cambridge, UK). The electrodes were used to record: 1) evoked
186 muscle action potentials (peak-to-peak amplitude of the M-wave); and 2) EMG to estimate
187 muscle activity and the output of spinal motoneurons (motor unit recruitment and firing
188 frequency). EMG was average rectified using the root mean square method (EMG_{RMS}).
189 EMG_{RMS} was then normalised to the pre-exercise maximum (or maximal EMG signal) and
190 the local M-wave amplitude (closest measure of the M-wave to the MVC) in order to exclude
191 any changes to the EMG trace to changes in local excitability. The ground electrode was
192 placed over the patella of the right leg.

193

194 *Peripheral Nerve Stimulation*

195 Electrical stimulation was applied by a constant current stimulator (Digitimer Stimulator
196 DS7, Digitimer, UK). M-waves were elicited by supramaximal percutaneous electrical
197 stimulation of the femoral nerve (200 μ s duration). The cathode was placed over the femoral
198 nerve in the inguinal fossa, approximately 3–5 cm below the inguinal ligament in the femoral

199 triangle. Once the M-wave was elicited, the maximum amplitude (peak-to-peak) of the M-
200 wave was determined (M_{\max}) for the vastus lateralis and vastus medialis. To determine the
201 stimulation intensity (current), single stimuli were delivered in 20 mA step-wise increments
202 from 100 mA until a plateau in quadriceps pTw and M-wave were observed. To ensure a
203 supramaximal response, the current was increased by an additional 30% (mean \pm SD current
204 = 194 ± 81 mA; Burke, 2002; Goodall et al. 2010; Neyroud et al. 2014). The average M_{\max}
205 was obtained from 3 stimuli, with ~8-10 s separating each pulse at rest. Peak torque, maximal
206 rate of force development, half relaxation time and contraction time were analysed for all
207 pTw.

208

209 *Data Analyses*

210 Data were analysed using a custom written script developed in Spike2 software (CED,
211 Cambridge, UK). Mean torque for each 3 s contraction was determined for all tests as the
212 mean value over a 1 s period which approximated the plateau level of the highest torque (i.e.
213 500 ms before and after the peak torque). The pTw was calculated as the peak torque
214 achieved following the single pulse delivered 1 s post-MVC. The twitch torque superimposed
215 onto the peak force production of the MVC (sTw) was calculated as the increment in torque
216 immediately following the pulse during MVCs. The torque impulse was calculated as the area
217 under the torque-time curve by accumulating the time integral of each MVC (3 s) by the
218 difference in torque between MVCs. The end-test torque (i.e. CT) during the 60 MVC test
219 was defined as the mean of the last 12 contractions (i.e. the last 60 s; Burnley, 2009). The W'
220 was calculated as the area above the CT from the torque-time curve (i.e. impulse above CT).
221 Central fatigue was assessed as the maximal voluntary activation of the motoneuron pool
222 (VA, %), calculated using the interpolated twitch method from peripheral nerve stimulation
223 (Merton, 1954). Specifically, the increment in torque evoked during the MVCs was expressed

224 as a fraction of the amplitude of the potentiated twitch produced with the same stimulus in the
225 relaxed muscle post MVC. The level of voluntary drive was then quantified as a percentage:
226 $[1 - \text{evoked torque (superimposed on voluntary torque, sTw)} / (\text{mean control evoked}$
227 $\text{response, pTw}) \times 100]$ (i.e. Allen et al. 1998).

228

229 The changes in maximal voluntary torque, pTw, VA and EMG_{RMS}, were used to quantify
230 peripheral fatigue and central fatigue. The maximal EMG was taken from the first MVC
231 during the 60 MVC task and compared to the last MVC at task end. The neuromuscular
232 parameters extracted from the three sets of maximal contractions completed post-exercise
233 were tested for statistical differences between sets of contractions and then compared to the
234 first set of MVCs completed pre-exercise (Froyd et al. 2013; Pageaux et al. 2015a; Doyle-
235 Baker et al. 2017). Neuromuscular function was also measured for each of the stimulated
236 contractions during the exercise and normalised to the corresponding pre-exercise values at
237 100% MVC. All neuromuscular parameters and torque were averaged across the protocol
238 using 6 (30 s) bin averages.

