

Title: The evolution of multiple-insecticide resistance in UK populations of tomato leafminer, *Tuta absoluta*

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Abstract

BACKGROUND: The tomato leafminer, *Tuta absoluta*, is an economically important pest of tomatoes in Europe, Africa, Asia and South America. In the UK this species is controlled using an integrated pest management (IPM) programme which incorporates the insecticides spinosad and chlorantraniliprole. In response to UK grower concerns of loss of efficacy of these compounds at certain sites, insecticide bioassays were performed on five populations collected from four commercial glasshouses and potential mechanisms of resistance investigated.

RESULTS: We observed high levels of resistance to spinosad in four of the strains, and in two of these tolerance to chlorantraniliprole. Selection of one of these strains with chlorantraniliprole rapidly resulted in a line exhibiting potent resistance to this compound. Sequencing of messenger RNA encoding the nicotinic acetylcholine receptor (nAChR) $\alpha 6$ subunit, target of spinosad, revealed *Ta $\alpha 6$* transcripts in the spinosad resistant strains that lack exon 4 and encode a highly truncated protein, or contain a triplet deletion in the predicted first transmembrane domain resulting in the loss of a highly conserved amino acid. Sequencing of the ryanodine receptor gene, encoding the target of diamide insecticides, of the chlorantraniliprole-selected line revealed an amino acid substitution (G4903V) that has been previously linked to diamide resistance in populations of *T. absoluta* in the Mediterranean and South America.

CONCLUSION: Taken together our results reveal emerging resistance in UK populations of *T. absoluta* to two of the most important insecticides used as part of IPM, with significant implications for the control of this species in the UK.

Introduction

The tomato leafminer, *Tuta absoluta*, is an economically important pest of tomatoes which spread from South America to Spain in 2006, subsequently invading a range of other European countries, Africa and the Middle East¹. *T. absoluta* arrived in the UK in 2009² and caused profound damage to commercial tomato crops at sites across the country before an Integrated Pest Management (IPM) strategy was developed that provided effective control³. This IPM strategy integrated biological control using the predatory bug, *Macrolophus pygmaeus*, with some physical control measures and three insecticides: the spinosyn spinosad, the diamide chlorantraniliprole and the oxadiazine indoxacarb. These three insecticides were selected in part due to their different modes of action, in order to prevent or delay the emergence of resistance. Recently, however, some UK growers have reported a loss of efficacy of both spinosad and chlorantraniliprole against *T. absoluta* (R. Jacobson pers. Comm.). While, to date, resistance in UK populations of *T. absoluta* to these compounds has not been confirmed, both resistance and the underlying mechanisms involved, have been characterised in populations of *T. absoluta* in other countries. In the case of spinosad, resistance has been described in field populations from Brazil with subsequent selection of one of these strains resulting in a line with extremely high levels of resistance to this compound⁴. Spinosad acts on the insect nervous system by targeting the $\alpha 6$ subunit of the nicotinic acetylcholine receptor (nAChR), a neurotransmitter-gated ion channel that plays a critical role in nerve signalling at the postsynaptic membrane. Sequencing of the gene encoding this subunit in the spinosad selected strain identified a point mutation in exon 9 of *Tac6* that results in a G275E amino acid substitution⁵. The causal role of this mutation in resistance was subsequently confirmed by CRISPR/Cas9 gene editing⁶. In a second study characterisation of the *Tac6* gene in a resistant strain selected from a field population collected in Portugal revealed an alternative mechanism of resistance conferred by skipping of exon 3 in transcripts of *Tac6*⁷.

Resistance to chlorantraniliprole has been recently described in populations of *T. absoluta* from Italy, Greece, Spain and Brazil⁸⁻¹⁰. Sequencing of the gene encoding the ryanodine receptor, a ligand-gated calcium-channel located in the sarco- and endoplasmic reticulum of neuromuscular tissues, identified several target-site mutations in the resistant strains⁸. Two of these, G4903E and I4746M, had been previously identified in diamide resistant diamondback moth^{11, 12}, while two others, G4903V and I4746T, were novel to *T. absoluta*. Radioligand binding studies and CRISPR/Cas9 gene editing provided unequivocal functional evidence that these mutations alter the affinity of the RyR to diamides and confer resistance^{8, 13, 14}.