239

240 *Statistics*

241 Paired-samples *t*-tests were used to compare the mean torque, total impulse, CT and *W'* (i.e.
242 impulse above CT) between ACT and PL conditions. In addition, paired samples *t*-tests were
243 used to assess parameters of neuromuscular function at task end between trials including
244 pTw, maximal rate of force development, half relaxation time and contraction time. The
245 profiles of torque, VA, pTw and m-wave amplitude were analysed using two-way ANOVAs
246 with repeated measures (using 12 contraction averages; i.e. 6 time points). Normalised
247 EMG_{RMS} were analysed using two-way ANOVAs with repeated measures using 30 s mean
248 values (i.e. 10 time points). Where the ANOVA revealed a significant interaction effect, post-

249 hoc tests were completed using a Bonferroni correction. For calculation of effect size, partial
250 eta squared (η^2) was used for omnibus tests. Cohen's d was used to calculate the effect size
251 for paired t-tests and post-hoc comparisons. A t-test was also conducted on the differences in
252 EMG and force production (torque) from task end (i.e. 300 s) to 150 s (mid-point) to assess
253 the rate of change as the protocol progressed. All statistical tests were performed both on %
254 change and raw data. Where sphericity was violated, a greenhouse-geisser correction factor
255 was used. For all tests, results were considered statistically significant when $P < 0.05$. Data are
256 presented as means \pm SD unless otherwise indicated. All statistical analyses were conducted
257 using IBM SPSS Statistics version 23.

258

259 **Results**

260 The mean MVC torque achieved prior to the 60 MVC protocol was 232 ± 47 and 228 ± 48
261 N.m for PL and ACT, respectively. Voluntary activation of the knee extensors achieved
262 during the preliminary MVCs was $88 \pm 7\%$ and 87 ± 5 for PL and ACT, respectively.
263 Baseline MVC and VA were not different between conditions ($P > 0.05$).

264

265 *60 MVC Performance*

266 The profile for mean torque across all subjects and a representative individual plot during
267 each contraction for the 60 MVC protocol is shown in Fig. 2. During the PL trial, torque
268 declined from a peak of $99 \pm 3\%$ MVC (relative to pre-exercise MVC) during the first
269 contraction to $40 \pm 15\%$ MVC during the last 12 contractions ($P < 0.0001$, $\eta^2 = 0.908$; Table 1,
270 Fig. 2). The mean torque (relative to pre-exercise MVC) achieved across the 60 MVCs was
271 greater with ACT ($61 \pm 11\%$, 97.1 ± 23.2 N.m) compared to PL ($58 \pm 14\%$, 83.8 ± 22.7 N.m;
272 $P = 0.030$, $d = 0.656$) and there was a significant interaction effect (time \times condition; $P = 0.036$,
273 $\eta^2 = 0.260$). Post-hoc tests revealed significant differences in torque between conditions at task

274 end ($P=0.044$, $d=0.656$) but at no other time-points (all $P>0.084$ and $d<0.548$). CT was
275 higher with ACT compared to placebo (ACT: 44 ± 13 % vs. PL: 40 ± 15 %, $P=0.011$,
276 $d=0.691$), with no between-condition differences in W' (PL: 6.97 ± 2.43 , ACT: 6.91 ± 2.54
277 N.m.s; $P=0.879$, $d=0.026$). Total impulse in the 60 MVC protocol was higher with ACT
278 (24386 ± 3793 N.m.s) compared to PL (22055 ± 3885 N.m.s; $P=0.006$, $d=0.973$). As time
279 progressed, the difference in force production between conditions increased as evidenced by
280 a significant difference in rates of change between conditions (i.e. difference between 300
281 and 150 s time points; $P=0.035$, $d=0.696$). The individual responses following ACT
282 supplementation to the torque-impulse can be seen in Fig. 3. Table 2 illustrates the
283 parameters of the 60 MVC test for PL and ACT for individual subjects and is available as an
284 online supplement.

285

286 *Neuromuscular Function*

287 There was a main effect for time on pTw (Fig. 4a, $P<0.001$, $\eta^2=0.754$), voluntary activation
288 (Fig. 4b, $P<0.001$, $\eta^2=0.647$) and m-wave amplitude (Fig. 4c, $P=0.005$, $\eta^2=0.281$), which all
289 declined as the protocol progressed. VA declined from 89 ± 8 % to 59 ± 19 % and from $88 \pm$
290 5 % to 62 ± 16 % in the PL and ACT conditions ($P<0.0001$, Fig 3), respectively. However,
291 there were no main condition effect on M-wave amplitude ($P=0.733$, $\eta^2=0.012$), pTw
292 ($P=0.783$, $\eta^2=0.032$) or VA ($P=0.841$, $\eta^2=0.004$) between the PL and ACT conditions (Fig.
293 4c). Likewise, there was no significant interaction effect (i.e. time \times condition) on M-wave
294 amplitude ($P=0.993$, $\eta^2=0.009$) or VA ($P=0.387$, $\eta^2=0.097$). In addition pTw declined from
295 68 ± 8 to 30 ± 20 N·m and from 71 ± 12 to 31 ± 23 N·m (both $P<0.0001$) for PL and ACT,
296 respectively, but there was no significant difference in end-exercise pTw between conditions
297 ($P=0.763$, $d=0.047$). There was also no significant difference in end-exercise maximal rate of

298 force development ($P=0.181$, $d=0.383$), half relaxation time ($P=0.234$, $d=0.341$) or
299 contraction time ($P=0.595$, $d=0.232$) between conditions.

300

301 Using 6 contraction (30-s) bin means, EMG_{RMS} decreased from $99 \pm 4\%$ to $59 \pm 17\%$ in the
302 PL trial (from first 6 to last 6 contractions; $P<0.0001$, $\eta^2=0.502$; Fig. 5). This decline in
303 EMG_{RMS} was attenuated following ACT ingestion (100 ± 5 to $87 \pm 28\%$), demonstrating a
304 significant main effect of condition ($P=0.033$, $\eta^2=0.381$) and an interaction effect ($P=0.043$,
305 $\eta^2=0.229$). The EMG_{RMS} was elevated at 60 s ($P=0.036$, $d=0.730$), 90 s ($P=0.020$, $d=0.835$),
306 210 s ($P=0.070$), 240 s ($P=0.015$, $d=0.886$), 270 s ($P=0.019$, $d=0.844$) and 300 s ($P=0.001$,
307 $d=1.306$) in ACT compared to PL (Fig. 5). As time progressed, the difference in EMG
308 amplitude between conditions increased as evidenced by a significant difference in rates of
309 change between conditions (i.e. difference between 300 and 150 s time points; $P=0.024$,
310 Cohen's $d=0.804$).

311

312 **Discussion**

313 Consistent with our experimental hypotheses, the principal findings of this study were that
314 acute ACT ingestion increased mean MVC torque, and increased CT and EMG amplitude
315 during the latter stages of the 60 MVC protocol. These observations offer insights into the
316 mechanisms by which ACT blunts fatigue development and suggest that the ergogenic effect
317 of ACT ingestion may be linked to increases in CT and muscle activation during fatiguing
318 skeletal muscle contractions.

319

320 In the present study, fatigue development and some of its underpinning mechanisms were
321 assessed during the completion of 60 MVCs of the knee-extensors using the protocol
322 described by Burnley (2009). Consistent with Burnley (2009), torque and pTw declined by

323 60% and 30%, respectively, and VA declined to 59%, reflecting peripheral and central
324 fatigue development. However, although neuromuscular fatigue was also manifest during the
325 ACT trial, the acute ingestion of ACT increased the mean torque during the fatiguing 60
326 MVC protocol by ~4% compared to the PL condition. This finding is in line with previous
327 observations that acute ACT ingestion can improve performance during whole body exercise
328 in humans (Foster et al. 2014; Mauger et al. 2010a).