The resistance mechanisms described above have not yet been described in UK populations of *T. absoluta*. However, the rapid invasion of this species into the UK from Europe suggests that European populations carrying resistance alleles may be readily imported into the UK – especially as *T. absoluta* is no longer a notifiable (quarantine) pest in the UK. Alternatively the use of spinosad and chlorantraniliprole in the UK for the control of *T. absoluta* might lead to the *de novo* emergence of resistance. In the current study we explored these possibilities by carrying out phenotypic and molecular characterisation of resistance in several UK populations of *T. absoluta*.

Materials and Methods

Insect strains

After reports of control failure using spinosad four strains of *T. absoluta* were collected in 2015 from commercial glasshouses at four UK sites; IOW from Wight salads, Isle of Wight, LAN from Flavourfresh Salads Ltd, Lancashire, NY from Jan Bezemer and sons, North Yorkshire and EVH from R and L Holt, near Evesham, Worcestershire. In 2016 a second strain, EVH16, was collected from R and L Holt after reports of control failure using chlorantraniliprole. An additional Spanish strain that is susceptible to spinosad and chlorantraniliprole was acquired from Rothamsted Research, Hertfordshire. Insects were

housed in controlled environment rooms at 25°C, 60% R.H., 16:8 light-dark cycle and fed *ad libitum* on tomato plants (var. Money Maker).

Leaf-dip assays

Leaf-dip bioassays were completed according to test method 22 guidelines produced by the Insecticide Resistance Action Committee¹⁵. Probit analysis and the calculation of LC₅₀ values were performed with GenStat 15th Edition (VSN International).

Chlorantraniliprole selection

Infested tomato leaves containing in excess of 500 larvae of the EVH16 strain were taken from cages and placed on a course net suspended over tomato branches that had been treated with chlorantraniliprole. This was repeated over the course of 12 months at concentrations of 1 ppm, 5 ppm, 10 ppm and 50 ppm. After each round of selection the line was left unselected for two full generations to allow the population to recover. The resulting selected line was named EVH16sel.

RNA extraction and cDNA synthesis

T. absoluta L2-L4 larvae were snap frozen in liquid nitrogen and stored at -80°C prior to RNA extraction. RNA extractions were carried out using the ISOLATE II RNA Mini Kit (Bioline) on individual larvae or pools of ten larvae. cDNA was then synthesized from 1µg of total RNA using the Maxima H Minus First Strand cDNA Synthesis kit (Thermo Scientific) according to the manufacturer's protocol.

Amplification and sequencing of the RyR and nAChR alpha six subunit

A 1353 bp region of the *T. absoluta* RyR gene (encompassing the sites of previously reported diamide resistance-conferring mutations) was amplified by PCR using DreamTaq Green PCR master mix (Thermo Fisher Scientific) and the primers detailed in Table S1. 10ng (1µl) of cDNA was used as template. Temperature cycling conditions comprised 3 min at 95°C followed by 35 cycles of 95°C for 30s, 54.5 for 30s and 72°C for 60s, and a final

extension step of 72°C for 6 min. Amplification of the nAChR $\alpha 6$ subunit gene was performed using the primers detailed in Table S1 and using a nested PCR protocol with 10ng (1 μ l) of cDNA used in the primary PCR, and 1 μ l of a 1:10 dilution of the product used in a secondary reaction. Temperature cycling conditions were as above. PCR products were purified using the GeneJET PCR purification kit (Thermo Scientific) and purified DNA quantified by spectrophotometry (NanoDrop, Thermo-Fisher) prior to Sanger sequencing using the primers detailed in Table S1.

Association of F238del with spinosad resistance

To examine the association of the F238del mutation and resistance to spinosad we utilised the LAN strain as it is heterogeneous for this mutation (frequency of 0.57). Twenty replicates of 10 larvae of this strain were placed on tomato leaves treated with 150 ppm of spinosad. As a control five replicates of 10 larvae were placed on a tomato leaf treated only with the insecticide dilutant. Mortality was scored after 72 h and all larvae were snap-frozen in liquid nitrogen and stored at -80°C. DNA was subsequently extracted from over 200 individual larvae using DNAzol following the manufacturer's protocol. A TaqMan SNP genotyping assay was designed using the Custom TaqMan Assay Design Tool (Applied Biosystems) and used to score the genotypes of dead and alive individuals generated above for the F238del mutation. Primers and MGB probes are detailed in Table S1. Each probe was labeled with a 5' reporter dye specific to either the wild-type or mutant allele. The VIC reporter dye was used to detect the wild-type allele and the 6FAM reporter dye to detect the mutant allele. Each probe also carried a 3' non-fluorescent quencher and a minor groove binder at the 3' end that provides more accurate allelic discrimination by increasing the T_M between matched and mis-matched probes. PCR reactions contained 2 μ l genomic DNA, extracted from individual insects using DNAzol reagent, 7.5 μ l of SensiFAST SYBR No-ROX kit (Bioline), 800nM of each primer and 200nM of each probe. Samples were run on CFX96 real-time PCR machine (Bio-Rad) using the temperature cycling conditions of 10 minutes at 95°C followed by 40 cycles of 95°C for 10 s and 60°C for 45 s. The increase in fluorescence