329

330 The attenuation of neuromuscular fatigue development following ACT ingestion was not
331 accompanied by specific alterations in central fatigue (voluntary activation), peripheral
332 fatigue (potentiated twitch, pTw) or peripheral neuromuscular excitability (M-wave) between
333 the ACT and placebo trials. However, while EMG declined across the 60 MVC protocol in
334 both conditions, it declined to a lesser extent with ACT (declined to ~87 %) compared to
335 PLA (declined to ~59%). This observation suggests that improved muscle activation might
336 have contributed to the ergogenic effect of ACT ingestion. This interpretation is strengthened
337 by our observation of a positive correlation between the inter-trial changes in EMG and
338 torque ($r = 0.85$). Although ACT ingestion has previously been reported to increase cortico-
339 spinal excitability at rest (Mauger & Hopker, 2013), the neuromuscular mechanisms for
340 improved performance during exercise had not been explored in previous studies reporting an
341 ergogenic effect of ACT ingestion (Foster et al. 2014; Mauger et al. 2010, 2014). Therefore,
342 by suggesting that ACT can improve muscle activation during repeated fatiguing skeletal
343 muscle contractions, our findings extend previous observations by providing insight into the
344 potential neuromuscular bases for the ergogenic effect of ACT ingestion.

345

346 It is possible that the effect of ACT may be linked to a sub-conscious neuromuscular
347 alteration during exercise via a reduction in the magnitude of muscle afferent feedback.

348 During exercise, ascending group III and IV muscle afferents discharge in response to
349 noxious, metabolic and mechanical stimuli within the contracting skeletal muscles to regulate
350 the sensation of pain, muscle activation and peripheral fatigue development (Amann et al.
351 2009, 2011; Blain et al. 2016; Hureau et al. 2016; McCord and Kaufmann, 2010; Pollak et al.
352 2014). When the magnitude of type III and IV muscle afferent feedback is reduced,
353 performance is compromised and pacing strategy is adversely impacted (Amann et al. 2009,
354 2011; Blain et al. 2016), despite enhanced muscle activation, as peripheral fatigue
355 development is exacerbated. ACT administration, on the other hand, which seemingly acts
356 predominantly through central processes to blunt pain sensation (Anderson, 2008; Graham et
357 al. 2013; Smith, 2009; Toussaint et al. 2010), appears to have attenuated the decline in
358 muscle activation without impacting peripheral fatigue development, culminating in blunted
359 neuromuscular fatigue development in the current study. Taken together, these observations
360 suggest that reducing, but not abolishing, pain sensation can attenuate neuromuscular fatigue
361 development during exercise or that a higher magnitude of afferent feedback is needed to
362 trigger a given pain sensation. The findings, whilst suggesting that improved maintenance of
363 muscle activation might have contributed to the ergogenic effect of ACT ingestion, cannot
364 differentiate between a sub-conscious alteration in neuromuscular control or a conscious
365 ability to increase muscle recruitment via a reduction in pain sensation.

366

367 The increased muscle activation in the latter stages of the ACT trial compared to the placebo
368 trial was accompanied by a higher mean torque over the final 12 MVCs, and therefore a
369 higher CT (Burnley, 2009). The CT is an important physiological threshold that is linked to
370 neuromuscular fatigue development through influencing muscle metabolic homeostasis
371 (Jones et al. 2008; Vanhatalo et al. 2016), systemic respiratory and acid-base profiles (Poole
372 et al. 1988) and neuromuscular fatigue development (Burnley et al. 2012). During the 60

373 MVC protocol employed in the current study, there is a precipitous perturbation to skeletal
374 muscle homeostasis, central and peripheral fatigue development, a decline in muscle
375 activation and a hyperbolic reduction in torque that asymptotes at CT (Burnley, 2009;
376 Burnley et al. 2010). By lowering pain sensation (Foster et al. 2014; Mauger et al. 2010),
377 ACT might have permitted the attainment of a greater degree of intramuscular metabolic
378 perturbation, thereby leading to improved exercise tolerance. However, since potentiated
379 twitch was not different between the ACT and placebo trials, it appears that ACT ingestion
380 did not result in greater peripheral fatigue development in this study. Instead, ACT ingestion
381 enhanced muscle activation and increased CT over the latter stages of the 60 MVC protocol.
382 Therefore, it is also possible that ACT ingestion lowered fatigue development through
383 permitting greater muscle activation for a given degree of intramuscular metabolic
384 perturbation.