of the two probes VIC and FAM was monitored in real time. In all runs, at least one control for each genotype (mutant homozygous, heterozygous and wild-type homozygous) was included. Pearson Chi-square test was performed to examine the association between the F238del mutation and survival to spinosad exposure. To analyse the individual interactions between phenotype and genotype post-hoc analyses were conducted with adjusted residuals converted to a Chi-square score and compared to a Bonferroni adjusted α . Chi-square and post-hoc analyses were completed in SPSS (ver. 22, IBM corp.).

Results

Sensitivity of T. absoluta strains to spinosad and chlorantraniliprole

The sensitivity of the five UK strains of *T. absoluta* (EVH, EVH16, LAN, IOW and NY) to spinosad and chlorantraniliprole was compared to a reference susceptible strain (TA1) from Spain by full dose-response bioassays. In the case of spinosad significant differences in the LC_{50} values obtained were observed between four of the UK strains (EVH, EVH16, LAN and NY) and TA1 with resistance ratios ranging from 29 to > 480 (Table 1). The LAN strain exhibited moderate resistance to spinosad with an LC_{50} of 149 mg L⁻¹ while the EVH, EVH16 and NY strains showed high levels of resistance to this compound with LC_{50} values of 860 mg L⁻¹, >500 mg L⁻¹ and >2500 mg L⁻¹ respectively (Table 1). Just two of the UK strains, NY and EVH16, exhibited tolerance to chlorantraniliprole. The LC_{50} of the two strains was 8-28-fold higher than the susceptible TA1 strain. Selection of the NY strain with chlorantraniliprole failed to rapidly result in a strain with higher levels of resistance. However, progressive selection of the EVH strain with chlorantraniliprole over just 12 generations resulted in a strain with potent resistance to this compound (LC_{50} of >5000 mg L⁻¹).

Identification of altered nAChR $\alpha 6$ transcripts in spinosad resistant strains of T. absoluta

Amplification and sequencing of the nAChR $\alpha 6$ mRNA revealed the presence of two altered *Taa6* transcripts in the four resistant strains that were not observed in the susceptible strains TA1 and IOW. The first of these, observed in all *Taa6* transcripts of the NY and EVH strains

was a deletion of 109 bp that precisely corresponds to the position and size of exon 4. This deletion results in a frameshift leading to the introduction of a premature termination codon (PTC) in exon 5 (Figure 1). Amplification and sequencing of the *Taα6* gene from genomic DNA of the NY and EVH strains revealed that exon 4 was present and unaltered in both strains confirming that the loss of this exon in mRNA results from exon skipping rather than a deletion of the exon in genomic DNA.

The second alteration, observed only in the LAN strain, was an in frame triplet deletion resulting in the predicted loss of a phenylalanine amino acid at position 238 (Figure 1). The F238del mutation occurs in the first α -helical transmembrane domain (TM1) of the *Taα6* subunit and comparison of insect nAChR $\alpha 6$ sequences from a range of insect species (Figure 1) demonstrate that this residue is highly conserved across the Arthropoda. This mutation was also observed in the *Taα6* gene when amplified from genomic DNA, and at a frequency of 0.57 as determined by genotyping >100 individuals. To examine the association of the F238del mutation with spinosad resistance the LAN strain was treated with a discriminating dose of spinosad and alive, dead and affected larvae genotyped. Of the 200 individuals exposed to 150 mg/L of spinosad 108 died, 30 were affected, 55 survived the treatment and 7 escaped. To rapidly genotype these individuals, and to provide a diagnostic tool for future resistance monitoring studies, a TaqMan SNP genotyping assays was designed. This assay showed excellent discrimination of the resistant and susceptible alleles using two fluorescently labelled probes when tested with sequence verified DNA samples (Figure S1). 175 individuals from the discriminating dose assay were successfully genotyped using this assay with 31 individuals scored as homozygous for the wildtype allele, 56 as homozygous for the mutant allele and 88 heterozygous. A significant association was observed between the individual's genotype and its response to spinosad exposure ($\chi^2(4) = 78.499$, $p < 0.001$). Post-hoc analysis (Table S2) revealed that none of the 'insecticide affected' group showed a statistically significant deviation from the proportion of genotypes expected by chance. However, individuals that died in the assay had a lower than expected

proportion of the RR genotype ($p < 0.001$) and a greater than expected proportion of SS ($p < 0.001$) and RS genotypes ($p < 0.001$). In contrast, individuals surviving the assay had a higher than expected proportion of the RR genotype ($p < 0.001$) and a lower proportion of SS ($p < 0.001$) and RS ($p < 0.001$) genotypes.

A 1353 bp region of the RyR encompassing the sites of mutations previously shown to confer resistance to diamides in *T. absoluta*⁸ and *Plutella xylostella*¹² (G4903E/V and I4746M/T in *T. absoluta* corresponding to G4946E and I4790M in *P. xylostella*) was amplified from the chlorantraniliprole selected line (EVH16sel), the parent strain from which it was derived, (EVH16), NY and TA1 (Figure 3). The sequence obtained from a pooled sample of the parental EVH16 strain encoded the wildtype amino acid at all the positions detailed above, however, a very small secondary peak was observed in the sequence chromatogram at position 4903 suggesting the G4903V substitution is present at very low frequency in this strain (Figure 3). This was confirmed when the RyR sequence from the selected line was examined which showed fixation of the mutation leading to this amino acid substitution. No mutations were observed in RyR sequences of the NY and TA1 strains.

Discussion

Our data reveal high levels of spinosad resistance in strains of *T. absoluta* collected from nurseries in the north and south of the UK, indeed, only one of the UK strains, sampled from the Isle of Wight, exhibited full susceptibility to this compound. While our sample size is small this finding is consistent with reports by several British tomato growers of poor efficacy of spinosad against *T. absoluta* and suggests that resistance to this insecticide may be widespread in UK populations. In three of the strains, NY, EVH and EVH16, the levels of resistance observed were extremely high and would certainly be expected to compromise the use of spinosad at the recommended field rate (87-100 mg L⁻¹). Lower levels of resistance were observed in the LAN strain, however, the LC₅₀ value determined for this strain indicates that a significant percentage of individuals would be expected to survive exposure to the field rate application.

Sequence characterisation of transcripts encoding the nAChR $\alpha 6$ subunit revealed two novel mutations in the spinosad resistant strains. The first of these, observed in the three most resistant strains, is skipping of exon 4. This alteration has profound consequences for the encoded protein which would lack the ~36 amino acids encoded by exon 4, and would also be significantly truncated as the loss of exon 4 leads to a frameshift and the introduction of a PTC in exon 5. Together this would result in a protein of just 160 amino acids (compared to 510 amino acids in the wildtype) that lacks all four transmembrane domains and would almost certainly be non-viable. Work on *Drosophila melanogaster* first demonstrated that null mutations of $\alpha 6$ lead to spinosad resistance with a strain with a variant of *Daa6* lacking the TM3 and TM4 cytoplasmic loops, and the extracellular C-terminal tail domains exhibiting >1000-fold levels of resistance to spinosad¹⁶. Significantly, this study also demonstrated that loss-of-function mutations in insect $\alpha 6$ transcripts do not result in catastrophic loss of fitness as insects without a functional copy of the $\alpha 6$ gene are still viable¹⁶. Subsequent studies on several insect crop pests have described genetic alterations in nAChR $\alpha 6$ sequences that result in truncated non-functional proteins leading to resistance. For example, several mutations resulting in mis-splicing and the introduction of PTCs in nAChR $\alpha 6$ transcripts are associated with spinosad resistance in the diamondback moth, *Plutella xylostella*, and the oriental fruit fly *Bactrocera dorsalis*¹⁷⁻¹⁹. In a previous study of a spinosad resistant *T. absoluta* strain derived from a population collected in Portugal resistance was conferred by skipping of exon 3 rather than exon 4, however, in this case the reading frame of the altered *Taa6* transcripts was unaffected by the exclusion of exon 3⁷.