385

386 The blunted neuromuscular fatigue development in this study occurred in association with
387 increased muscle activation. However, it is surprising that the attenuation in neuromuscular
388 fatigue development was not accompanied by changes in central (i.e. voluntary activation) or
389 peripheral (i.e. potentiated twitch force) fatigue in this study. Indeed, since muscle activation
390 and torque were increased, this might be expected to result in greater intramuscular metabolic
391 perturbation and, by extension, greater peripheral fatigue development. Although we cannot
392 exclude the possibility that ACT enhanced systemic physiological responses or skeletal
393 muscle efficiency to mitigate this potential for greater peripheral fatigue development, we are
394 not aware of any published evidence to support such effects.

395

396 *Experimental Considerations*

397 It is acknowledged that a limitation of the current study was that pain sensation were not
398 directly assessed. Pain was not assessed due to the protocol requiring the completion of 60
399 MVCs with a short (2 s) recovery period; asking subjects to rate pain sensation in this setting
400 may have compromised their ability to focus on the exercise task and provide a true maximal
401 effort during the experimental protocol. Previous studies have observed a higher power
402 output for the same pain sensation, suggestive of a reduction in the sensation of pain during
403 cycle ergometry exercise (Foster et al. 2014; Mauger et al. 2010). However, these studies
404 used a higher ACT dose (1.5 g) and a different exercise modality (cycling) compared to the
405 current study. Therefore, we are unable to draw conclusions regarding pain sensation with
406 ACT in the current study. It should also be acknowledged that in the face of a relatively small
407 (~4%) improvement in torque, it is possible that the central and peripheral fatigue
408 measurements in our study were not sensitive enough to detect small defects in central
409 nervous system and peripheral muscle function. Moreover, while ACT increased EMG
410 during the latter stages of the 60 MVC protocol compared to placebo, which might be linked,
411 in part, to increased muscle activation and central motor drive, VA was not impacted by
412 ACT. This might have been a function of the continuous assessment of the EMG response
413 and torque throughout the 60 MVC protocol, compared to every 6th MVC for VA, such that
414 EMG might have provided a more complete picture of central fatigue development across the
415 protocol. Recent research has also challenged the validity of VA estimated using the
416 interpolated twitch technique as a measure of central fatigue (Neyroud et al. 2016), which
417 might contribute to the disparity between the EMG-inferred central motor drive and the VA
418 results. Accordingly, further research is required to resolve the underlying mechanisms for
419 the ergogenic effect of ACT ingestion. Although the current study is consistent with previous
420 studies reporting an ergogenic effect of acute ACT consumption (Foster et al. 2014; Mauger
421 et al. 2010, 2014), we do not advocate regular ACT use or exceeding a single dose of 1 g

422 given the potent hepatotoxicity of ACT ingestion (Graham et al. 2013). Therefore,
423 individuals wishing to explore the use of ACT to enhance exercise performance should do so
424 infrequently, and with caution.

425

426 In conclusion, acute ACT ingestion increased the mean torque across 60 MVCs of the knee-
427 extensors in agreement with earlier reports that ACT can attenuate neuromuscular fatigue
428 development and improve exercise performance. Our results extend these previous
429 observations by providing novel insights into the mechanisms for the potential ergogenic
430 effect of ACT ingestion. Specifically, the improved mean torque was accompanied by an
431 increase in CT and greater muscle activation during the latter stages of the 60 MVC protocol.
432 Therefore, ACT ingestion appears to attenuate fatigue development during repeated skeletal
433 muscle MVCs by enabling a better preservation of muscle activation during exercise.

434

435 **Conflict of interest**

436 The author declares no conflict of interest regarding the publication of this manuscript.

437 **References**

- 438 Allen, G.M., McKenzie, D.K., & Gandevia, S.C. 1998. Twitch interpolation of the elbow
439 flexor muscles at high forces. *Muscle Nerve*. 21(3):318-28.
440
- 441 Amann, M., Blain, G.M., Proctor, L.T., Sebranek, J.J., Pegelow, D.F., & Dempsey, J.A.
442 2011. Implications of group III and IV muscle afferents for high-intensity endurance exercise
443 performance in humans. *J Physiol*. 589, 5299-5309.
444
- 445 Amann, M., Proctor, L.T., Sebranek, J.J., Pegelow, D.F., & Dempsey, J.A. 2009. Opioid-
446 mediated muscle afferents inhibit central motor drive and limit peripheral muscle fatigue
447 development in humans. *J Physiol*. 587, 271-283.
448
- 449 Anderson, B.J. 2008. Paracetamol (Acetaminophen): mechanisms of action. *Paediatr*
450 *Anaesth*. 18, 915-921.
451
- 452 Black, M.I., Durant, J., Jones, A.M., & Vanhatalo, A. 2014. Critical power derived from a 3-
453 min all-out test predicts 16.1-km road time-trial performance. *Eur J Sport Sci*. 14, 217-223.
454
- 455 Blain, G. M., Mangum, T. S., Sidhu, S. K., Weavil, J. C., Hureau, T. J., Jessop, J. E., *et al.*
456 (2016). Group III/IV muscle afferents limit the intramuscular metabolic perturbation during
457 whole body exercise in humans. *J. Physiol*.
458
- 459 Burke, D. 2002. Effects of activity on axonal excitability: implications for motor control
460 studies. *Adv Exp Med Biol* 508, 33-37.
461
- 462 Burnley, M. 2009. Estimation of critical torque using intermittent isometric maximal
463 voluntary contractions of the quadriceps in humans. *J Appl Physiol*. 106, 975-983.
464
- 465 Burnley, M., & Jones, A.M. 2016. Power-duration relationship: Physiology, fatigue, and the
466 limits of human performance. *Eur J Sport Sci*. 3, 1-12.
467
- 468 Burnley, M., Vanhatalo, A., Fulford, J., & Jones, A.M. 2010. Similar metabolic perturbations
469 during all-out and constant force exhaustive exercise in humans: a ³¹P magnetic resonance
470 spectroscopy study. *Exp Physiol*. 95, 798-807.
471
- 472 Burnley, M., Vanhatalo, A., & Jones, A.M. 2012. Distinct profiles of neuromuscular fatigue
473 during muscle contractions below and above the critical torque in humans. *J Appl Physiol*.
474 113, 215-223.
475
- 476 Doyle-Baker, D., Temesi, J., Medysky, M.E., Holash, R.J., & Millet, G.Y. 2017. An
477 Innovative Ergometer to Measure Neuromuscular Fatigue Immediately after Cycling. *Med*
478 *Sci Sports Exerc*.
479

480 Enoka, R.M., & Duchateau, J. 2008. Muscle fatigue: what, why and how it influences muscle
481 function. *J Physiol.* 586, 11-23.

482

483 Forrest, J.A., Clements, J.A., & Prescott, L.F. 1982. Clinical pharmacokinetics of
484 paracetamol. *Clin Pharmacokinet.* 7(2): 93-107

485

486 Foster, J., Taylor, L., Christmas, B.C., Watkins, S.L., & Mauger, A.R. 2014. The influence of
487 acetaminophen on repeated sprint cycling performance. *Eur J Appl Physiol.* 114, 41-48.

488

489 Froyd, C., Millet, G.Y., & Noakes, T.D. 2013. The development of peripheral fatigue and
490 short-term recovery during self-paced high-intensity exercise. *J Physiol* 591(Pt 5), 1339-
491 1346.

492

493 Gandevia, S.C. 2001. Spinal and supraspinal factors in human muscle fatigue. *Physiol Rev.*
494 81, 1725-1789.

495

496 Goodall, S., Ross, E.Z., & Romer, L.M. 2010. Effect of graded hypoxia on supraspinal
497 contributions to fatigue with unilateral knee-extensor contractions. *J Appl Physiol.*
498 109(6):1842-51.

499

500 Graham, G.G., Davies, M.J., Day, R.O., Mohamudally, A., & Scott, K.F. 2013. The modern
501 pharmacology of paracetamol: therapeutic actions, mechanism of action, metabolism, toxicity
502 and recent pharmacological findings. *Inflammopharmacology.* 21, 201-232.

503

504 Hureau, T.J., Romer, L.M., & Amann M. 2016. The 'sensory tolerance limit': A hypothetical
505 construct determining exercise performance? *Eur J Sport Sci.* 7, 1-12.