A more subtle alteration, F238del, was observed in transcripts of *Taa6* in the LAN strain and this was correlated with lower levels of resistance to spinosad. This triplet deletion results in the loss of a phenylalanine amino acid in exon 7 and occurs in TM1 of the nAChR *Taa6* subunit. This amino acid is highly conserved in nAChR $\alpha 6$ subunit proteins across Arthropoda, suggesting that this residue is functionally constrained. The loss of the amino acid at this position is thus extremely unusual and consistent with it being a *bona fide*

resistance mutation. We provide further evidence of a causal role of this mutation in resistance by demonstrating that it cosegregates with spinosad resistance in survival bioassays. Finally, the F238del mutation exhibits parallels with a previously reported amino acid substitution (G275E) shown to confer spinosad resistance in both thrips and *T. absoluta* (the latter in strains from Brazil)^{5, 6, 20, 21}. Like F238del this mutation also occurs in a α -helical transmembrane domain (TM3) of the $\alpha 6$ subunit²¹. X-ray crystallisation studies of the glutamate-gated chloride channel (GluCl), a pentameric receptor with close structural similarity to nAChRs, bound to ivermectin, another macrocyclic lactone with structural similarity to spinosad, have shown that ivermectin makes direct associations (by hydrogen bonding and van der Waals interactions) with TM1, TM2 and TM3²². This finding is consistent with spinosad acting via an allosteric transmembrane site distinct from the conventional agonist binding site²¹, and it is thus likely that the F238del mutation confers resistance by directly modifying the spinosad binding site.

It is notable that neither the G275E mutation or exon 3 skipping, previously identified in strains of *T. absoluta* from Brazil and Portugal respectively, were observed in UK strains of *T. absoluta* suggesting that the mutations identified in our study arose *de novo* in UK populations rather than being imported via transfer of insects on plant material or packaging from outside the UK. The application of spinosad via the irrigation system in the UK, together with the redundancy of the nicotinic acetylcholine receptor (nAChR) $\alpha 6$ subunit, in which any nonfunctionalizing mutation will confer resistance, likely facilitated the rapid evolution of resistance.

Two of the strains tested in our study also showed tolerance to chlorantraniliprole - while the levels of tolerance displayed were relatively modest selection of one of these strains rapidly resulted in a line with potent resistance to this compound. This finding suggests that resistance alleles were almost certainly present in the population prior to selection. Sequencing of a region of the RyR encompassing the sites of known resistance mutations confirmed this and revealed that a mutation leading to a G4903V substitution present in the

parental strain at low frequency had reached fixation in the selected line. This mutation has recently been reported in *T. absoluta* strains from Greece and Italy⁸ suggesting *T. absoluta* with this resistance allele may have been imported into the UK. The functional significance of the G4903V substitution has been confirmed *in vitro* using radioligand binding studies and *in vivo* by CRISPR/CAS9 genome editing in *Drosophila*¹³. In the case of the latter genome modified flies bearing the mutation exhibited high levels of resistance to both the diamides flubendiamide (91-fold) and chlorantraniliprole (195-fold) when compared to flies of the same genetic background without the mutation¹³.

Our results have significant applied implications in relation to the control of *T. absoluta* in the UK. Spinosad performs a key role in the IPM strategy introduced against this pest in 2013, and is applied at the start of the season to provide control while the natural enemy *M. pygmaeus* is becoming established³. If necessary, a high volume spray of chlorantraniliprole is applied as a second line of defence later in the season to keep the pest and predator populations in balance³. The fact that we identified one strain with resistance mechanisms to both spinosad and chlorantraniliprole is concerning as it demonstrates that *T. absoluta* with resistance to both compounds can readily evolve and are viable. It would be useful to screen UK populations of *T. absoluta* more widely for the resistance mechanisms identified in this study. If such monitoring indicates that, as suggested by our study, spinosad resistance is widespread then we suggest that use of this insecticide is suspended for a period to examine if susceptibility is restored to populations in the absence of selection. The likelihood of this is dependent on any fitness costs associated with the resistance mechanisms we have identified. In *D. melanogaster* no evidence of fitness costs were identified under laboratory conditions in a Dα6 null mutant, suggesting the costs of loss of function mutations in this subunit may be minimal¹⁶. However, work on a spinosad selected strain of *T. absoluta* found that resistance was unstable in the absence of spinosad selection and dropped rapidly over relatively few (<10) generations²³. In our study we observed high levels of spinosad resistance in a strain (EVH16) collected from a nursery where this compound had not been

used for a year suggesting the exon 4 skipping identified in this strain may have a low fitness penalty. However, additional work is required to clarify fitness costs associated with this mutation and the others identified in this study in both the $\alpha 6$ nAChR subunit and the RyR under field-realistic conditions.