506

507 Józwiak-Bebenista, M., & Nowak, J.Z. 2014. Paracetamol: mechanism of action, applications
508 and safety concern. *Acta Pol Pharm.* 71, 11-23

509

510 Jones, A.M., Wilkerson, D.P., DiMenna, F., Fulford, J., & Poole, D.C. 2008. Muscle
511 metabolic responses to exercise above and below the "critical power" assessed using 31P-
512 MRS. *Am J Physiol Regul Integr Comp Physiol.* 294, 585-593.

513

514 Jones, A.M., Grassi, B., Christensen, P.M., Krstrup, P., Bangsbo, J., & Poole, D.C. 2011.
515 Slow component of VO₂ kinetics: mechanistic bases and practical applications. *Med Sci*
516 *Sports Exerc.* 43(11), 2046-2062

517

518 Mauger, A.R., & Hopker, J.G. 2013. The effect of acetaminophen ingestion on cortico-spinal
519 excitability. *Can J Physiol Pharmacol.* 91, 187-189.

520

521 Mauger, A.R., Jones, A.M., & Williams, C.A. 2010. Influence of acetaminophen on
522 performance during time trial cycling. *J Appl Physiol.* 108, 98-104.

523

524 Mauger, A.R., Taylor, L., Harding, C., Wright, B., Foster, J., & Castle, P.C. 2014. Acute
525 acetaminophen (paracetamol) ingestion improves time to exhaustion during exercise in the
526 heat. *Exp Physiol.* 99, 164-171.
527

528 McCord, J.L., & Kaufman, M.P. 2010. Reflex Autonomic Responses Evoked by Group III
529 and IV Muscle Afferents. In: L. Kruger & A.R. Light (eds). *Translational Pain Research:
530 From Mouse to Man*, Chapter 12, pp 283-300. CRC Press/Taylor & Francis, Boca Raton, FL.
531

532 Neyroud, D., Cheng, A.J., Bourdillon, N., Kayser, B., Place, N., & Westerblad, H. 2016.
533 Muscle Fatigue Affects the Interpolated Twitch Technique When Assessed Using
534 Electrically-Induced Contractions in Human and Rat Muscles. *Front Physiol.* 7, 252.
535

536 Neyroud, D., Vallotton, A., Millet, G.Y., Kayser, B., & Place N. 2014. The effect of muscle
537 fatigue on stimulus intensity requirements for central and peripheral fatigue quantification.
538 *Eur J Appl Physiol* 114(1):205-15.
539

540 O'Connor, P.J., & Cook, D.B. 1999. Exercise and pain: the neurobiology, measurement, and
541 laboratory study of pain in relation to exercise in humans. *Exerc Sport Sci Rev.* 27:119-66.
542

543 Pageaux, B., Angius, L., Hopker, J.G., Lepers, R., & Marcora, S.M. 2015a. Central
544 alterations of neuromuscular function and feedback from group III-IV muscle afferents
545 following exhaustive high intensity one leg dynamic exercise. *American journal of
546 physiology, regulatory, integrative and comparative physiology* 308(12), ajpregu 00280
547 02014. doi: 10.1152/ajpregu.00280.2014.
548

549 Pickering, G., Estève, V., Lorient, M.A., Eschalier, A., & Dubray, C. 2008. Acetaminophen
550 reinforces descending inhibitory pain pathways. *Clin Pharmacol Ther.* 84, 47-51.
551

552 Pickering, G., Lorient, M.A., Libert, F., Eschalier, A., Beaune, P., & Dubray, C. 2006.
553 Analgesic effect of acetaminophen in humans: first evidence of a central serotonergic
554 mechanism. *Clin Pharmacol Ther.* 79, 371-378.
555

556 Place, N., Yamada, T., Bruton, J.D., & Westerblad, H. 2010. Muscle fatigue: from
557 observations in humans to underlying mechanisms studied in intact single muscle fibres. *Eur
558 J Appl Physiol.* 110, 1-15.
559

560 Pollak, K.A., Swenson, J.D., Vanhantsma, T.A., Hughen, R.W., Jo, D., White, A.T., Light,
561 K.C., Schweinhardt, P., Amann, M., & Light, A.R. 2014. Exogenously applied muscle
562 metabolites synergistically evoke sensations of muscle fatigue and pain in human subjects.
563 *Exp Physiol.* 99, 368-380.
564