Mutations conferring resistance to chlorantraniliprole were only identified in a single strain in our study suggesting that with effective resistance management it may be possible to prevent the further development of resistance to this compound. This would require the rotation of this insecticide with insecticides of alternative modes of action and/or the use of alternative control methods. In regards to the latter recent trials in the UK using a synthetic sex pheromone as a mating disruption strategy against *T. absoluta* have provided encouraging results²⁴ and may allow the frequency of insecticide applications to be significantly reduced. However, our study clearly illustrates the danger of complacency and we suggest that the control of *T. absoluta* in the UK (and worldwide) should continue to rely on the use of a range of integrated approaches in order to reduce selection pressure on any single control in order to prevent or slow the emergence of resistance.

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Table 1

Sensitivity of UK strains of *Tuta absoluta* to the insecticides spinosad and chlorantranilprole. The resistance ratio (RR) of these strains was compared to a susceptible strain from Spain (TA1). CI: confidence interval.

Treatment	Strain	Location	LC ₅₀ (mg / L)	95% CI	RR
Spinosad	TA1	Spain	5.2	3.1 – 7.3	1
	NY	North Yorkshire	>2500		>480.77
	EVH	Evesham	860	484 - 2114	165.38
	EVH16	Evesham	>500		>96.15
	LAN	Lancashire	149	31 - 5385	28.65
	IOW	Isle of Wight	8.6	5.3 - 13.9	1.65
Chlorantranilprole	TA1	Spain	1.2	0.095 – 5.00	1
	NY	North Yorkshire	33	11.2 - 97.9	27.5
	EVH	Evesham	0.12	0.005 - 1.00	0.1
	EVH16	Evesham	9.53	5.025 - 18.09	7.94
	EVH16sel	Evesham	>5000		>4166.67
	LAN	Lancashire	4.75	1.7 - 12.8	3.96
	IOW	Isle of Wight	3.6	0.9 - 14	3

Figure legends

Figure 1. Schematic of mutations identified in *Taa6* transcripts of spinosad resistant *T. absoluta* strains. (A) Schematic of the exon structure of *Taa6*. (B-D) Alignment of the sequences obtained from six *T. absoluta* strains illustrating the skipping of exon 4 observed in three of the spinosad resistant strains that leads to the introduction of a premature termination codon in exon 5. (E-F) The LAN strain has a triplet deletion in the encoded first transmembrane domain of *Taa6* that leads to an in frame deletion of an amino acid that is highly conserved in nAChR $\alpha 6$ subunit sequences across Arthropoda.

Figure 2. Genotype of *T. absoluta* larvae for the F238del mutation that died, survived or were affected following exposure to a discriminating dose of spinosad. n Dead = 108, n affected = 30, n survived = 55.

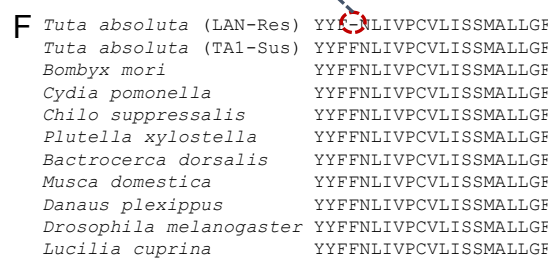
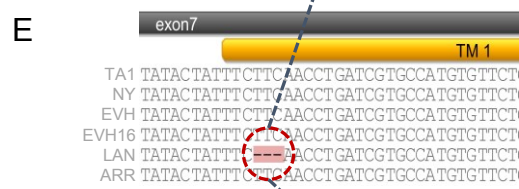
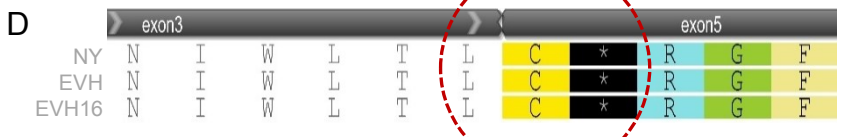
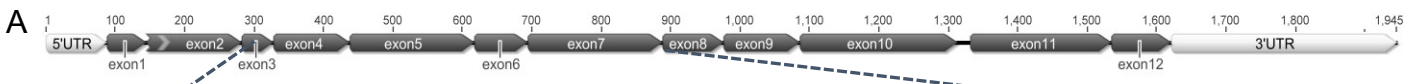
Figure 3. Identification of the G4903V amino acid substitution in a diamide resistant strain of *T. absoluta*. (A) Alignment of RyR amino acid sequences obtained from three UK strains of *T. absoluta* and a strain from Spain. (B) Sequence chromatograms of the same strains at the G4903 codon illustrating a slight secondary 'T' peak in the second nucleotide of the codon EVH16 strain that has become fixed following chlorantraniliprole selection (EVH16sel).