565 Poole, D.C., Ward, S.A., Gardner, G.W., & Whipp, B.J. 1988. Metabolic and respiratory
566 profile of the upper limit for prolonged exercise in man. *Ergonomics.* 31, 1265-79.
567

568 Smith, H.S. 2009. Potential analgesic mechanisms of acetaminophen. *Pain Physician*. 12,
569 269-80.
570
571 Taylor, J.L., Amann, M., Duchateau, J., Meeusen, R., & Rice, C.L. 2016. Neural
572 Contributions to Muscle Fatigue: From the Brain to the Muscle and Back Again. *Med Sci*
573 *Sports Exerc*. 48, 2294-2306.
574
575 Toussaint, K., Yang, X.C., Zielinski, M.A., Reigle, K.L., Sacavage, S.D., Nagar, S., & Raffa,
576 R.B. 2010. What do we (not) know about how paracetamol (acetaminophen) works? *J Clin*
577 *Pharm Ther*. 35, 617-638.
578
579 Vanhatalo, A., Doust, J. H., & Burnley, M. 2008. A 3-min all-out cycling test is sensitive to a
580 change in critical power. *Medicine and Science in Sports and Exercise*, 40(9), 1693–1699.
581
582 Vanhatalo, A., Black, M.I., DiMenna, F.J., Blackwell, J.R., Schmidt, J.F., Thompson, C.,
583 Wylie, L.J., Mohr, M., Bangsbo, J., Krustup, P., & Jones, A.M. 2016. The mechanistic bases
584 of the power-time relationship: muscle metabolic responses and relationships to muscle fibre
585 type. *J Physiol*. 1;594(15):4407-23.

586 **Legends to figures**

587 *Figure 1*

588 Graphic overview of the procedures used in the current study. A and C: Procedures employed
589 pre and post (<10 s) high-intensity exercise, respectively. 10 s separated each single pulse
590 stimulation administered at rest (small dashed arrows). A 45 s rest period separated maximal
591 efforts (MVCs). Single pulse stimuli were administered during peak force production of
592 MVCs (large solid arrow) and immediately (1-2 s post; small grey arrows). B: 60 MVC
593 protocol of the knee extensors. The figure presents a period of 30 s which is repeated
594 sequentially for 5 min. Each MVC was held for 3 s and interspersed by a 2 s passive recovery
595 period. Every 6th MVC was accompanied by single pulse stimuli administered during peak
596 force production (Large solid arrow) and immediately following (<2 s post; small grey
597 arrows). This cycle was repeated 10 times such that the protocol spanned 5 minutes requiring
598 the completion of 60 MVCs. MVC, maximal voluntary isometric contraction. Surface
599 electromyography (EMG) was measured throughout.

600

601 *Figure 2*

602 Torque profile during the 60 maximal contractions for placebo (clear circles) and
603 acetaminophen (filled circles) trials. All contractions were normalized to a control maximal
604 voluntary contraction (MVC) performed before the test commenced. Mean \pm SE torque
605 responses are presented in panel A with the torque response from a representative individual
606 presented in panel B. Note that torque falls over the first ~150 s before reaching stable values
607 between 240 and 300 s (the end-test torque; last 12 MVCs).

608

609 *Figure 3*

610 Individual responses to ACT supplementation on 60 MVC performance (torque-impulse).

611 *Significantly different from placebo ($P<0.05$).

612

613 *Figure 4*

614 Mean \pm SE potentiated twitch (A), voluntary activation (B), and M-wave amplitude (C)

615 responses during the 60 maximal voluntary contraction (MVC) test for placebo (clear circles)

616 and acetaminophen (filled circles) trials.

617

618 *Figure 5*

619 Surface electromyography (EMG) responses (expressed relative to M-wave amplitude)

620 during the 60 MVC test for placebo (clear circles) and acetaminophen (filled circles) trials.

621 Mean \pm SE EMG responses are presented in panel A with the EMG response from a

622 representative individual presented in panel B. *Significantly different from placebo

623 ($P<0.05$).