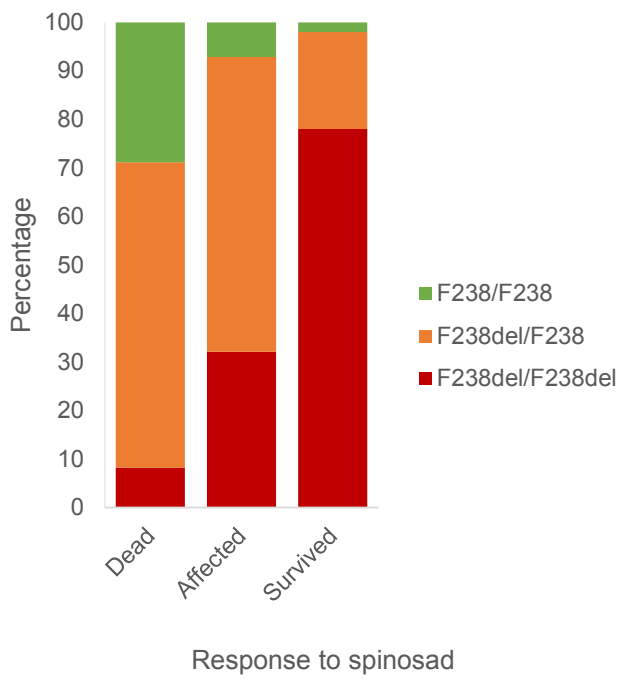
Supporting Information

Table S1. Oligonucleotide primers and probes used in this study.

Table S2. Results of Chi-square post-hoc analysis of genotype at F238 and response to spinosad exposure.

Figure S1. Real-time TaqMan detection of the F238del mutation. Fluorescence of the MGB probe labelled with VIC (specific for wild-type allele) is shown by brown trace, and the fluorescence of the probe labelled with 6FAM is shown by blue trace. A) Fluorescent signal derived from the DNA of a homozygous F238 (wild type) individual. B) Fluorescent signal derived from the DNA of a heterozygous individual. C) Fluorescent signal derived from the DNA of a homozygous F238del (mutant) individual.





A

TA1	N	I	A	A	A	L	H	S	I	V	S	L	L	I	I	I	4,750	Y	Y	H	L	K	V	L	A	I	4,760	F	F	F	F
NY	N	I	A	A	A	L	H	S	I	V	S	L	L	I	I	I	4,750	Y	Y	H	L	K	V	L	A	I	4,760	F	F	F	F
EVH16	N	I	A	A	A	L	H	S	I	V	S	L	L	I	I	I	4,750	Y	Y	H	L	K	V	L	A	I	4,760	F	F	F	F
EVH16-sel	N	I	A	A	A	L	H	S	I	V	S	L	L	I	I	I	4,750	Y	Y	H	L	K	V	L	A	I	4,760	F	F	F	F
TA1	K	R	E	K	E	I	A	R	K	L	E	F	D	G	L	Y	4,770	E	A	4,780	E	D	D	D	L	K	S	S	S	S	
NY	K	R	E	K	E	I	A	R	K	L	E	F	D	G	L	Y	4,770	E	A	4,780	E	D	D	D	L	K	S	S	S	S	
EVH16	K	R	E	K	E	I	A	R	K	L	E	F	D	G	L	Y	4,770	E	A	4,780	E	D	D	D	L	K	S	S	S	S	
EVH16-sel	K	R	E	K	E	I	A	R	K	L	E	F	D	G	L	Y	4,770	E	A	4,780	E	D	D	D	L	K	S	S	S	S	
TA1	H	W	D	K	L	L	V	I	S	A	K	S	P	V	N	Y	4,800	W	D	4,810	V	K	K	V	V	R	A	K	K	K	
NY	H	W	D	K	L	L	V	I	S	A	K	S	P	V	N	Y	4,800	W	D	4,810	V	K	K	V	V	R	A	K	K	K	
EVH16	H	W	D	K	L	L	V	I	S	A	K	S	P	V	N	Y	4,800	W	D	4,810	V	K	K	V	V	R	A	K	K	K	
EVH16-sel	H	W	D	K	L	L	V	I	S	A	K	S	P	V	N	Y	4,800	W	D	4,810	V	K	K	V	V	R	A	K	K	K	
TA1	Y	S	E	T	Y	D	D	S	I	S	N	M	L	G	M	E	4,830	E	K	4,840	A	A	A	O	E	D	E	G	G	G	
NY	Y	S	E	T	Y	D	D	S	I	S	N	M	L	G	M	E	4,830	E	K	4,840	A	A	A	O	E	D	E	G	G	G	
EVH16	Y	S	E	T	Y	D	D	S	I	S	N	M	L	G	M	E	4,830	E	K	4,840	A	A	A	O	E	D	E	G	G	G	
EVH16-sel	Y	S	E	T	Y	D	D	S	I	S	N	M	L	G	M	E	4,830	E	K	4,840	A	A	A	O	E	D	E	G	G	G	
TA1	S	K	G	L	K	Y	I	I	T	I	D	W	R	R	Y	O	4,860	V	W	4,870	A	G	V	T	F	T	N	S	S	S	
NY	S	K	G	L	K	Y	I	I	T	I	D	W	R	R	Y	O	4,860	V	W	4,870	A	G	V	T	F	T	N	S	S	S	
EVH16	S	K	G	L	K	Y	I	I	T	I	D	W	R	R	Y	O	4,860	V	W	4,870	A	G	V	T	F	T	N	S	S	S	
EVH16-sel	S	K	G	L	K	Y	I	I	T	I	D	W	R	R	Y	O	4,860	V	W	4,870	A	G	V	T	F	T	N	S	S	S	
TA1	F	L	Y	S	L	W	Y	F	F	S	V	M	G	N	F	4,890	F	F	4,900	A	A	A	H	L	L	L	D	V	V	V	
NY	F	L	Y	S	L	W	Y	F	F	S	V	M	G	N	F	4,890	F	F	4,900	A	A	A	H	L	L	L	D	V	V	V	
EVH16	F	L	Y	S	L	W	Y	F	F	S	V	M	G	N	F	4,890	F	F	4,900	A	A	A	H	L	L	L	D	V	V	V	
EVH16-sel	F	L	Y	S	L	W	Y	F	F	S	V	M	G	N	F	4,890	F	F	4,900	A	A	A	H	L	L	L	D	V	V	V	
TA1	A	V	G	F	K	L	R	R	T	T	I	L	O	V	H	N	4,920	C	L	4,930	L	V	L	T	V	M	L	L	L	L	
NY	A	V	G	F	K	L	R	R	T	T	I	L	O	V	H	N	4,920	C	L	4,930	L	V	L	T	V	M	L	L	L	L	
EVH16	A	V	G	F	K	L	R	R	T	T	I	L	O	V	H	N	4,920	C	L	4,930	L	V	L	T	V	M	L	L	L	L	
EVH16-sel	A	V	G	F	K	L	R	R	T	T	I	L	O	V	H	N	4,920	C	L	4,930	L	V	L	T	V	M	L	L	L	L	
TA1	T	I	I	V	Y	I	Y	T	T	V	I	A	N	N	F	4,940	V	Y	4,950	E	E	D	D	D	E	V	N	N	N	N	
NY	T	I	I	V	Y	I	Y	T	T	V	I	A	N	N	F	4,940	V	Y	4,950	E	E	D	D	D	E	V	N	N	N	N	
EVH16	T	I	I	V	Y	I	Y	T	T	V	I	A	N	N	F	4,940	V	Y	4,950	E	E	D	D	D	E	V	N	N	N	N	
EVH16-sel	T	I	I	V	Y	I	Y	T	T	V	I	A	N	N	F	4,940	V	Y	4,950	E	E	D	D	D	E	V	N	N	N	N	

B

